

A Study on the Effects of *Walnut oil* on Plasma Levels of Testosterone Pre and Post Puberty in Male Rats

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ABSTRACT

Background: As there is little knowledge about the impact of walnut oil on the physiology of the animals' reproductive system, and considering the fact that walnut oil contains a high proportion of some important unsaturated fatty acids including alpha-linolenic acid (ALA) and Omega-3, the present study aimed to investigate the possible effects of walnut oil on plasma levels of testosterone pre and post puberty in male wistar rats.

Materials and Methods: In this experimental study, 48 male wistar rats were randomly divided into two groups of pre and post puberty. Then each group was divided into three sub groups of control, sham and experimental, each of which with 8 animals. Walnut oil was administered by gavage at a dose of 20 mg/ kg daily at the specified time (10 am) for 30 days. Blood samples were obtained by cutting off the tail and plasma levels were measured by using a radioimmunoassay method (RIA).

Results: In the groups receiving walnut oil in the pre-puberty period, testosterone levels significantly increased ($P < 0.05$). In the groups receiving walnut oil post puberty, testosterone levels significantly increased, too ($P < 0.01$). A significant difference between pre puberty and post puberty groups was observed. ($P < 0.001$)

Conclusion: This study showed that walnut oil had stimulating effects on the male reproductive system and could increase plasma testosterone levels by influencing the pituitary - testicular axis.

Keywords- Walnut oil, Testosterone, Male rats.

INTRODUCTION

Walnut is the oldest tree food known to man with a longevity extended to thousands of years. Walnut originated in ancient Persia. That is why it is often referred to as the Persian walnut. Walnut contains significant amounts of antioxidants, omega-3 fatty acids and vitamin E, minerals, iron, sodium, calcium, magnesium, manganese, copper, potassium, phosphorus, and also protein and fiber, which have made it a varied nutritious meal¹.

Walnut belongs to the family of *Juglandaceae* and has been widely used in traditional medicine throughout the world. The genus is called *Juglandaceae* because there are 5 – hydroxyl - 1, and 4-naphthoquinonein its leaf, fruit shell, wood and root². Walnut oil, is extracted from English walnut (also known as the Persian walnut). This yellowish oil contains all lipid groups. The maximum percentage of which are triglycerides and sterile esters (96.9%). And only 3.1% of its lipids are polar lipids. 73.4% of polar lipids are sphingolipids and the remaining 26.6% are phospholipids. In walnut oil, however, these amounts are 2.3% and 0.8%, respectively^{3,4}. Their major components are triglycerides, mono cyclic unsaturated fatty acids (mainly oleic acid) and polycyclic unsaturated fatty acids (linoleic and alpha-linolenic acid), which are present in high levels. The presence of other organic substances, such as phenols, tocopherols and phytosterols, has been also proved³.

Walnut oil is also a good source of omega-3 fatty acids that are essential for human nutrition⁴. The major fatty acids found in walnut oil are linoleic, oleic acid and linoleic acid⁵. The preventive roles of monounsaturated fatty acids and polycyclic unsaturated fatty acids (MUFA and PUFA) in cardiovascular diseases have been identified.

It has been reported that the consumption of walnut (kernel and oil) lowers blood cholesterol levels^{6,7}. Studies have shown that walnut oil has antioxidant properties and reduces coronary heart disease risks, inflammation, and is useful in the treatment of skin disease and high blood pressure⁸⁻¹³. Walnut kernels are used to reduce blood lipids, that is, to increase high density lipoprotein, and to reduce low density lipoprotein^{14,15}. Walnut is also effective in the treatment of type 2 diabetes and enhancing cardiovascular flexibility¹⁶. It has been reported that due to its high concentration of natural antioxidants, walnut can be consumed as a protection against certain types of cancer. It may also reduce the risk of cardiovascular diseases^{17,18}.

Testosterone is a steroid hormone from the androgen group and is found in mammals, reptiles¹⁹, birds²⁰ and other vertebrates. In mammals, testosterone is primarily secreted in the testes of males and the ovaries of females; however, small amounts are also secreted by the adrenal glands. It is the principal male sex hormone and an anabolic steroid product. In men, testosterone has a key role in the development of male reproductive tissues like testis and prostate. In addition, it improves the secondary sexual characteristics such as increasing muscle mass, bone mass and hair growth²¹. Testosterone is also essential to health maintenance²² and to prevent osteoporosis²³.

Testosterone is a major form of androgen derived from sterol C-19. Like all other steroid hormones, testosterone, is predominantly synthesized from cholesterol in the Leydig cells of the testes²⁴. The testicular Leydig cells, are influenced by the central nervous system. The hypothalamus controls the pituitary gland, which results in the production of luteinizing hormone (LH) and follicle-stimulating hormone (FSH) in both sexes. However, females, mainly produce testosterone, and males mainly

produce estrogen²⁵. In addition, based on the type of the receptor that is expressed in cells, different hormonal responses can be generated. The hypothalamus produces gonadotropin releasing hormones (G_nRH) which stimulate FSH and LH to enter blood circulation from the pituitary gland and then to spread to the Leydig cells in order to produce testosterone²⁶.

On the other hand, walnut can be considered as an effective drug for impotency because of its compounds such as: niacin, which plays an important role in the production of steroid hormones by the adrenal gland; arginine and aspartic acid which have stimulatory effects on G_nRH and LH; oleic acid which is a 5 - alpha reductase inhibitor; and bromine (Br) which increases estrogen in postmenopausal women.

Since very few studies had been conducted on the effects of walnut oil on the reproductive system²⁷⁻³², this survey was designed to study the effect of walnut oil on hormonal changes in the reproductive system of male rats, the results of which could be important in the area of pharmaceutical applications of walnut oil, especially in reproductive system.

MATERIALS AND METHODS

Animal

The present study was an experimental - laboratory research in which the control and the treated samples were compared. At first, a number of male and female rats within the weight range of 180-200gr and on the verge of puberty were purchased from Razi Vaccine & Serum Research Institute, Fars branch (Iran) and were kept in the animal center of Arsanjan Azad University. During the experiment, the animals were kept in standard conditions of temperature, light and moisture, and had free access to food and water. Before getting started, rats spent 4 days at adaptation

condition to remove stress and to adapt to new conditions.

Measuring testosterone levels before and after puberty

Then rats were put together for mating. After the babies were born, male rats were isolated and 24 rats were randomly put into three groups of 8 rats as the pre-puberty group and the next 24 were similarly put into 3 groups of 8 rats as the post puberty group. For all the members of the pre- puberty group the test and gavage started at day 20 after birth and continued up to day 50 for 30 days. Then on the fiftieth day, blood samples were collected from rats. But in post puberty groups the test began on day 50 after birth, just when the rats reached puberty and it continued over a period of 30 days. On day 80 bloods sampling from these rats was done.

Preparation of walnut oil and gavage

Walnuts were collected from Bavanat (Fars province of Iran) in late spring and a sample was kept in the herbarium of Medicinal Plants Research Center, Islamic Azad University of Arsanjan. After cleaning and drying in the shade, walnut kernels were ground into powder. Then, a certain amount of powdered walnut was kept in a solvent called n - hexane in the lab condition for 24 hours, then it was strained and the solution obtained was poured into the rotary device (RV10D, IKA England) at 40-50°C to let the solvent evaporate. To ensure that there was no moisture left it was kept in a desiccator device (GCD-051X, KIKO, Japan) which had a powerful vacuum pump, for another 24 hours. At the end a bright yellow oily substance with the concentration of 1.1363gr/mL suitable for gavage was obtained (Figure 1). Gavage was done once a day at 10 am for 30 days. At day 31, blood samples were taken from the animals. To do so, the animals were anesthetized by

chloroform and blood samples were taken from tail end. Then, blood serums were separated from blood samples; finally, by using a radioimmunoassay method (RIA) the serum testosterone levels were measured.

Statistical analysis

The results were analyzed by using SPSS version 11.5 and independent-sample T test.

RESULTS

Statistical analysis showed that the plasma testosterone level in pre-pubertal rats receiving walnut oil had significantly increased ($P < 0.05$). Also in the groups receiving walnut oil post puberty the testosterone levels significantly increased ($P < 0.01$). Significant difference was observed between the post puberty groups and the pre- puberty ones ($P < 0.001$). Table 1 represents the plasma levels of testosterone in the control, sham and experimental groups, pre and post puberty.

DISCUSSION

The results showed that testosterone levels in the experimental group receiving a dose of 20 g/ kg walnut oil had a significant increase both before and after puberty compared to the control group ($P < 0.00$). According to the results of this research the increase in testosterone level indicates the positive effect of walnut oil on pituitary – testicular axis.

The hypothalamus - pituitary – testicular axis can be affected by various negative and positive feedbacks. Nitric oxide (NO) is one of the factors affecting this axis. High levels of Arginine in walnut can be converted to nitric oxide.

Nitric oxide increases the release of G_nRH , which in its turn increases gonadotropin secretion by activating neuron

nitric oxide synthase enzyme in the pituitary gland^{33,34}.

Nitric oxide activates Guanylate cyclase enzyme that causes the release of cyclic guanosine monophosphate and eventually by raising G_nRH , LH and FSH, enhances sperm motility and induces erection in males²⁸.

Aspartic acid, which is one of the amino acids found in walnut, has a stimulatory effect on the secretion of LH and G_nRH . Experiments have shown that this amino acid regulates the synthesis of testosterone and LH through cyclic guanosine mono-phosphate (cGMP), and cyclic adenosine mono-phosphate (cAMP) as the second messengers in pituitary and testes, respectively³⁵. Increased testosterone level in this experiment has had been the secondary consequence of increased G_nRH , particularly the LH.

Walnuts contains a large amount of polyunsaturated fatty acids such as linolenic acid, linoleic acid and oleic acid³⁶. Studies have shown that in a large number of androgen-sensitive organs such as the prostate, testosterone via 5 alpha – reductase is converted to 5-alpha – dihydrotestosterone.

Unsaturated fatty acids are able to inhibit 5 alpha - reductase in cell cultures and cell-free systems. Isomers of cis-linolenic acid, linoleic acid and oleic acid have been shown to have great power in the inhibition of 5 alpha – reductase, thereby preventing the conversion of testosterone to dihydrotestosterone. In this way walnuts can prevent the reduction of plasma testosterone level³¹. However, it appears that the increase in testosterone levels with walnut oil was due to its direct effect on Leydig cells, as well as its interfere with the biosynthesis of testosterone which was probably done through stimulating the synthesis of prostaglandins series 2.

Walnut kernels contain alpha-linolenic acid, which can be converted to

arachidonic acid, as a precursor to make type 2 prostaglandins like E₂³⁷. Arachidonic acid seems to play an important role in testicular steroidogenesis. As research indicates arachidonic acid increases cyclic adenylylase, thus enhancing the rate of cholesterol side-chain breakage and stimulating the production of testosterone. So, these compounds mediate the testosterone production via messaging. Studies on a kind of fish showed that all E-series prostaglandins stimulated testosterone production in the testes. E₂ was more powerful than E₁ and E₃³⁸.

Both n-3 and n-6 polyunsaturated fatty acids can affect reproductive processes through different mechanisms. They provide precursors for the synthesis of prostaglandins and ultimately regulate the expression patterns of key enzymes involved in the metabolism of steroids and prostaglandins¹⁵.

CONCLUSIONS

The increase in testosterone concentration indicated the positive impact of walnut oil on the hypothalamic - pituitary - testicular axis. These positive effects could be done in several mechanisms some of which were mentioned in the discussion section above. According to this study, this effect was exerted both pre and post puberty. However, the post pubertal effects of walnut oil ingredients were much more than those of pre pubertal ones which could be caused by the completion of the hypothalamic-pituitary-gonadal axis (HPG axis). Yet, further study in this area is necessary.

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Conflict of interest

None declared.

REFERENCES

1. Cosmulescu S, Baciú A, Achim G, Botu M, *et al.* Mineral Composition of Fruits in Different Walnut (*Juglans regia* L.) Cultivars. *Cluj* 2009; 37 (1), 156-160.
2. Mokhtari M, Abedinzade M, Naseran N. Effect of Walnut (*Juglans regia*) Extract on Serum LH, FSH and Testosterone Levels in Adult Male Rat. *J Ardabil Univ Med Sci.* 2012; 12(2):157-16.
3. Martinez ML, Mattea MA, Maestri DM. Varietal and crop year effects on lipid composition of walnut (*Juglans regia* L.) genotypes. *J Am Oil Chem Soc.* 2006; 83:791-796.
4. Ozcan MM, Iman C, Arslan D. Physico-chemical properties, fatty acid and mineral content of some walnuts (*Juglans regia* L.) types. *Agricultural Sciences*, 2010; 1(2), 62-67.
5. Tsamouris G, Hatziantoniou S, Demetzos C. Lipid analysis of Greek walnut oil (*Juglans regia* L.). *Z Naturforsch C.* 2002; 57(1):51-6.
6. Rajaram S, Haddad EH, Mejia A, Sabate J. Walnuts and fatty fish influence different serum lipid fractions in normal to mildly hyperlipidemic individuals: a randomized controlled study. *Am J Clin Nutr.* 2009; 89(5):1657-1663.
7. Damasceno NR, Perez-Heras A, Serra M, Cofan M, Sala-Vila A, Salas-Salvado J, *et al.* Crossover study of diets enriched with virgin olive oil, walnuts or almonds. Effects on lipids and other cardiovascular risk markers. *Nutr Metab Cardiovasc Dis.* 2011; 21(1):14-20.
8. Diana O, Labuckas D. Phenolics from walnut (*Juglans regia* L.) kernels: Antioxidant activity and interactions with proteins. *Food Chem.* 2008; 107(2):607-612.

9. Reiter RJ, Manchester LC, Tan D. Melatonin in walnuts: Influence on levels of melatonin and total antioxidant capacity of blood. *Nutrition*. 2005; 21(9):920-924.
10. Fladman EB. The scientific evidence for a beneficial health relationship between walnuts and coronary heart disease. *J Nutr*. 2002; 132:1062-1101.
11. Qadan F, Thewainni Aj, Ali DA, Afifi R, Elkhawad A, Matalaka KZ. The antimicrobial activities of psidium guajava and Juglans regia leaf extract to acne-developing organisms. *J Chin Med*. 2005; 33(2):197-204.
12. McPherson A. Juglans regia, common walnut, black walnut. *Nutr*. 2005; 65(1):3-10.
13. Deirdre K, Frank B. Effects of walnut consumption on blood lipids and other cardiovascular risk factors: a meta-analysis and systematic review. *Am J Clin Nutr*. 2009; 90(1):56-63.
14. Iwamoto M, Imaizumi K, Sato M, Hirooka Y, Sakai K, Takeshita A. Serum lipid profiles in Japanese women and men during consumption of walnuts. *Eur J Clin Nutr*. 2002; 56(7):629-37.
15. Tavakoli Darestani A, Kimiagar SM, Velaei N. Persian. Walnut effect on serum lipids in postmenopausal women. *J. Mazandaran Uni. Med. Sci*. 2005; 14(44):21-32.
16. Tapsell LC, Batterham MJ, Teuss G, Tan SY, Dalton S, Quick CJ. Long-term effects of increased dietary polyunsaturated fat from walnuts on metabolic parameters in type II diabetes. *Eur J Clin Nutr*. 2009; 63(8):1008-1015.
17. Miraliakbari H, Shahidi F. Antioxidant activity of minor components of tree nut oils. *Food Chem*. 2008; 111:421-427.
18. Yang J, Liu RH, Halim L. Antioxidant and anti-proliferative activities of common edible nut seeds. *LWT-Food Sci Technol*. 2009; 42:1-8.
19. Cox R, John-Alder H. Testosterone has opposite effects on male growth in lizards (*Sceloporus* spp.) with opposite patterns of sexual size dimorphism. *J. Exp. Biol*. 2005; 208(24):4679-4687.
20. Read W, Clark M, Parker P, Raouf S, Arguedas N, Monk D, Snajdr E, Nolan V, Ketterson E. Physiological effects on demography: a long-term experimental study of testosterone's effects on fitness. *Am. Nat*. 2006; 167(5):667-683.
21. Barrett KE, Barman SM, Boitano S, Brooks H. Ganong's Review of Medical Physiology, 24th Ed. McGraw- Hill Pub.2012;section 3, chapter 23, p 419.
22. Bassil N, Alkaade S, Morley J. The benefits and risks of testosterone replacement therapy: a review. *Ther Clin Risk Manag*. 2009; 5(3):427-448.
23. Tuck S, Francis R. Testosterone, bone and osteoporosis. *Front Horm Res*. 2009; 37:123-132.
24. Riggs BL, Khosla S, Melton LJ. Sex steroids and the construction and conservation of the adult skeleton. *Endocr. Rev*. 2002; 23:279-302.
25. Burger HG. Androgen production in women. *Fertil. Steril*. 2002; 77(4):S3-S5.
26. Neill JD. GnRH and GnRH receptor genes in the human genome. *Endocrinology*. 2002; 143:737-743.
27. Leighton RF, Gordon NF, Small GS, Davis WJ, Ward ES. Dental and gingival pain as side effects of niacin therapy. *Am Coll Chest Physic*. 1998; 114(5):1472-1474.
28. Miraglia E, De Angelis F, Gazzano E, Hassanpour H, Bertagna A, et al. Nitric oxide stimulates human sperm motility via activation of the cyclic GMP/protein kinase G signaling pathway. *Reproduction*. 2011; 141(1):47-54.
29. Sato Y, Tsukamamoto T. Effects of nitric oxide stimulation on the brain. *Drug Today*. 2000; 36(2):83-92.
30. D Aniello A. D-aspartic acid: An endogenous amino acid with an important neuroendocrine role. *Brain Res Rev*. 2007; 53(2):215-234.
31. Park WS, Son ED, Nam GW, Kim SH, Noh MS, Lee BG, et al. Torilin from *Torilis japonica*, as a new inhibitor of testosterone 5 alpha-reductase. *Planta Med*. 2003; 69(5): 459-61.
32. Raton B, James A. Handbook of phytochemical constituents of herbs and other economic plants. USA: CRC Press; 2000. p. 59-207.

33. Gonzalez LC, Pinilla L, Tena-Sempere M, Bellido C, Aguilar E. Effects of systemic blockade of nitric oxide synthases on pulsatile lh, prolactin, and GH secretion in adult male rats. *Horm Res.* 2001; 55(5):229-235.
34. Barnes MJ, Lapanowski K, Rafols JA, Lawson DM, Dunbar JC. Chronic nitric oxide deficiency is associated with altered leutinizing hormone and follicle-stimulating hormone release in ovariectomized rats. *Exp Biol Med.* 2002; 227(9):817-22.
35. Topo E, Soricelli A, D'Aniello A, Ronsini S, D'Aniello G. The role and molecular mechanism of D-aspartic acid in the release and synthesis of LH and testosterone in humans and rats. *Reprod Biol Endocrinol.* 2009; 27; 7:120.
36. Aiello F, Garofalo A, Aloisi AM, Lamponi S, Magnani A, Petroni A. Synthesis of esters of androgens with unsaturated fatty acids for androgen requiring therapy. *J Endocrinol Invest.* 2013; 36(6):390-5.
37. Kobayashi M, Hori T, Kawakami E. Changes in prostaglandin E2 levels in seminal plasma during ejaculation and the effect of exogenous prostaglandin E2 on semen volume in the dog. *J Vet Med Sci.* 2013; 75(9):1249-52.
38. Wade MG, Van der Kraak G. Arachidonic acid and PGE2 stimulate testosterone production. *Gen Comp Endocrinol.* 1993; 90(1):109-118.

Table 1. Results of T test analysis of all pre-pubertal and post pubertal groups

P	Testosteroneng/ml	Group	
-	.84±.05	Control	Pre-puberty
NS	.88±.05	Sham	
P<0.05	1.34±.17	Experimental	
-	1.91±.14	Control	Post puberty
NS	2.07±.09	Sham	
P<0.01	3.70±.40	Experimental	

Values of P are presented in comparison to the sham group, represents no significant difference (NS) compared with control group.

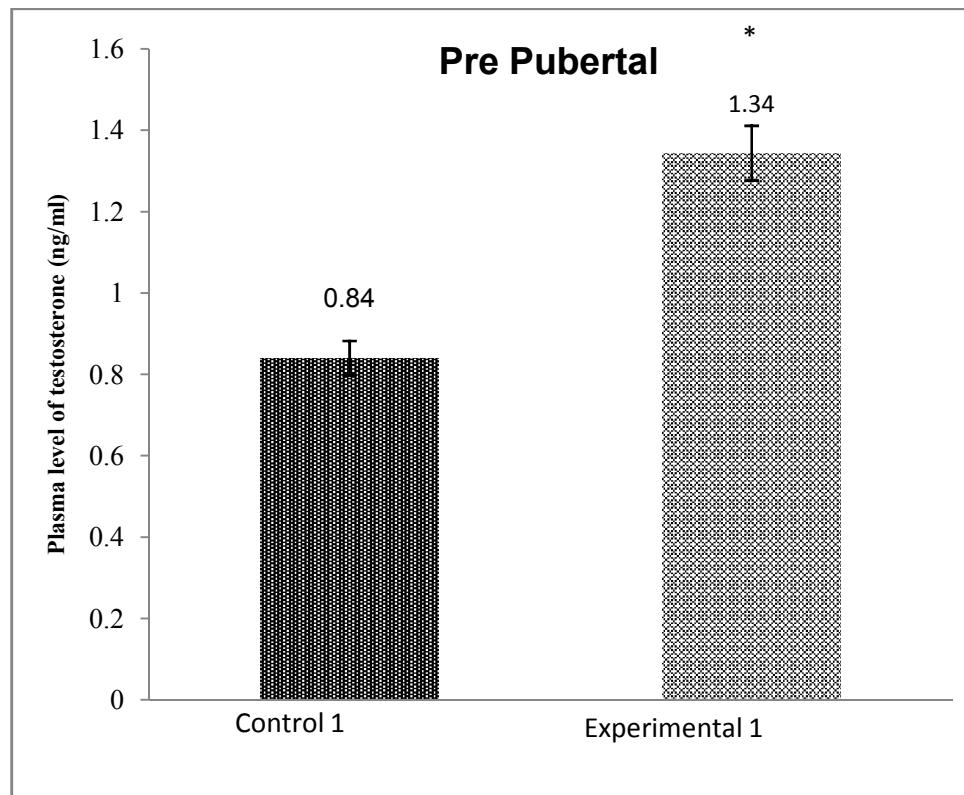


Figure 1. Compares the plasma testosterone levels between experimental and control groups in the pre-pubertal period. Each column represents the mean \pm standard error (Mean \pm SEM). Difference at $P < 0.05$ was significant

* indicates significant difference with control group at $P < 0.05$.

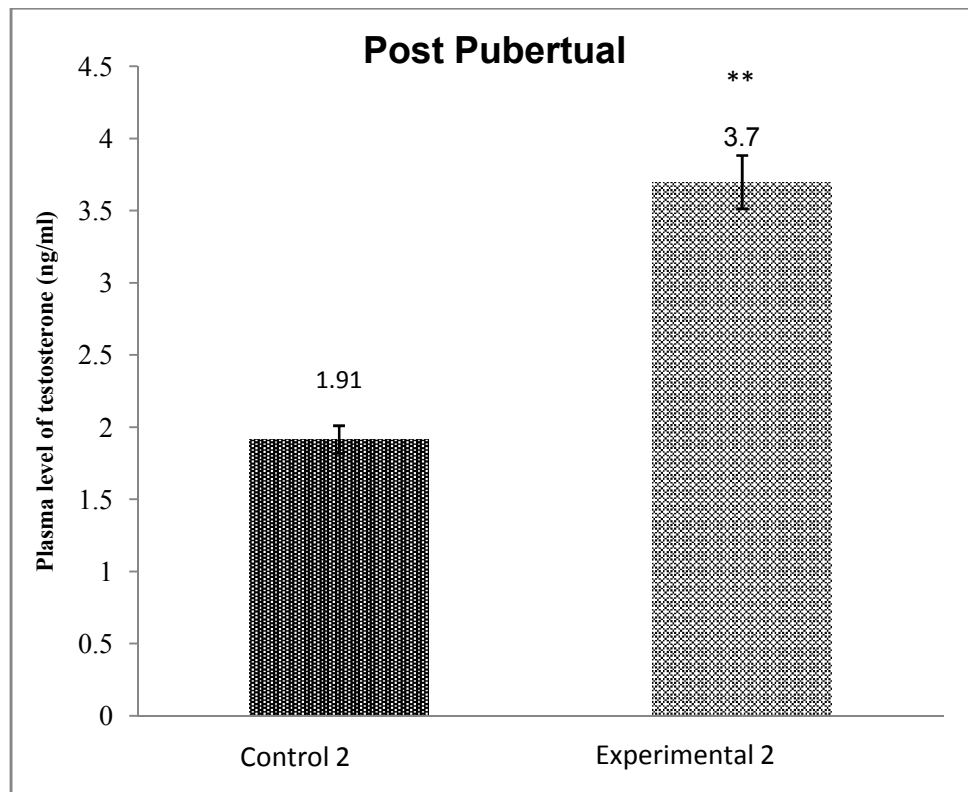


Figure 2. Compares the testosterone levels between experimental and control groups in post pubertal period. Each column represents the mean \pm standard error (Mean \pm SEM). Difference at $P < 0.01$ was significant

** indicates significant difference with control group at $P < 0.01$.

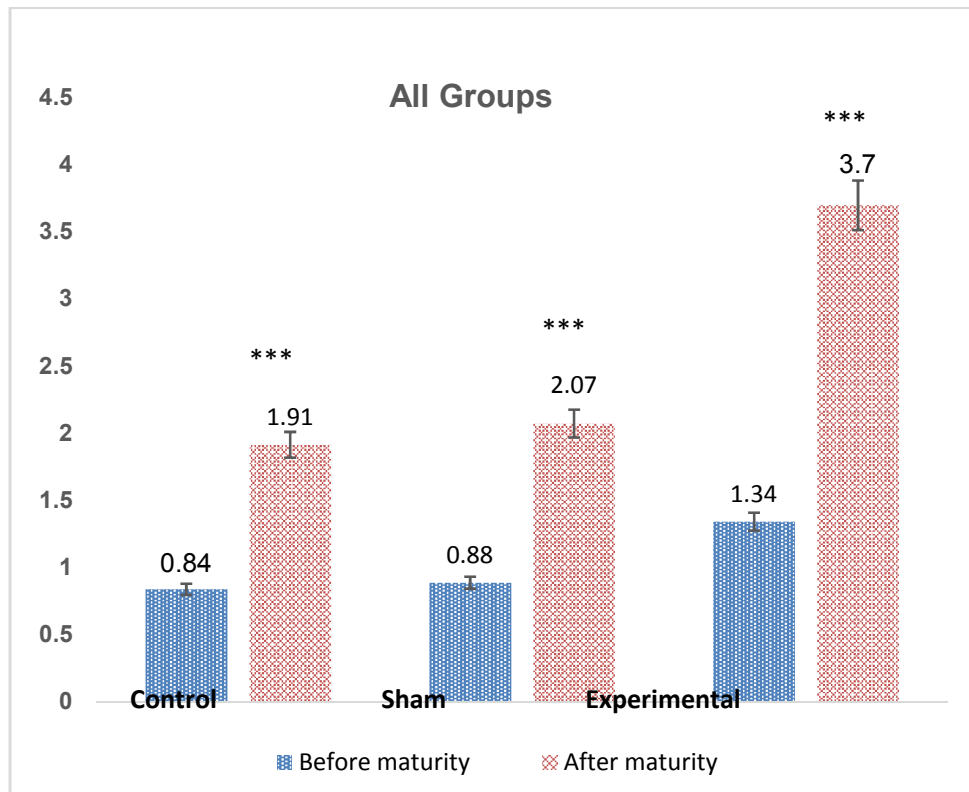


Figure 3. Compares the Testosterone levels among all groups of control, sham and experimental pre and post puberty

** indicates significant difference with pre and post-puberty groups at $P < 0.001$.

