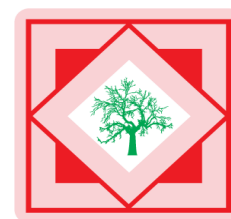




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RP-HPLC method for the determination of Tenatoprazole in pharmaceutical formulations

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ABSTRACT

A reverse phase high performance liquid chromatography (RP-HPLC) has been developed for the estimation of Tenatoprazole in bulk drug and pharmaceutical dosage form. The quantification was carried out using C_{18} column and mobile phase consisting of 10mM phosphate buffer at p^H 2.4: acetonitrile (60:40 v/v), at flow rate of 1 mL/min. The separation was performed at ambient temperature. Eluents were monitored by UV detector set at 307 nm. The method was statistically validated for the linearity, precision, accuracy, LOD and LOQ. The linearity was found to be in the range of 1-6 $\mu\text{g/mL}$. The proposed method was found to be simple, precise, accurate, rapid and reproducible for the estimation of Tenatoprazole in bulk drug and tablets.

Key words: Tenatoprazole , RP-HPLC and UV-detector.

INTRODUCTION

Tenatoprazole (TPZ) is chemically, 3-methoxy-8- [(4-methoxy-3,5-dimethyl-pyridin-2 yl) methyl sulfinyl] 2,7,9-triazabicyclo [4.3.0] nona-2,4,8,10-tetraene. (Figure 1).It is a prodrug of the proton pump inhibitor (PPI) class, which is converted to the active sulfenamide or sulfenic acid by acid in the secretory canaliculus of the stimulated parietal cell of the stomach. This active species binds to lumenally accessible cysteine of the gastric $H^+ K^+ \text{-ATP}$ ase resulting in disulfide formation and acid secretion inhibition [1]. Literature review revealed that only HPLC methods are reported for the estimation of TPZ in rat and dog plasma and stability-indicating thin-layer chromatographic and chiral separation LC method for pharmaceutical dosage forms [2-5]. The objective of this study was to develop a simple, rapid, economic and sensitive RP-HPLC method for the analysis of TPZ in its tablet formulation using the most commonly employed RP- C_{18} column with UV-detection.

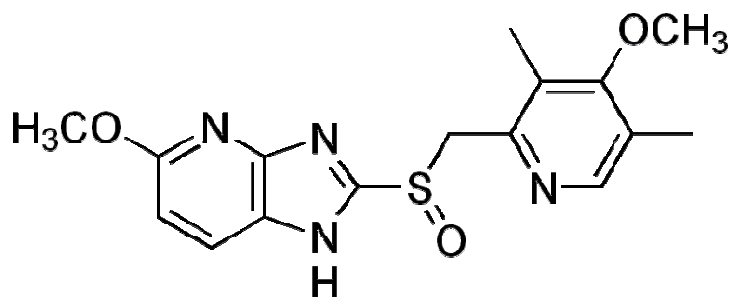


Figure 1 structural formula of tenatoprazole

MATERIALS AND METHODS

The pure drug sample of TPZ was obtained as gift sample from Reddy's Laboratory, Hyderabad. Allegro (enteric coated tablets) containing 40 mg of TPZ was purchased from SIDEM Pharmacy, U.K. Qualigens fine chemicals, Mumbai, supplied HPLC grade acetonitrile and water, sodium dihydrogen orthophosphate AR grade and Phosphoric acid AR grade. An isocratic high pressure liquid chromatograph (Schimadzu HPLC class VP series) with LC-10 ATVP pump, variable wavelength programmable UV/Visible detector SPD-10 AVP system and operating software winchrome was used. The chromatography column used was a reverse phase phenomenax C₁₈ column (250mm × 4.6 mm i.d, particle size 5μ). A mixture of 10mM phosphate buffer and acetonitrile (adjusted to pH 2.4 using ortho phosphoric acid) in the ratio of 60:40 v/v was used as mobile phase and was filtered before use through 0.45μ membrane filter. The flow rate of mobile phase was maintained at 1.0 ml/ min. Detection was carried out at 307nm at ambient temperature.

Preparation of 10 mM phosphate buffer pH 2.4

1.3609 g of potassium dihydrogen orthophosphate was dissolved in sufficient quantity of distilled water and made to produce 1000 mL and adjusted to pH 2.4 with ortho phosphoric acid [6].

Preparation of standard solution

25 mg of TPZ was dissolved in a minimum quantity of acetonitrile and the total volume was brought to 50 mL with acetonitrile, further dilutions were made with acetonitrile to get 10 μg/mL.

Preparation of calibration curve

Aliquots of standard solution of TPZ 1 to 6 mL was transferred into 10 mL volumetric flask and made up to the mark to get the concentration range from 1 to 6 μg/mL and the calibration curve was plotted between concentration and peak area.

Quantification of Tenatoprazole in formulation

Twenty tablets containing 40mg of TPZ were accurately weighed and finely powdered. The powdered tablet equivalent to 25 mg of TPZ was weighed and transferred into a 50 ml volumetric flask, added sufficient quantity of acetonitrile and was sonicated for few minutes and made up to the mark with acetonitrile. The solution was filtered through Whatmann filter paper No.41. From this clear solution, further dilution was made with acetonitrile to produce 10

detection (LOD) and limit of quantification (LOQ) were separately determined based on the standard deviation of response of the calibration curve. The standard deviation of y- intercepts of regression lines and slope of the calibration curves were used to calculate LOD and LOQ. The LOD and LOQ values are 0.2515 and 0.7623 $\mu\text{g/mL}$ respectively.

Table 1 Quantification of TPZ formulation

Labeled claim (mg/tab)	Amount found(mg)	Percentage purity obtained	Mean \pm S.D	% RSD
40	40.08	100.20	100.35 \pm 1.6431	1.6046
40	40.20	100.50		
40	40.08	100.20		
40	40.20	100.50		
40	40.08	100.20		
40	40.20	100.50		

RSD: Relative standard deviation

The proposed method was validated as per the standard analytical procedures [8]. Each sample was injected six times and the retention times were same. Accuracy of the method was calculated by recovery studies (n=3) at three levels. Standard drug solution containing drug in the range 100.0, 66.66 and 33.33% of nominal concentration was added to previously analyzed test solution. Amount of drug recovered at each level was calculated. The sample recovery in the formulation (Table 2) was in good agreement with the label claim.

Table 2 Recovery studies of TPZ formulation

Percentage	Amount present ($\mu\text{g/ml}$)	Amount Added ($\mu\text{g/ml}$)	Total Estimated* ($\mu\text{g/ml}$)	Amount recovered * ($\mu\text{g/ml}$)	% recovery	Mean \pm S.D	% RSD
100.00	3.15	3	6.11	2.96	98.60	99.20 \pm 0.7211	0.7269
66.66	3.12	2	5.12	2.00	100.00		
33.33	3.06	1	4.05	0.99	99.00		

** Mean of three observations*

Table 3 System suitability parameters of TPZ

Parameter	Value
Retention Time	3.45
Tailing factor	1.08
Capacity factor	1.54
Asymmetric factor	1.08
No of theoretical plates	5273

High percentage recovery and low % RSD showed that the method was free from interference of the commonly used excipients in the formulation. System suitability parameters of the proposed method for TPZ are given in Table 3. No interfering peaks were found in the chromatogram indicating that excipients used in the tablet formulation did not interfere with the estimation of the drug by the proposed RP-HPLC method.

CONCLUSION

The procedure described here is simple, rapid, sensitive, selective and cost effective. It is evident from the results that the recommended procedure is well suited for assay and evaluation of drug, in dosage forms. It can be applied for direct determination of TPZ in drug control laboratories.

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