Role of Ethylene inhibitors in tissue culture

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Ethylene is a plant hormone which controls growth and senescence of plants. It is also produced by plant tissues developed in vitro. At times ethylene accumulates in large amount in the cultures, especially in suspension cultures or rapidly growing non-differentiated callus, thus it probably affects the growth and development in various plant systems. This review exhibits the role of agents which inhibits accumulation of ethylene in the different types of tissue culture systems. Although in some cases its influence seems negligible but in many types of tissue culture ethylene may act either as a promoter or inhibitor depending on the type of culture. Ethylene plays an important role in in vitro regeneration of plants.

Introduction

Profitable shoot organogenesis technique for plant regeneration is based on proper establishment of medium components, an appropriate explant and regulation of the physical environment (Brown et al., 1986). Ethylene (C2H4) is the main factors of physical environment in plant tissue culture which is a gaseous plant hormone. It plays an important role in plant growth and development(Kende et al., 1993). Similar researches has been done in the previous years using ethylene inhibitors, like cobalt chloride (CoCl2), aminoethoxyvinylglycine (AVG), silver nitrate (AgNO3), benzyl isothiocyanate (BITC), 1methylcyclopropene (1-MCP), aminocarboxypropionic acid, polyamines, 3,4,5trichlorophenol, silver thiosulphate (STS) and salicylic acid (2-hydroxybenzoic acid) for instigating shoot organogenesis in different plant species (Bai et al., 2013). Recently some in vitro plant regeneration studies were conducted in various plants has been reported (Ruduś et al., 2013; Siddikee et al., 2012; Luo et al.,2014).

Ethylene biosynthesis

Biosynthesis of ethylene needs intensive study. The biosynthesis of ethylene begin with conversion of the

amino acid methionine to S-adenosyl-Lmethionine by the enzyme Met Adenosyltransferase. SAM is consequently converted to 1- aminocyclopropane-1carboxylic-acid (ACC) by the enzyme ACC synthase (ACS). The action of ACS is the rate-limiting step in ethylene synthesis. The closing step involves oxygen and enzyme action of ACC-oxidase (ACO), also known as the ethylene forming enzyme (EFE) (Pech et al., 2010). Establishment of S-adenosylmethionine (SAdoMet) and ACC as the precursors of ethylene becomes a major breakthrough in the ethylene synthesis pathway. Based on this fact, the enzymes catalyzing these reactions were characterized and purified. The molecular cloning of the ACC (Jafari et al., 2013) and ACO (Shi et al., 2012) genes results in demonstration of these enzymes that fit in to a multigene family and are controlled by a complex network of developmental and environmental signals responding to both internal and external stimuli. Over and above being an indispensable building protein synthesis block, almost 80% of cellular methionine is converted to S-AdoMet by S-AdoMet synthetase at the rate of utilization of ATP (Osborne et al., 1996). S-AdoMet being the foremost methyl donor in plants, is used as a substrate for numerous biochemical pathways going on in plants simultaneously, including polyamines and ethylene biosynthesis (Ravanel et al. 1998).

Mechanisms of action of ethylene inhibitor on inhibition of ethylene production

Active elements of ethylene inhibitor compounds are capable of producing ethylene insensitivity in plants (Wang et al. 2004). Ethylene-insensitive mutations (Stepanova *et al.*, 2005) interrupt with the ethylene binding sites (Sisler *et al.*, 2003). ETR1, ethylene receptor, have one ethylene-binding site per homodimer and binding is intervened by a single copper ion present in the ethylene-binding site. The replacement of the copper co-factor by inhibitor element also functions in locking the receptor into such a conformation where it continuously represses ethylene responses (Sisler *et al.*, 2003).

Conclusion

In this review, an attempt has been made to discuss the role of ethylene, its biosynthesis and the mechnanism how ethylene inhibitors interact and inhibit the action of ethylene on the plant cells. Although a lot of studies require the complete process. Current communication exhibits new dimensions in understanding plant morphogenesis. Thus, it is essential to explicate the physiological mechanisms at the gene regulation level to find out the definite role of ethylene inhibitors in signaling and to get to know how they influence regulation of ethylene action in plants.

References

Brown, D.C. and Thorpe, T.A., 1986. Plant regeneration by organogenesis. *Cell culture and somatic cell genetics of plants*, *3*, pp.49-65.

Kende, H., 1993. Ethylene biosynthesis. *Annual review of plant biology*, 44(1), pp.283-307.

Bai, B., Su, Y.H., Yuan, J. and Zhang, X.S., 2013. Induction of somatic embryos in Arabidopsis requires local YUCCA expression mediated by the downregulation of ethylene biosynthesis. *Molecular plant*, 6(4), pp.1247-1260.

Ruduś, I., Sasiak, M. and Kępczyński, J., 2013. Regulation of ethylene biosynthesis at the level of 1aminocyclopropane-1-carboxylate oxidase (ACO) gene. *Acta Physiologiae Plantarum*, *35*(2), pp.295-307.

Siddikee, M.A., Chauhan, P.S. and Sa, T., 2012. Regulation of ethylene biosynthesis under salt stress

in red pepper (Capsicum annuum L.) by 1-aminocyclopropane-1-carboxylic acid (ACC)

deaminase-producing halotolerant bacteria. *Journal* of plant growth regulation, 31(2), pp.265-272.

Luo, X., Chen, Z., Gao, J. and Gong, Z., 2014. Abscisic acid inhibits root growth in Arabidopsis through ethylene biosynthesis. *The Plant Journal*, *79*(1), pp.44-55.

Pech, J.C., Latché, A. and Bouzayen, M., 2010. Ethylene biosynthesis. In *Plant Hormones* (pp. 115-136). Springer, Dordrecht.

Jafari, Z., Haddad, R., Hosseini, R. and Garoosi, G., 2013. Cloning, identification and expression analysis of ACC oxidase gene involved in ethylene production pathway. *Molecular biology reports*, *40*(2), pp.1341-1350.

Shi, H.Y. and Zhang, Y.X., 2012. Pear ACO genes encoding putative 1-aminocyclopropane-1carboxylate oxidase homologs are functionally expressed during fruit ripening and involved in response to salicylic acid. *Molecular biology reports*, *39*(10), pp.9509-9519.

Osborne, D.J., Walters, J., Milborrow, B.V., Norville, A. and Stange, L.M., 1996. Special publication evidence for a non-ACC ethylene biosynthesis pathway in lower plants. *Phytochemistry*, 42(1), pp.51-60.

Wang, K.L.C., Yoshida, H., Lurin, C. and Ecker, J.R., 2004. Regulation of ethylene gas biosynthesis by the Arabidopsis ETO1 protein. *Nature*, *428*(6986), pp.945-950.

Stepanova, A.N., Hoyt, J.M., Hamilton, A.A. and Alonso, J.M., 2005. A link between ethylene and auxin uncovered by the characterization of two rootspecific ethylene-insensitive mutants in Arabidopsis. *The Plant Cell*, *17*(8), pp.2230-2242.

Sisler, E.C. and Serek, M., 2003. Compounds interacting with the ethylene receptor in plants. *Plant Biology*, *5*(05), pp.473-480.