

Phytochemical Investigation of High Altitude Medicinal Plants *Cinnamomum tamala* (Buch-Ham) Nees and Eberm and *Rhododendron arboreum* Smith

Shagun Gill¹, Preeti Panthari² and Harsha Kharkwal*³

¹Amity Institute of Biotechnology, Amity University Uttar Pradesh, Noida-201303

²Amity Institute of Phytomedicine and Phytochemistry, Amity University Uttar Pradesh, Noida-201303

³Amity Center for Carbohydrate Research, Amity University Uttar Pradesh, Noida-201303

Address for Correspondence

Amity Center for Carbohydrate Research, Amity University Uttar Pradesh, Noida-201303

E-mail: hkharkwal@amity.edu

ABSTRACT

The present review provides a detailed information about two high altitude medicinal plants, namely *Cinnamomum tamala* and *Rhododendron arboretum* used in curing a number of ailments since ancient time. It highlights the work done since last one decade on chemical constituents, uses, formulations and pharmacological activities of these important medicinal species. These medicinal plants have therapeutic value that aids for alleviating various ailments of humankind. Both these plants are known to have antioxidant, antifungal, antimicrobial activities, anti-inflammatory and anti-diarrhoeal activities.

Keywords: Pharmacology, Phytochemical, Antioxidant, Antimicrobial, Anti-inflammatory, Anti-diarrhoeal.

INTRODUCTION

High altitude medicinal plants are used worldwide in both traditional and Western medicine systems. India is known for its vast biological diversity and knowledge rich ancient traditional systems of medicine, which provides a strong base for utilization of a large number of plants in general health care. The high altitude ecosystem is considered to be hotspots of medicinal plant diversity and are most

neglected regions for research in view of their inaccessibility and harsh climatic conditions. In a survey, it was found that only 20% high altitude medicinal plants are used in Indian drug trade which is collected from wild sources¹.

Proper methodologies for the research and development are the need of the day for tapping the full therapeutic potential of plants. To find out the efficacy

of medicinal plants one needs to analyze the phytochemicals present in it. Scientists and pharmaceutical companies are working out for the development of new and effective drugs all over the world. Phytochemicals are the natural chemical compounds which are present in plants, having various characterized property such as antioxidant, anti-bacterial, anti-fungal, anti-diabetic activity. The main purpose of phytochemical screening is to detect the presence of secondary metabolites such as phenols, saponins, coumarins, alkaloids, terpenoids and flavonoids which have great medicinal importance. Any drug is mostly derived from the phytochemicals present in such medicinal plants, which has been used since time immemorial. Alkaloids helps in controlling development system of living organisms, plays some metabolic role and have protective property². They are used as steroidal alkaloids as a medicine. While phenolic compounds and flavonoids are known to possess biological activity such as antioxidant, anti-inflammatory, anti carcinogenic etc². According to Okuda *et al* 1983 tannins have the reducing power which prevents liver injury by inhibiting the formation of lipid peroxides³. Flavonoids have been reported to exhibit the anti-oxidative, antiviral, antimicrobial and anti-platelet activities⁴. Polyphenols, particularly flavonoids, which are present in considerable amounts in fruits, vegetables, spices, medicinal herbs and beverages, have been used to treat many human diseases, such as diabetes, cancer and coronary heart disease⁵⁻⁶.

To determine bioactive compounds from plant parts successfully, it is necessary to use different solvent in extraction method which highlights the need to use as much solvent as possible in the screening of biologically active phytochemicals⁷.

Cinnamomum tamala and *Rhododendron arboretum* are the two

medicinal plants present at altitude of 900-2500m in tropical and sub-tropical Himalayas⁸ and above 1500m to 6000m in Himalayan forests or Nilgiri Hills in Southern India⁹ respectively and both have therapeutic and economical value.

THE POPULAR CHEMICAL CONSTITUENTS ISOLATED

Chemicals constituents which have been widely reviewed and investigated are Eugenol, Cinnamaldehyde from leaves of *C. tamala*. Leaves from Gorakhpur were analyzed for oleoresins present in them, it was found that the oleoresins varied with respect to the solvent used. With Methanol, ethanol, iso-octane and carbon tetrachloride 28, 34, 42 and 41 components were obtained respectively. Eugenol (66.1%), spathulenol (4.8%), viridiflorene (2.4%), methyleugenol (1.9%), aromadendrene (1.5%), with other minor compounds were detected¹⁰. Chemical structure of some isolated compound of *Cinnamomum tamala* (Nees and Eberm) is given in Fig. 1

While in case of *Rhododendron arboretum* isolated compounds are Taraxerol, Betulinic acid, Ursolic acid acetate from bark; Arbutin, Hyperoside, Amyrin, Epifridilenol from leaves; and Quercetin, Rutin, Coumaric acid from flower part¹¹. The chemical structure of some chemical constituents of *Rhododendron arboretum* is given in Fig. 2

The present review highlights the work done till now on the phytochemical and pharmacological aspect of these two medicinal plants and also gives a brief on the recent research and future prospects which will help in finding new lead compounds for drug discovery. The phytochemical screening test is given in Table 4.

PHYTOCHEMISTRY

The compounds present in *C. tamala* belonging to different localities varies widely due to environmental conditions prevailing over that region. The chemical constituents of essential oils present in the leaves and bark of *C. tamala* reported from North East India are given in Table 1.

While the Chemical constituents of *Rhododendron arboretum* from Himalayan region are listed in Table 2.

PHARMACOLOGICAL ACTIVITIES OF CINNAMOMUM TAMALA AND RHODODENDRON ARBOREUM

Antibacterial activity

Pankaj *et al* (2009) showed that the stem bark of *Cinnamomum tamala* has antibacterial activity. To prove it further they prepared aqueous, ethanol, methanol, ethyl acetate and hexane extract in a final concentration of 100mg/ml and performed Agar well diffusion assay to evaluate the antibacterial potential of *C. tamala* bark extract, and concluded that almost all extract of the stem bark showed variable degrees of inhibition zone against different bacterial species except hexane which was found inactive. Ethanol, methanol & ethyl acetate fraction were found to have significant activity against *Salmonella typhi*, *Staphylococcus aureus*, *Streptococcus pyogenes*, *Bacillus (subtilis, cereus)* except *E.coli*. Overall methanolic extract was found to be more effective with zone size ranging from 11.26 mm to 20.77 mm while ethanolic extract showed inhibition zone ranging from 11.83 mm to 17.90 mm; mild activity was shown by the ethyl acetate extract ranging from 12 mm to 15 mm. Aqueous extract was found to be effective against *Staphylococcus aureus* and *Bacillus cereus* with zone size 11 mm approx. against each bacterium²⁷. Vardar *et al* (2003) showed that volatile oil was 100% effective against

Fusarium moniliforme, *Aspergillus niger*, *A. oryzae* and *A. solani* but not for *A. awamori* in inverted patri-plate method²⁸. Md. Hemayet *et al* showed that the ethanolic extract of the leaves of *C. tamala* (500 µg/disc) showed moderate anti-microbial activity against *Staphylococcus epidermidis* (10mm), *Vibrio cologet* (Zone of inhibition 11mm), *Streptococcus agalactiae* (9mm), *Shigella sonnei* (9 mm), *Streptococcus pyogenes* (9mm), *Staphylococcus saprophyticus* (11mm), *Staphylococcus aureus* (8 mm).

Mohammad *et al* (2013) reported the antibacterial activity of the methanolic extract of various parts such as such, as flowers, leaves, bark, stem and roots of *Rhododendron arboreum* by determining the zone of inhibition against the tested bacterial strains such as *Escherichia coli*, *Staphylococcus aureus*, *Bacillus subtilis* and *Salmonella typhi*.

Antifungal activity

A.K *et al*, 2012 reported that *C. tamala* has fungicidal characteristic. It was shown that oil extracted had the potent antifungal efficacy against *Aspergillus niger*, *A. fumigatus*, *Candida albicans*, *Rhizopus stolonifer* and *Penicillium spp.*, which was determined by measuring the diameter of the zone of inhibition (ZOI) surrounding wells, in agar diffusion assay and concluded that *Candida albicans* showed the maximum ZOI of 25 mm in comparison to other fungi.

The MFC values of oil against all the test fungi were found to be 230µg/ml. While the Minimum fungicidal concentration (MFC) of oil extracted was evaluated at four different concentrations of oil (115,230,460 and 690µg/ml) and was concluded that higher concentration of oil showed complete fungicidal activity and MFC values of oil against all fungal isolates tested were found to be 230µg/ml²⁹.

Nisar *et al* (2013) in his study reported that the various fractions of *Rhododendron arboreum* bark showed anti-fungal activity in agar well diffusion method. It was shown that higher concentration of extracts, i.e. 50 mg/ml exhibited maximum inhibition zone. For methanolic extract it was recorded to be 25 ± 1.19 , 29 ± 1.02 , 28 ± 1.98 , 17 ± 1.44 , 28 ± 1.56 , 32 ± 1.66 mm against fungi (*A. niger*, *C. albican*, *C. flavus*, *F. solani*, *M. canis* and *D. glaberata* respectively; while for ethyl acetate it was reported to be 22 ± 1.92 , 28 ± 1.02 , 25 ± 1.32 , 16 ± 1.16 , 24 ± 1.11 and 28 ± 1.00 respectively. The chloroform extract was reported to produce an antifungal effect which was lesser than ethyl acetate and methanolic extract while n-hexane extract failed to produce an outstanding fungicidal effect.

Antidiabetic activity

Palanisamy *et al* described the anti-diabetic activity of *Cinnamomum tamala* Linn, with special reference to its curative and protective role in streptozotocin-induced diabetic animal model. The efficacy of 50% ethanolic extract of the leaves of *Cinnamomum tamala* showed significant decrease in the blood glucose level and increase in the antioxidant efficacy in streptozotocin induced diabetes. It was demonstrated that the oral administration of the 50% ethanolic extract of leaves of *C. tamala* extract to streptozotocin induced diabetic rats showed significant positive changes in the biochemical and physiological parameters related to carbohydrate, protein and lipid metabolism³⁰. It has been shown by Subash *et al.* that oral administration of cinnamaldehyde produces a significant antihyperglycemic effect which lowers both total cholesterol and triglyceride levels and, at the same time, increases HDL-cholesterol in STZ induced diabetic rats³¹. Mega *et al*

(2010) showed the antidiabetic activity using α -amylase inhibition assay of methanolic and successive water extract of bark of *C. tamala* and concluded that methanol extract have high potent activity than later³². While Usha *et al* (2010) demonstrated that the administration of 250 mg/kg aqueous extract of *C. tamala* in streptozotocin induced diabetic rats showed significant decrease in levels of blood glucose and urine sugar³³.

M.R. Bhandary and J. Kuwabata (2008) examined anti-diabetic property in *R. arboreum* and isolated active compounds from it. Administration of the aqueous methanolic extract of the flower was found to exhibit inhibitory activity on the rat intestinal α -glucosidase. Both the water soluble and ethyl acetate-soluble portions from the aqueous methanolic extract showed inhibitory activities on α -glucosidase, revealing higher activity by the ethyl acetate soluble portion. α -glucosidase inhibitor quercetin-3-O- β -D-galactopyranoside (hyperin) was isolated from the ethyl acetate-soluble portion, through enzyme-assay guided separation. The isolated compound showed a dose dependent α -glucosidase inhibitory activity with IC50 values of 1.66 mM and 0.76 mM for sucrase and maltase, respectively. From this study, they could conclude that the flower possessed antidiabetic potential which could be utilized for medicinal preparations, nutraceutical or functional food for diabetes.

Anti-diarrhoeal activity

Md. Hemayet *et al* performed the castor oil induced diarrheal method in mice of ethanolic extract of *C. tamala* and showed that an extract inhibited the mean number of defecation which were 24.49% and 40.82% at doses of 250mg/kg and 500mg/kg respectively.

N. Verma *et al*, 2011 reported that the ethyl acetate fraction of *Rhododendron*

arboretum (flowers) has potent antidiarrheal activity. High performance thin layer chromatography (HPTLC) method was used for the determination of hyperin. Using HPLC the concentration of hyperin in flowers of *R. arboreum* was found to be 0.148%. Oral administration of EFRA in 100, 200 and 400 mg/kg showed dose-dependent and significant ($P < 0.05$ - 0.001) antidiarrhoeal potential in castor oil and magnesium sulfate-induced diarrhoea. EFRA was found to possess an anti enteropooling in castor oil-induced experimental animals by reducing both weight and volume of intestinal content significantly. It is also reported that the ethyl acetate fraction of *R. arboreum* flowers reduce magnesium sulfate-induced diarrhoea significantly, which could be due to increased absorption of water and electrolytes. The extract inhibits gastrointestinal motility in diarrhoea through anticholinergic effect. Phytochemical screening exhibit the presence of numerous constituents such as flavonoids, saponins, tannins, phytosterols, reducing sugars and phenolic compounds. Hence tannins, reducing sugars and sterols may be responsible for the mechanism of antidiarrhoeal activity of EFRA³⁴.

Antioxidant activity

The antioxidant potential of Cinnamomum oil and oleoresins for mustard oil has been evaluated by different methods such as peroxide, p- anisidine, thiobarbituric acid and total carbonyl value method. For determining antioxidant activity of essential oil and oleoresins of *Cinnamomum tamala* linoleic acid system and scavenging effect on DPPH³⁵⁻³⁶. An experiment was conducted to evaluate *in-vitro* antioxidant activity of Indian Bay Leaf on rat by S. Lakshmi Devi *et al* (2007) and reported increases in the levels of lipid and lipid peroxidation products and a decline in antioxidant potential in diabetic rat brain synaptosomes

when induced with methanolic extract of bay leaf. Extract of Bay leaf displayed scavenging activity against superoxide and hydroxyl radical in a concentrate dependent manner. It can be concluded that synaptosomes from diabetic rats are susceptible to oxidative damage³⁷.

A.K *et al* 2012 showed that a different extract of *C. tamala* (Petroleum ether, Benzene, Chloroform, Ethyl acetate, Acetone, ethyl alcohol, water) has antioxidant activity 66.3%, 34.8%, 38.9%, 5.2%, 31.1%, 28.5%, 4.0% respectively; after 3 hours using β -carotene linoleate model system³⁸.

Research carried out by Swamidasan *et al* (2008) on mice and rats had shown that ethanolic extract of *Rhododendron arboreum* exhibit adaptogenic property. For the evaluation of adaptogenic activity, anoxia stress tolerance, swimming endurance, immobilization stress models was used. Related treatment with ethanolic extract at doses 250 and 500 mg/kg, showed markedly increased in anoxia stress tolerance and swimming endurance time as compared to control group. Similarly, pre-treatment with extract showed marked decrease in blood glucose, cholesterol, triglyceride level as compared to stress control group in immobilization stress. Weights of the liver and adrenal glands are markedly decreased, but no weight changes in the spleen and testes were observed³⁹. The secondary flavonoids isolated from the leaves of *R. arboreum* were found to have potent antioxidant property⁴⁰.

Anti-ulcer activity

Lima *et al* (2010) showed that the hydro alcoholic extract of *C. tamala* leaves have the potential to protect the gastric mucosa from chemical, stress and physically induced ulcers. They inhibit gastric acid secretion probably by blocking H⁺ K⁺-ATPase action and offer antioxidant

protection against oxidative stress-induced gastric damage. The findings of this experimental study lead to further isolation, and pharmacological activity of new therapeutic compounds effective against ulcer⁴¹.

Anti-inflammatory activity

Thamizhselvam *et al* (2012) in his study evaluated that the methanolic extract of *C. tamala* has anti-inflammatory activity in the carrageenan induced paw edema in Wistar albino rats. The percentage inhibition of edema formation was 66.75% and 73.71% at 250 and 500 mg/kg dosage respectively⁴².

N. Verma *et al* (2011) has observed that the ethyl acetate extract fraction of *R. arboreum* exhibit remarkable anti-inflammatory and anti-nociceptive potential in animal models. Oral administration of *Rhododendron arboreum* extract (EERA) (100, 200 and 400 mg/kg) showed dose dependent and significant anti-inflammatory activity in arachidonic induced hind paw edema ($p < 0.01$), cotton pellet granuloma model of inflammation ($p < 0.01$) and Freund's adjuvant-induced paw arthritis ($p < 0.01$). A significant ($p < 0.05$) anti-nociceptive activity was evidenced in mice, protected in acetic acid-induced writhing. EERA at the dose of 100, 200 and 400 mg/kg showed significant ($p < 0.001$) resistance against analgesimeter induced pain in mice. The anti-inflammatory effect of the extract may be due to the presence of flavonoids (hyperin), tannins, saponins and other phytochemicals present either as single or in combination. The significant level of anti-inflammatory activity of the ethyl acetate extract could be attributed to the high amount of flavonoids present in the extract⁴³.

Alzheimer prevention Potential

Alzheimer is the most common form of dementia among older people, which affects person's ability to carry out daily activities. In a recent clinical study, it was found that two compounds (Cinnamaldehyde & Epicatechin) have potential to ward off or even prevent Alzheimer disease. The stated compound present has a protective power which can protect component protein of neuron, called tau, from oxidative stress, thus allowing neuron to function properly⁴⁴.

Unlike *C. tamala*, there is no report on Anti-Alzheimer activity for *Rhododendron arboreum*.

Hepatoprotective activity

Selvam *et al* (2010) reported the hepatoprotective activity of methanolic extract of *Cinnamomum tamala* (nees) against paracetamol intoxicated swiss albino mice.

N. Verma *et al* (2011) showed the hepatoprotective potential of the ethyl acetate fraction of *Rhododendron arboreum* against CCl_4 -induced liver damage in preventive and curative models. Fraction at a dose of 100, 200 and 400 mg/kg was administered orally once daily for 14 days in CCl_4 -treated groups (II, III, IV, V and VI). The serum levels of glutamic oxaloacetic transaminase (SGOT), glutamate pyruvate transaminase (SGPT), alkaline phosphatase (SALP), γ -glutamyltransferase (γ -GT), and bilirubin were estimated along with the activities of glutathione S-transferase (GST), glutathione reductase, hepatic malondialdehyde formation, and glutathione content. The substantially elevated serum enzymatic activities of SGOT, SGPT, SALP, γ -GT, and bilirubin due to CCl_4 treatment were restored to normal in a dose-dependent manner. Meanwhile, the decreased activities of GST and glutathione reductase were also restored to normal. In

addition, ethyl acetate fraction also significantly prevented the elevation of hepatic malondialdehyde formation and depletion of reduced glutathione content in the liver of CCl₄ -intoxicated rats in a dose-dependent manner⁴⁵. Some

Phytochemical Analysis

Qualitative phytochemical analysis of the aqueous, methanol, ethanol, ethyl acetate, chloroform and hexane extracts of *Cinnamomum tamala* was carried out to test the presence of phytochemicals such as alkaloid, flavonoids, terpenoids, sterols, tannins, glycosides (Table no. 4). Phytochemical screening of different solvent extracts of *Cinnamomum tamala* (Kasargod district of Kerala) and *Rhododendron arboreum* (Western ghats) is listed in the Table no. 5 and 6 respectively.

Future Prospects

Cinnamomum tamala and *Rhododendron arboreum* have beneficial effects with wider uses. Numerous studies have been conducted in different parts of both medicinal plants, but these plants have not yet developed as a drug by pharmaceutical industries. Newer concepts aligning with industrial norms for extracting oil, which has tremendous utility in the food industry can be developed.

The field of nanotechnology has evolved rapidly and this technology in the area of medicine can be considered as a boon to treat human disease in near future. Katti, director of Cancer Nanotechnology Platform (2010), discovered that phytochemicals present in *Cinnamomum* have the ability to treat cancer when combined with gold nanoparticles because active pharmaceutical ingredients are carried by gold nanoparticles into cancer cells and assist in the destruction or imaging of malignancies. Chanda *et al* (2011) showed

that cinnamon coated gold nanoparticles have the ability to detect cancer cells⁵¹.

This review elicits on all the aspects of the herb and throws attention to set the mind of the researchers to carry out the work for developing various formulations, which can ultimately be beneficial for the human beings as well as animals. Further research is required to isolate the bioactive principle of this plant as well as further studies on its bioefficacy against human pathogens. Moreover, chemistry of genus *Cinnamomum* is quite interesting as this medicinal plant exhibits several chemotypes within a species and very little attention has been paid to nonvolatile compounds and also a very little work has been reported in the bark and root oil composition of *C. tamala*, so this is an area worth exploring.

CONCLUSION

The review suggests that *Cinnamomum tamala* and *Rhododendron arboreum* both are high altitude medicinal plant having wide application in the fields like pharma, medicines, cosmetics, etc. Various parts are exported due to their high medicinal values, hence the protection of these endemic species should be stressed upon. *Rhododendron arboreum* is an ornamental tree and can be used in avenue decoration. *Cinnamomum tamala* a miraculous plant should be explored in the pharmaceutical industry for drug development. The plant has traditional importance and an in-depth systematic study of the different aspects of the plant is the need of the hour. The detailed cataloguing will help the researchers to carry out their research on this highly medicinal plant which will lead to new formulations both to man and animals. The essential oil and oleoresins possess high antioxidant and antimicrobial activity against bacterial and fungi. It can be concluded that there is a

huge scope of the oil in food industry as natural food additive¹⁰.

Rhododendron arboretum extracts have high antioxidant potential which can replace synthetic antioxidants in pharma field. In the treatment of cancer, suitable extract of different parts of the tree can be used. The review suggests that this plant is one of the important plants of the Himalayan region and needs protection and have a high commercial utility of different part extract.

ACKNOWLEDGEMENT

The authors are thankful to the Chancellor and Vice chancellor Amity University Uttar Pradesh for their constant support and guidance.

REFERENCES

1. Kaul M K. 2010. High altitude botanicals in integrative medicine-Case studies from Northwest Himalaya. *Indian Journal of traditional Knowledge*. 18-25.
2. P. Jayanthi, Thamaraiselvi P. Preliminary studies on phytochemicals and antimicrobial activity of solvent extracts of *Eichhornia crassipes* (Mart.) Solms. *Asian Journal of Plant Science and Research*, 2012, 2(2):115-122.
3. Okuda T, Kimura Y, Yoshida T, Hatano T, Okuda H, Arichi S. 1983. Studies on the activities of tannins and related compounds from medicinal plants and drugs. *Chem. Pharm. Bull.* 31: 1625–1631.
4. Middleton E, Kandaswami C.1993. The impact of plant flavonoids on mammalian biology: implications for immunity, inflammation and cancer.619-652.
5. Broadhurst C.L, Polansky M.M, Anderson R.A.2000. Insulin-like activity of culinary and medicinal plant aqueous extracts *in vitro*. *J. Agric. Food Chem.* 48: 849–852.
6. Fabri R.L, Nogueira M.S, Braga F.G, Coimbra E.S.2009. *Scio E. Mitracarpus frigidus* aerial parts exhibited potent antimicrobial, antileishmanial and antioxidant effects. *Biores. Technol.* 100: 428–433.
7. Charles S, Uche A, Ifeanyi O. Preliminary screening of different solvent extracts of stem bark and roots of *Dennetia tripetala* G. Baker. *Asian Journal of Plant Science and Research*. 2013; 3(3): 10-13.
8. J.Rema 2005), N.K Leela, P.A Mathew. Chemical composition of *Cinnamomum tamala* essential oil- A review. *Journal of medicine and Aromatic Plant Sciences*. 2005. 27:515-519.
9. Pankaj Kumar Sonar, Ranjit Singh, Shagufta Khan. Isolation, Characterization and activity of the flowers of *Rhododendron arboreum* (Ericaceae). *E-Journal of Chemistry*.2012. 9(2): 631-636
10. Kapoor IPS, Singh B, Singh G, Isidorov V, Szczepaniak L.2009. *Cinnamomum tamala* Nees & Eberm. (Tejpat) essential oil and oleoresins. *Natural Product Radiance*, 8(2): 106-116.
11. Srivastava P. 2012. *Rhododendron Arboreum: An overview Journal of Applied Pharmaceutical Science*, 02(01): 158-162.
12. J. Rema, N.K Leela.2005. Chemical composition of *cinnamomum tamala* essential oil. *Journal of medicinal and aromatic Plant sciences*. 515-519.
13. Shrestha M, Budhathoki N.P.2012. The chemical compositions of *Rhododendron arboretum*, "Laligunras". *J. Nepal chem. Soc.* Vol 30.
14. Rangaswamy S and Sambamurthy K. Crystalline chemical components of flowers of *Rhododendron nilagiricum* Zenk. *Proc math Sci.* 1960; 51(6): 322-327.
15. Gupta R.2009. Hypoglycaemic Activity of Ethanol Extract of *Cinnamomum tamala* Leaves in Normal and Streptozotocin Diabetic Rats.

- Iranian Journal of Pharmacology. 8: 17-21.
16. Chakraborty U, Das H. 2010. Antidiabetic and Antioxidant Activities of Cinnamomum tamala Leaf Extracts in Stz-Treated Diabetic Rats. *Global Journal of Biotechnology & Biochemistry*. 5(1):12-18.
 17. Bisht B, Sisodia S.S. 2011. Assessment of antidiabetic potential of C.tamala leaves extract in streptozotocin induced diabetic rats. *Indian Journal of Pharmacology*. 43(5): 582-585.
 18. Thamizhselvam N, Soumya S, Sanjayakumar YR, Salinichandran K, Jaya. N.2012. Anti-inflammatory, Analgesic and antipyretic activity of methanolic extract of Cinnamomum Tamala(Nees) in experimental Animal Models. *Int J Bioas*. 01(09)
 19. Ullah N, Khan M, Khan T, Ahmad W.2013. Protective Effect of Cinnamomum tamala Extract on Gentamicin-Induced Nephrotic Damage in Rabbits. *Tropical Journal of Pharmaceutical Research*. 12(2): 215-219.
 20. Dhan P, Garima U, Singh BN, Ruchi D, Sandeep K, Singh KK.2007. Free radical scavenging activities of Himalayan Rhododendrons. *Curr Sci*. 92(5): 26–32.
 21. Verma N, Singh AP, Amresh G, Sahu PK, Rao CV.2010. Anti-inflammatory and anti-nociceptive activity of *Rhododendron arboreum*. *J Pharm Res*.76-80.
 22. Verma N, Singh A, Rao.2011. Protective effect of ethyl acetate fraction of Rhododendron arboretum flowers against carbon tetrachloride-induced hepatotoxicity in experimental models. *Indian J Pharmacol*. 43(3): 291-295.
 23. Verma N, Singh P.2012. Antihyperglycemic and antihyperlipidemic activity of ethyl acetate fraction of Rhododendron arboreum Smith flowers in streptozotocin induced diabetic rats and its role in regulating carbohydrate metabolism. *Asian Pacific Journal of Tropical Biomedicine*. 2(9): 696-701.
 24. Kumar P, Singh R, Bansal P, Balapure A.2012. R. Arboreum Flower And Leaf Extracts: RP-HPTLC Screening, Isolation, Characterization and Biological Activity. *Rasayan J.Chem*. 5(2):165-172.
 25. Thangaraj V.2013.Hypolipidemic effect of Rhododendron arboretum Sm. linn flower juice in experimentally induced hypercholestermic rabbits. *Int J Pharm Biomed Res*. 4(1): 46-49.
 26. Muhammad N, Ali S, Qaisar M, Ali G.2013. Antifungal activity of bioactive constituents and bark extracts of Rhododendron arboretum. *Bangladesh J Pharmacol*. 8: 218-222
 27. Goyal P, Chauhan A.2009.Laboratory Evaluation of crude extract of Cinnamomum tamala for Potential Antibacterial Activity. *Electronic journal of Biology*. 5(4): 75-79.
 28. Vardar Unlu G, Candan, Sokmen A, Domez Eand Tepe B. 2003. Antimicrobial activity of essential oil from C.tamala (Lauraceae). *J Agric Food Chem*. 51:63-67.
 29. Pandey A.K, MishraA.k.2012. Antifungal and antioxidative Potential of Oil and extracts Derived from leaves of Indian Spice Plant Cinnamomum tamala. *Cell. Mol. Biol*.58(1): 142-147.
 30. Palanisamy P, Srinath K.R, Chowdary P.2011. Evaluation of Anti-oxidant and Anti-diabetic activities of *Cinnamomum tamala* Linn leaves in Streptozotocin induced diabetic rats. *Int. Research Journal of Pharmacy*.2(12): 157-162.
 31. Prabuseenivasan S, Ignacimuthu S, Subash P.2007.Cinnamaldehyde-A potential antidiabetic agent. *Science Direct* .14(1): 15–22.
 32. Roux GF, Perrier J, Forest E, Mouren GM, Puigserver A, and Santimone M.1998. Screening of bark of

- Cinnamomum tamala (Lauraceae) by using α -amylase inhibition assay for anti-diabetic activity. *Biochim Biophys Acta*.10-20
33. Chakraborty U, Das H.2010.Antidiabetic and Antioxidant Activities of Cinnamomum tamala Leaf Extracts in Stz-Treated Diabetic Rats. *Global Journal of Biotechnology & Biochemistry*. 5 (1): 12-18.
 34. Verma N, Singh A, Gupta A, Sahu PK, Rao C.2011. Antidiarrheal potential of standardized extract of Rhododendron arboretum Smith flowers in experimental animals. *Indian Journal of Pharmacology*. 43(6):689-693.
 35. Osawa T, Namaki M.1983,. A novel type antioxidant isolated from leaf wax of Eucalyptus leaves. *Agric Biol Chem.*, Vol 45:735-739.
 36. Miller NJ, Rice- Evans CA.1997. Factors affecting the antioxidant activity determined by ABTS+ radical cation assay. *Free Radic Res*. Vol 26: 195-199.
 37. Devi S.L, Kannappan S, Anuradha C.V.2007. Evaluation of *in-vitro* antioxidant activity of Indian bay Leaf, Cinnamomum tamala using rat brain synaptosomes as model system. *Indian Journal of Experimental biology*. Vol 45:778-784.
 38. Pandey A.K, Mishra A.K. 2012.Antifungal and antioxidative Potential of Oil and extracts Derived from leaves of Indian Spice Plant Cinnamomum tamala.*Cell.Mol.Biol*. Vol 58(1) : 142-147.
 39. R. Swamidasan, Prakash T, Divakar G, Kamalesh D.R, Dixit P, Chandrasekar S.B. 2008. Adaptogenic activity of ethanolic extract of leaves of rhododendron arboretum in mice and rats. *Indian J Pharmacol*. Vol 40(2): 402.
 40. P. Dhan, U. Garima, Singh B.N, D. Ruchi, K. Sandeep & Singh K.K. 2007. Free radical scavenging activities of Himalayan Rhododendrons. *Curr Sci*. Vol 92: 526-32.
 41. Shah M, Panchal M. 2010. Ethanopharmacological Properties of Cinnamomum Tamala – A REVIEW.Article-025. *International journal of Pharmaceutical sci. review and research* .5(3).
 42. Soumya S, Sanjayakumar YR, Salinichandran K. 2012. Anti-inflammatory, Analgesic and antipyretic activity of methanolic extract of Cinnamomum tamala in experimental animal models. *Int. Journal of Bioassays*. Vol 1(9): 1-4
 43. Verma N, Singh A.P, Amresh G, P.K Rao C.V. 2011. Protective effect of ethyl acetate fraction of rhododendron arboretum flowers against carbon tetrachloride-induced hepatotoxicity in experimental models. *Indian Journal of Pharmacology*. 43(3): 291-295.
 44. Anderson J. 2014. The healing potential of cinnamon. *Natural News and scientific Discoveries*.
 45. Verma N, Singh A.P, Sahu PK. Protective effect of ethyl acetate fraction of Rhododendron arboreum flowers against carbon tetrachloride-induced hepatotoxicity in experimental models. *Indian Journal of Pharmacology*.2011, 43(3):291-295.
 46. Mishra AK, Mishra A, Kehri HK, Sharma B, Pandey AK. 2009. Inhibitory activity of Indian spice plant Cinnamomum zeylanicum extracts against Alternaria solani and Curvularia lunata, the pathogenic dematiaceous moulds. *Bio Med Central*. 8-9.
 47. Tiwari P, Kumar B, Kaur G.2011.Phytochemical screening and Extraction: A review. *Internationale Pharmaceutica Scientia*.1(1).
 48. http://shodhganga.inflibnet.ac.in/bitstream/10603/11645/12/12_chapter%203.pdf

49. Sadasivam S, Manickam A.1996. Biochemical Methods, 2nd edition. New Delhi New Age International (P) Ltd: 192-93.
50. Kiruba S, Mahesh M, Nisha SR, Miller P. 2011. Phytochemical analysis of flower extracts of *Rhododendron arboretum* Sm. ssp. *nilagiricum* Tagg. *Asian Pacific J Tropical Biomedicine*.
51. Chanda N, Shukla R, Zambre A, Mekapothula S, Kulkarni RR, Katti K.2011. An effective strategy for the synthesis of biocompatible gold nanoparticles using cinnamon phytochemicals for phantom CT imaging and photoacoustic detection of cancerous cells. *Pub med.* 28(2): 279-91.

Table 1. Some examples of Chemical compounds with content percentage in *Cinnamomum tamala*(Nees and Eberm)

Plant Parts	Compound	Percentage
Leaves ¹²		
	β - caryophyllene	0.50%
	α - pinene	2.25%
	β - pinene	0.50%
	camphene	0.34%
	Linalool	1.20%
	Cinnamaldehyde	0.20%
	Eugenol	68.10%
	p-cymene	4%
Bark	Cinnamic aldehyde	70-80%
Roots	Eugenol	-
	Safrole	-
	Benzaldehyde	-
	Terpine	-

Table 2. Different compounds isolated from *Rhododendron arboreum* Smith

Plant parts	Compound present	Percentage
Wood (Stem) ¹³	Ash content	0.75%
	Extractives	4.39%
	Homocelluloses	69.82%
	Hemicelluloses	22%
	Alpha-cellulose	47.98%
	Pentosans	16.84%
	Lignins	24.88%
Leaves ¹⁴		Molecular formula
	Ericolin(arbutin)	C ₁₂ H ₁₆ O ₇
	Ursolic acid	C ₃₀ H ₄₈ O ₄
	Alpha-amyrin	C ₃₀ H ₅₀ O
	Epifriedelinol	C ₃₀ H ₅₂ O
Flowers ¹⁴	Triterpenoids(Campanulin, quercetin & hyperoside)	C ₂₁ H ₂₀ O ₁₂
	Quercetin-3-rhamnoside	C ₂₁ H ₂₀ O ₁₁
Phenolic compound	Quecetin	C ₁₅ H ₁₀ O ₇
	Rutin	C ₂₇ H ₃₀ O ₁₆
	Coumaric acid	C ₉ H ₈ O ₃

Table 3. Various Activities reported on *Cinnamomum tamala* (Nees and Eberm) and *Rhododendron arboreum* Smith

Species	Part utilized	Activity
<i>C. tamala</i> (Nees& Eberm)	Leaves(ethanolic extract)	Hypoglycaemic activity ¹⁵ .
<i>C. tamal</i> (Nees& Eberm)	Leaves(aqueous extract)	Antidiabetic and anti-oxidant activities ¹⁶ .
<i>C. tamala</i> (Nees& Eberm)	Leaves (ethanol extract)	Antidiabetic ¹⁷ .
<i>C. tamala</i> (Nees& Eberm))	Leaves (methanolic extract)	Analgesic, anti- inflammatory and antipyretic activity ¹⁸ .
<i>C. tamala</i> (Nees& Eberm)	Leaves(ethanolic extract)	Reno-protective properties ¹⁹ .
<i>Rhododendron arboreum</i> (Smith)	Leaves(flavonoids)	Anti-oxidant activity ²⁰ .
<i>Rhododendron arboreum</i> (Smith)	Flower(ethyl acetate fraction)	Anti-inflammatory and anti-nociceptive activity ²¹ .
<i>Rhododendron arboreum</i> (Smith)	Flower(ethyl acetate fraction)	Antidiarrheal, hepatoprotective potential ²⁰ , Antihyperglycemic & antihyperlipidemic activity ²¹ .
<i>Rhododendron arboreum</i> (Smith)	Flower & leaves(alcoholic extract)	Anticancer activity ²⁴ .
<i>Rhododendron arboreum</i> (Smith)	Flower juice	Hypolipidemic effect ²⁵ .
<i>Rhododendron arboreum</i> (Smith)	Bark(n-hexane, chloroform, ethyl acetate, methanol extract)	Antifungal activity ²⁶ .

Table 4. Preliminary test

S. No	Experiment	Observation	Inference
1.	20 mg powder was dissolved in 2 ml distilled water and filtered then 2 ml FeCl ₃ was added to the filtrate ⁴⁶ .	Blue – black precipitate	Indicates the presence of Tannin
2.	20 mg extract was dissolved in 2 ml distilled water and filtered. To the filtrate, 2–4 drops of 1% HCl was added and steam was passed through it. To the 1 ml of this solution 6 drops of Wagner's reagent was added ⁴⁶ .	Brownish red ppt	Indicates the presence of Alkaloids
3.	0.5ml of filtrate obtained from alkaloid test 5ml distilled water was added(Wall et al,1954)	Honey comb froth persistence for 15-30 minutes	Indicates the presence of Saponins
4.	20 mg extract were dissolved in 10 ml ethanol and filtered. 0.5 ml conc. HCl and magnesium ribbon were added to 2 ml filtrate ⁴⁶ .	Pink or magenta red or crimson colour	Indicates presence of Flavonoids
5	Extract were dissolved individually in 5ml distilled water , filtered and the filtrate was tested for the presence of carbohydrate using the procedures described by Harper (1975) and Balbaa (1986).		
a)	Molisch's test: Filtrates were treated with 2 drops of alcoholic α-naphthol solution in test tube.	Bluish violet ring at the junction	Indicates the presence of carbohydrates and or glycosides .
b)	Fehling's test: To 5ml filtrates were treated with 5ml Fehling's(A & B) and heated	Formation of red precipitate	Indicates presence of reducing sugars .
c)	Benedict's test: To 1ml of filtrate,5ml of Benedict's reagent were added and heated	Red precipitate	Indicates the presence of reducing sugar .
6)	Glycosides test: Extract were hydrolysed with dil.HCl, and subjected to glycoside test		
a)	Legal's Test: Extract were treated with sodium nitroprusside in sodium hydroxide and pyridine.	Formation of pink to blood red colour	Presence of cardiac glycosides .
7. a)	Phytosterols test: Salkowski's test: Extracts were treated with chloroform and filtered. To the filtrates few drops	Golden yellow colour	Indicates presence of triterpenes

	of conc. Sulphuric acid was added.		
b)	Libermann Burchard's test: Extract were treated with chloroform and filtered. To the filtrate few drops of acetic anhydride were added followed by conc. Sulphuric acid ⁴⁷ .	Formation of brown ring at the junction	Indicates the presence of phytosterol .
8)	To 1ml extract 10 ml chloroform were added followed by addition 10 ml of conc. Sulphuric acid.	Change of color from violet to blue /green	Presence of steroids
9)	To 1gm of sample in test tube covered with filter paper moistened with dil. NaOH and heated on water bath for few minutes. (El-Tawil , 1983).	Yellow fluorescence on filter paper observed under U.V light	Indicates the presence of coumarins :
10.	Plant extract were dissolved in 10 ml distilled water, filtered and subjected to various test.		
a)	Millon's Test- Few drops of Millon's reagent were added to 2ml filtrate	White precipitate	Indicate presence of Protein
b)	Biuret Test- Drop of 2%CuSO ₄ were added to 2ml filtrate followed by 1ml 95% ethanol and excess potassium hydroxide solution 60%	Appearance of pink colour in ethanol layer	Indicates presence of Protein ⁴⁸
11)	Methanolic extract of plant were dissolved in distilled water and 0.5ml Folin-ciocal teau reagent was added followed by 2ml 20% Na ₂ CO ₃ solution.	Formation of bluish colour	Indicates the presence of Phenol ⁴⁹ .

Table 5. Preliminary screening of phytochemicals present in *Cinnamomum tamala* (Nees and Eberm)⁴⁸

Phytochemical constituents	Ethanol	chloroform	Ethyl acetate	Methanol	water
Flavonoids	+	-	-	+++	-
Terpenoids	-	++	+	-	-
Steroids	-	++	++	-	-
Alkaloids	++	+	++	-	-
Tannins	-	++	-	-	-
Glycosides	+	-	-	+++	-
Carbohydrates	-	-	-	+	++
Saponins	-	+	-	-	-
Protein	-	-	-	+	+

(--)= Not present;

(+)= Present but low in abundance;

(++)= Present in moderate amount;

(+++)= Present in high amount.

Table 6. Preliminary screening of phytochemicals obtained in *Rhododendron arboreum* Smith⁵⁰

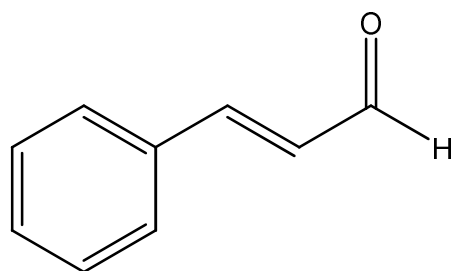
Phytochemical constituents	Acetone	benzene	chloroform	Ethanol	Petroleum ether	water
Alkaloids	-	-	-	-	-	-
Phenol	+++	++	+	+++	+	++
Flavonoids	-	-	-	-	-	-
Saponins	-	+++	+++	-	+++	-
Protein	-	-	+	+	-	-
Steroids	-	+	-	+++	-	-
Tannins	+++	-	-	+++	-	+++
Xanthoprotein	+++	-	-	-	-	-
Carboxylic acid	-	-	-	-	-	-
Coumarins	+++	-	+	-	+	-
Carbohydrates	-	+	+++	+	+	-

(-)= absent;

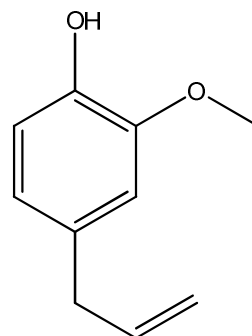
(+)= low;

(++)= average;

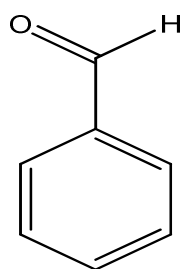
(+++)= high.



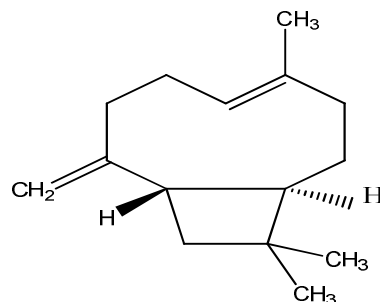
cinnamaldehyde



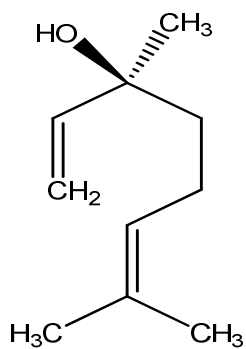
Eugenol



Benzaldehyde



Caryophyllene



Linalool

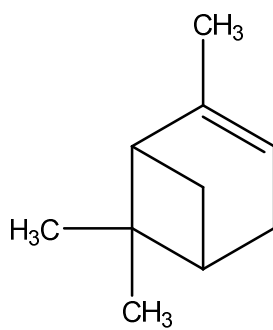
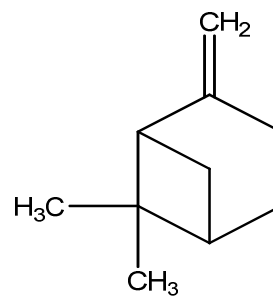
 α -pinene β -pinene

Figure 1. Chemical structures of some compounds found in different parts of *Cinnamomum tamla*(Nees and Eberm)

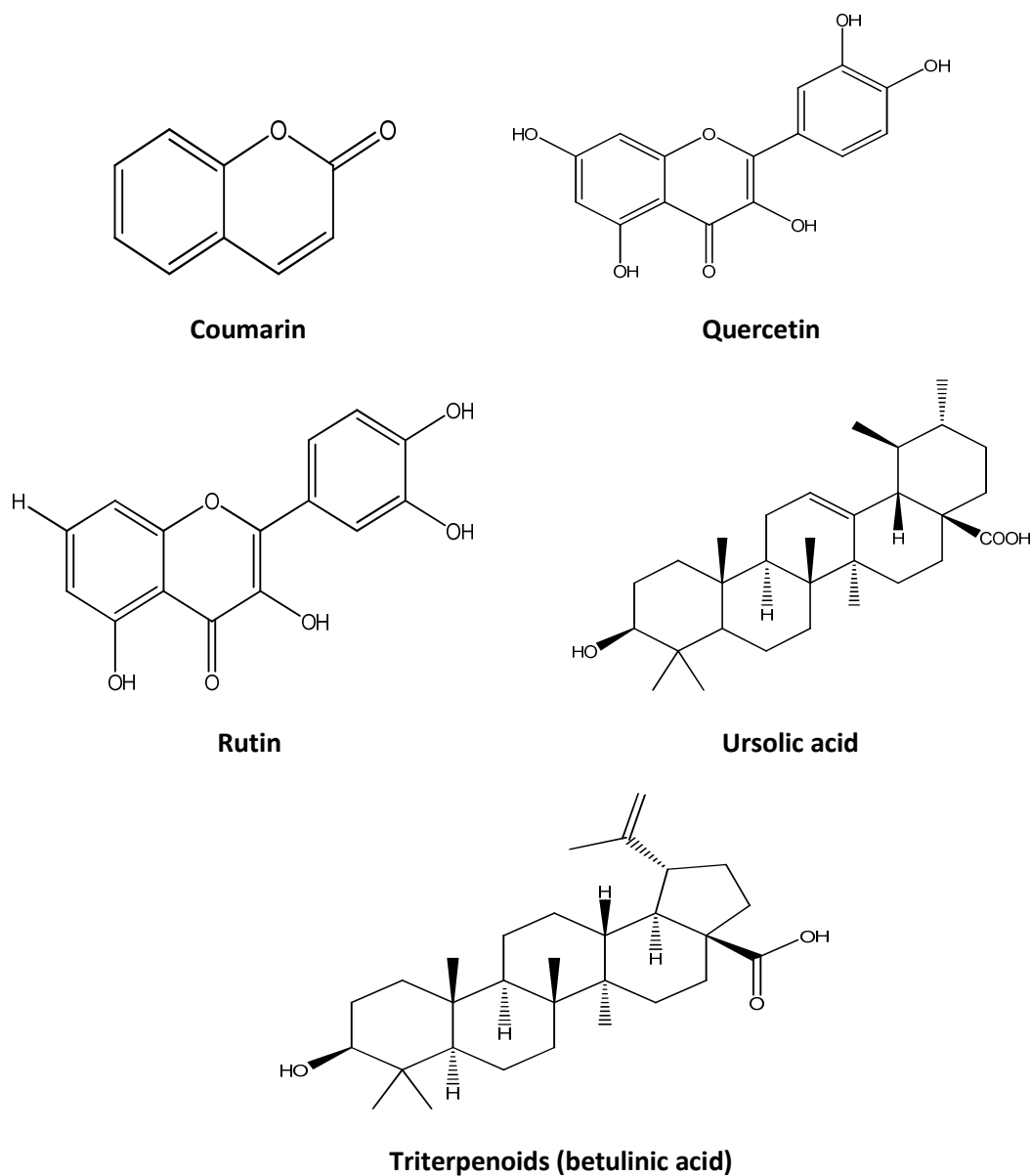


Figure 2. Chemical Structures of some isolated compounds of *Rhododendron arboreum* Smith