

Antimicrobial Effect of Pyocyanin Extracted from *Pseudomonas aeruginosa*

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Received date: Oct 21, 2016; Accepted date: Dec 06, 2016; Published date: Dec 10, 2016

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Citation: Abdul-Hussein ZR, Atia SS. Antimicrobial Effect of Pyocyanin Extracted from *Pseudomonas aeruginosa*. Eur J Exp Bio. 2016, 6:6.

Abstract

Pyocyanin was extracted from environmental *Pseudomonas aeruginosa* isolated from soil at Basra city, Bacteria was identified by using Vitek-2 system analysis. Pigment was purified after extraction and identified by using thin layer chromatography, spectrophotometric, and GC mass analysis. Antibacterial and antifungal activities of different concentrations of pigment were studied against urinary tract and wound infection bacteria and pathogenic fungi. Results showed that pigment gave a good effect against all bacteria and fungi.

Keywords: Pyocyanin; *Pseudomonas aeruginosa*; Antimicrobial

Introduction

Pseudomonas aeruginosa is gram-negative rod shaped, asporogenous, and monoflagellated bacterium. It is about 1-5 µm long and 0.5-1.0 µm wide. *P. aeruginosa* is obligate bacteria that respire aerobically as its optimal metabolism. However it can also respire anaerobically on nitrate or other alternative electron acceptors. Therefore this makes *P. aeruginosa* very invasive microorganisms and has been found in environments such as soil, water, humans, animals, plants, sewage, and hospitals [1]. One of the applications of *P. aeruginosa* in biotechnology is its ability to degrade aromatic hydrocarbon such as methylbenzenes, which are the by-products of petroleum industries and used as solvents for enamels and paints as well as in production of drugs and chemicals. Methylbenzenes are considered as environmental contaminants that ubiquities in atmosphere, underground and soil, and in surface water [2]. *P. aeruginosa* can also degrade toluene, the simplest form of methylbenzene by the oxidation of methyl group to aldehyde, alcohol, acid, and convert it to catechol. Therefore it can be used in pollution control [3]. Phenazines are redox-action pigments produced by these bacteria. These pigments are involved in virulence and iron acquisition [4]. Pyocyanin is a water-soluble blue green, phenazine pigment produced by active cultures of *pseudomonas aeruginosa*. Pyocyanin also has antibiotic activity toward different microorganisms [5,6].

Materials and Methods

Soil samples were collected from different region of Basra city and were serially diluted in sterile distilled water. Processed samples were cultured on nutrient agar. The blue green colonies grown on nutrient agar were sub cultured onto nutrient agar and Gram stained. Cultured plate were incubated for 24-48 h at 25-37°C. Bacteria was identified biochemically with catalase, oxidase and then subjected to Vitek 2 system analysis.

Nutrient broth, mineral salt medium, and peptone water medium was used for the production of pyocyanin pigment. *Pseudomonas aeruginosa* was inoculated into one of the medium following up and grown for 72 h at 37°C. Bacteria were then removed by centrifugation (10,000xg 30 min) and filtration of the supernatant through 0.45 µm filters [7]. After that, 5 ml *Pseudomonas* culture was taken in a sterilized tube and same volume of chloroform was added. The solution was mixed well by using shaker for 2 min, and again centrifugation at 10,000xg for 15 min. Two distinct layers separated out, in which one was the pigment, the other remaining material of culture. The pyocyanin was removed and stored in screw capped tube at 4°C. Pigment was passed through TLC plates using chloroform-methanol in the ratio (9:1 v/v) as an eluent [8]. For spectrophotometric analysis, pigment was extracted with chloroform followed by centrifugation at 4000-70,000 rpm for 10-15 minutes and the cell debris was removed by a second centrifugation step. Finally the supernatant was transferred to a cuvette for the measurement of absorbance at different scanning range.

Gas chromatography(GC-Mass) analysis of pyocyanin were performed on a shimadzu Qp2010 quadrupole gas chromatography Mass spectrophotometer (GC-MS) instrument equipped with a carbowax (30 mm × 0.25 mm ID; 0.25 µm film thickness) capillary column (intercut DB5Ms. Japan). 1 µl of the sample was injected into the capillary column. Helium was used as the carrier gas. Injector and detector temperatures were set at 280°C. Injection was performed in split mode (1:30), the column temperature was programed initially at 40°C for 1 min and then increase at a rate of 5° per min at final temperature of 280°C. Pigments were separated at constant pressure (96.1 Kpa) 6 split ratio 30.0, column flow 1.71 ml/min and peaks were identified by comparing the mass spectra with mass spectral database.

Antimicrobial activity

Effect of pigment against pathogenic bacteria from wound (*Pseudomonas aeruginosa*, *Staphylococcus*, *E. coli*) and urinary tract infection (*Pseudomonas aeruginosa*, *Staphylococcus*, *E. coli* and *Bacillus*) and pathogenic fungi *Cryptococcus neoformans*, *Candida albicans* fungi *Aspergillus niger* and *Aspergillus fumigatus* were performed by using agar diffusion methods. Data were summarized as mean \pm SD. All data were subjected to statistical analysis using one way analysis of variance (ANOVA) by using mini tab program. The differences were considered significant if $P < 0.05$.

Results and Discussion

Blue green pigment producing colonies were picked up from cultures grown on nutrient agar and subcultured on nutrient agar and gave name (ps), purified colonies were Gram stained, and catalase and oxidase tests examined, in addition to some biochemical tests (Figure 1).

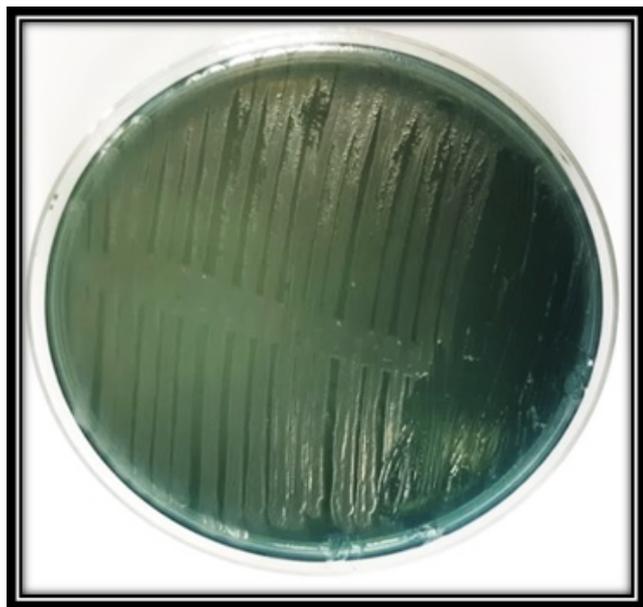


Figure 1 Blue green pigment of *Pseudomonas aeruginosa*.

Pseudomonas suspected was further diagnosed by subjecting to Vitek system analysis. Results of vitek2 system analysis revealed that bacterial isolates were identified as ps=*Pseudomonas aeruginosa* (Figure 2).

Enhancement of pyocyanin production

Different media were used for enhancement of pigment production for *Pseudomonas* (green). Mineral salt medium and peptone water were much better support medium for producing of blue-green pigment, while mineral mannitol, including broth and broth malt with cooked meat extract showed less improvement at same temperature and period of incubation.

Identification Information		Card:	GN	Lot Number:	241334140	Expires:	Feb 6, 2016 12:00 CST										
Completed:		May 14, 2015 02:59 CDT	Status:		Final	Analysis Time: 6.00 hours											
Selected Organism		99% Probability <i>Pseudomonas aeruginosa</i>		Biometer: 0003453103500240		Confidence: Excellent identification											
SRF Organism																	
Analysis Organisms and Tests to Separate:																	
Analysis Messages:																	
Contraindicating Typical Biopattern(s)																	
Biochemical Details																	
2	APPA	-	3	ADO	-	4	PyrA	-	5	IARL	-	7	dCEL	-	9	BGAL	-
10	H2S	-	11	BNAG	-	12	AGLTp	-	13	dGLU	+	14	GGT	+	15	OFF	-
17	BGLU	-	18	dMAL	-	19	dMAN	+	20	dMNE	+	21	BXYL	-	22	BAlap	+
23	ProA	+	26	LIP	+	27	PLE	-	29	TyrA	+	31	URE	-	32	dSOR	-
33	SAC	-	34	dTAG	-	35	dTRE	-	36	CIT	+	37	MNT	+	39	SKG	-
40	ILATk	+	41	AGLU	-	42	SUCT	+	43	NAGA	-	44	AGAL	-	45	PHOS	-
46	GlyA	-	47	ODC	-	48	LDC	-	53	IHISa	-	56	CMT	+	57	BGRU	-
58	O129R	-	59	GGAA	-	61	IMLTa	+	62	ELLM	-	64	ILATa	-			

Figure 2 Vitek analysis sheet of *Pseudomonas aeruginosa*.

The other bacterial pigment was cultured on nutrient agar (NA) and peptone glycerol casein (PGC) gave a good recorded result.

Vitek analysis *Pseudomonas aeruginosa*

Results of current study showed that isolated pigmented bacteria with greenish color ps exhibited phenomenal probability 99% and confidence excellent identification PS identified as *Pseudomonas aeruginosa*, this results harmonious with previous studies of who identified different species of bacteria including *A. xylosoxidans*, *A. baumani*, *Pseudomonas aeruginosa* and *P. fluorescens* by using vitek. Marko and his colleagues revealed that it can identify different species bacteria isolated from Cf patients and accredit on characteristic features of non-fermenting, Gram negative, Bacilli [9,10].

Extraction and chemical analysis of pigment

Chloroform extracted pyocyanin showed bluish extract converted to red when acidified with 0.1 (N) HCl (Figure 3).

GC-MS Chromatograph of *Pseudomonas aeruginosa* chloroform extracted shown a sharp peak at acquisition time 23.17 minutes on gas Chromatographic analysis which identified by mass spectrum analysis as alpha-Hydroxy phenazine (Hemipyocyanin) that gave intense molecular ion peak at 196 m/z with its structure shown in Figure 4.

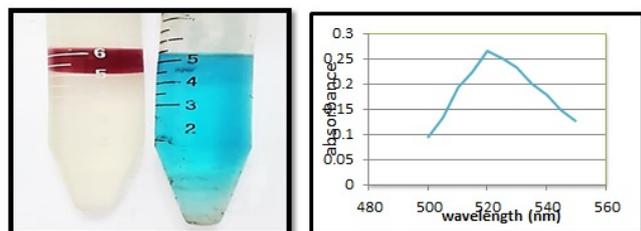


Figure 3 Bluish extract of pyocyanin converted color to red with HCl (left), and spectrophotometric analysis of pyocyanin showed a peak at 520 nm.

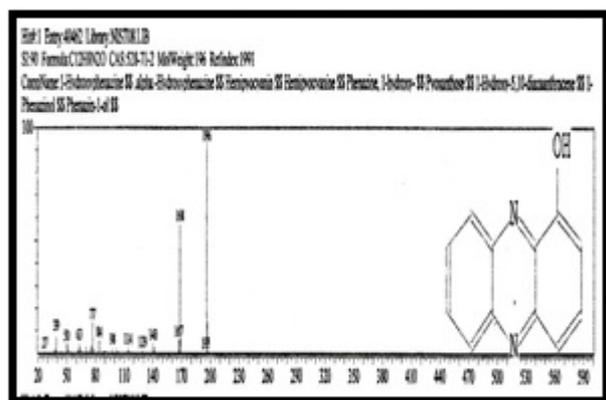


Figure 4 GC-mass spectral analysis of pyocyanin extracted from *Pseudomonas aeruginosa* revealed the chemical structure of pigment which cauterized as hemi pyocyanine.

TLC

The chromatogram of colored spot on TLC plate showed an RF value of 0.83 for pyocyanin. The result was comparable with previous studies [8]. Baron and John obtained RF value of pyocyanin (0.71) by eluting with chloroform solvent [5]. In the present study pyocyanin was separated from culture supernatants by the addition of chloroform solvent. Extractable chloroform layer was mixed with 1 ml of 0.2 (N) HCl which converted pyocyanin to pinkish red color which indicated the presence of pyocyanin pigment. The result was consonant with the previous studies [11,12]. Partially purified of pyocyanin was spectrophotometrically scanned at range of 200-800 nm. Absorption was measured at 520 nm, as was also observed by other authors [12,13].

Gas chromatography-Mass of pyocyanin in the present study, showed the presence of Hemipyocyanin and phenazine compound. Previous GC-Ms analysis confirmed the result of the study of who showed the related hemi-pyocyanin pigment extracted from *Pseudomonas aeruginosa* and identified by its electron impact mass spectrum after gas chromatography at ions at m/z 196 [14].

Antimicrobial effect of pyocyanin

The most affected bacteria to pyocyanin were *E. coli* extracted from urinary tract, followed by *Bacillus* and *Staphylococcus* at the same level. The effect of pyocyanin on wound bacteria was most obvious on *Pseudomonas* and *E. coli*, followed by *Staphylococcus* respectively in the concentration range of 10.000 to 100 $\mu\text{g/ml}$ ($P < 0.008$) (Figure 5).

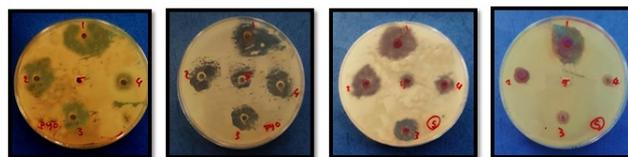


Figure 5 Effect of different concentration (1=10000, 2=5000, 3=1000, 4=500, 5=100) of pyocyanin on pathogenic bacteria (from left to right) *Pseudomonas*, *E. coli*, *Staphylococcus* and *Bacillus* respectively.

The most affected mould to pyocyanin pigment *Aspergillus niger* and *Aspergillus fumigatus* respectively, followed by yeast *Cryptococcus neoformans*, while *Candida tropicalis* and *C. albicans* were equally effected. *C. krusi* was less affected yeast to (pyocyanin) pigment in the concentration range of 10.000 to 100 $\mu\text{g/ml}$ ($P > 0.000$) (Figure 6).

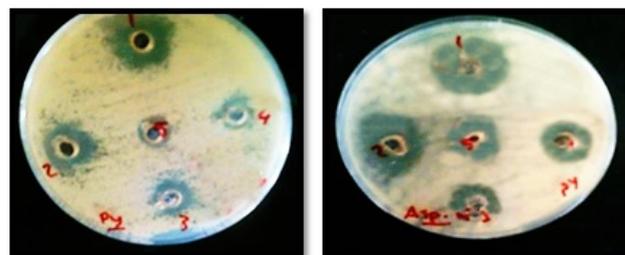


Figure 6 Effect of different concentration of pyocyanin (10000, 5000, 1000, 500, and 100 $\mu\text{g/ml}$ against *Candida albicans* and *Aspergillus niger* (from left to right).

Activity of phenazin antibiotics are comparable with other study who revealed phenazin antibiotic have a marvellous effects on Gm+ve and Gm-ve, these results are in agreement with that previous result who reported that phenazin antibiotic have antimicrobial activity only against strains of *Bacillus subtilis*, *Escherichia Coli* and *Candida albicans* [15]. Pyocyanin has been detected in an oil-degrading culture containing *Pseudomonas aeruginosa* and is a redox-active compound capable of inhibiting the growth of pyocyanin-sensitive of the microbial community [16]. The antagonistic effects of all of phenazine derivatives are ascribed to on general characteristic redox-activity [17].

Conclusion

Present study concluded that pyocyanin extracted from environmental isolate of *Pseudomonas aeruginosa* was

hemipyocyanin and has antimicrobial effect against infectious bacteria and some pathogenic fungi.

References

1. Leaderbery J (2000) *Pseudomonas*. Encyclopedia of Microbiology 3: 876-891.
2. Pieper D, Stadler-fritzche K, Schoolman M, Knachmuss H (1992) Metabolism of 2-Chloro-4-Methylphenoxyacetate by *alcaligenes eutrophus* JMP 134: Implications for the degradation of chloro- and methyl-substituted aromatics via ortho cleavage. FEMS Symposium. pp: 268-272.
3. Johnson GR, Olsen RH (1997) Multiple pathways for toluene degradation in *Burkholderia* sp. strain JS150. Appl Environ Microbiol 63: 4047-4052.
4. Dietrich LE, PriceWhelan A, Petersen A, Whiteley M, Newman DK (2006) The phenazine pyocyanin is a terminal signalling factor in the quorum sensing network of *Pseudomonas aeruginosa*. Mol Microbiol 61: 1308-1321.
5. Baron SS, Rowe JJ (1981) Antibiotic action of pyocyanin. Antimicrob Agents Chemother 20: 814-820.
6. Liang H, Li L, Dong Z, Surette MG, Duan K (2008) The YebC family protein PA0964 negatively regulates the *Pseudomonas aeruginosa* quinolone signal system and pyocyanin production. J Bacteriol 190: 6217-6227.
7. Hassett DJ, Charniga L, Bean K, Ohman DE, Cohen MS (1992) Response of *Pseudomonas aeruginosa* to pyocyanin: mechanisms of resistance, antioxidant defenses, and demonstration of a manganese-cofactored superoxide dismutase. Infect Immun 60: 328-336.
8. Genevive G, Sharief B, Sebastein R, Tjeerd V, Ben J, et al. (2006) Pip, a novel activator of phenazine biosynthesis in *pseudomonas chlororaphis* PCL1391. J Bacteriol 188: 8283-8293.
9. Martiny D, Busson L, Wybo I, Haj RAE, Dediste A, et al. (2012) Direct bacterial identification in positive blood cultures by use of two commercial matrix-assisted laser desorption ionization–time of flight mass spectrometry systems. J clin microbial 50: 1313-1325.
10. Marko DC, Saffert RT, Cunningham SA, Hyman J, Walsh J, et al. (2012) Evaluation of the Bruker Biotyper and Vitek MS matrix-assisted laser desorption ionization–time of flight mass spectrometry systems for identification of nonfermenting Gram-negative bacilli isolated from cultures from cystic fibrosis patients. J clin microbial 50: 2034-2039.
11. Raoof WM, Latif IAR (2010) In vitro study of the swarming phenomenon and antimicrobial activity of pyocyanin produced by *Pseudomonas aeruginosa* isolated from different human infections. Eur J Sci Res 47: 405-421.
12. Essar DW, Eberly L, Hadero A, Crawford IP (1990) Identification and characterization of genes for a second anthranilate synthase in *Pseudomonas aeruginosa*: interchangeability of the two anthranilate synthases and evolutionary implications. J Bacteriol 172: 884-900.
13. Kurachi M (1958) Studies on the biosynthesis of pyocyanine. Isolation and determination of pyocyanine. Bull Inst Chem Res Kyoto Univ 36: 163-173.
14. Kerr JR, Taylor GW, Rutmon A, Hoiby N, Cole PJ, et al. (1999) *Pseudomonas aeruginosa* pyocyanin and 1-hydroxyphenazine inhibit fungal growth. Clin Pathol 52: 385-387.
15. Makrand R, Prashant DS, Bhushan L, Sudhir B (2007) Detection, isolation and identification of phenazine -1-carboxylic acid produced by biocontrol strains of *Pseudomonas aeruginosa*. J Scientific and Industrial Res 66: 627-631.
16. Norman RS, Moellar P, McDonald TJ, Morris PJ (2004) Effect of pyocyanin on a crude-oil-degrading microbial community. Appl Environ Microbiol 70: 4004-4011.
17. Price-Whelan A, Dietrich LEP, Newman DK (2006) Rethinking 'secondary' metabolism: Physiological roles for phenazine antibiotics. Nat Chem Biol 2: 71-78.