Phytochemical Analysis and Anti-Microbial Activities of Rosa Indica Collected from Kohat Pakistan

Rumana Saeed^{*1}, Hameed-Ur-Rehman¹, Shaiq Ali¹, Hidayat Ullah¹, Mukhtar Ullah², Rohullah¹, Saeed Hassan⁴, Farhan¹, Shehzad Ahmed³ and Shomaila Akhwan¹

¹Department of Chemistry, Kohat University of Science & Technology (KUST), Kohat-26000 Khyber Pakhtunkhwa, Pakistan

²Department of Microbiology, Kohat University of Science & Technology (KUST), Kohat-26000 Khyber Pakhtunkhwa, Pakistan

³Department of Physics, International Islamic University Islamabad, 44000 Punjab, Pakistan

⁴Department of Biotecnology, Kohat University of Science & Technology (KUST), Kohat-26000 Khyber Pakhtunkhwa, Pakistan

ABSTRACT

Objectives: The aim of the research work was to assess out the antimicrobial activity and phytochemicals in medicinal plant, Rosa indica.

Methods: For phytochemical screening different reagents were used while antimicrobial i.e. antibacterial and antifungal activities were checked by employing various bacterial and fungal strains, using nutrient agar at 37°C and agar tube dilution method, the stock solutions of crude and other three fractions were prepared in DMSO i.e. in 4mg/ 1ml ratio respectively.

Results: All solvent soluble fractions of Rosa Indica showed superlative activity against Streptococcus pneumonia, Shigella flexneri, Proteus mirabilis, Pneumonia, Salmonella typhimurium, Vibia cholera, E. coli, Pneumonia aeruginosa but no activity was recorded against Proteus mirabilis by hexane and chloroform fraction. Similarly the crude extract exhibited complete anti-fungal activity against all fungal strains. Chloroform fraction was active in all fungal strains except Aspergillus fumigatus. The n-hexane fraction was active against Aspergillus fumigatus and Aspergillus Niger while n-butanol fractions exhibited best Antifungal activity against all fungal strains except Aspergillus Niger. The Rosa indica show best phytochemical results.

Conclusion: The above mentioned result recommends that Rosa Indica have excellent anti-fungal as well as anti-bacterial properties and can be used for disease management and therapy. Beside this Rosa Indica have also gain important in scientific ethno-botanical studies because of their phyto-chemicals..

Address for Correspondence

Department of Chemistry, Kohat University of Science & Technology (KUST), Kohat-26000 Khyber Pakhtunkhwa, Pakistan **E-mail:** <u>Saeedrumana</u> @gmail.com Keywords: Rosa indica, Phytochemicals, Antibacterial, Antifungal.

INTRODUCTION

Therapeutic plants are generously available throughout the world. Therapeutic plants are now much more motivated than ever before because they are capable of producing numerous benefits to community indeed to mankind, particularly inside the type of medication. These naturally occurring compounds are believed to form the building blocks of modern-day prescription medication as we recognize today¹⁻⁶.

has influenced cultures Rose artistically, economically, clinically. scientifically, psychologically and religiously since manhood could fragrance and appreciates its essences. The presence of anthocyanins in rose petals in high concentration give assurance to these signs because anthocyanins are recognized for their ability of having anti-bacterial, antiinflammatory and anti-oxidant activity. Beside that it also provides strength to plant's vascular system and reduce blood platelet stickiness⁷⁻¹². All members of Rosacea family have been cultivated worldwide. There are approximately 10,000 species of rose, including the most fragrant R. damascena, R. gallica, Rosa centifolia, R. muscatta, R. indica, R. rubiginosa and R. rugosa¹³. Amongst all, R. damascena Mill is one of the most popular rose specie, it has been commonly employed for the extraction of rose essential oil¹⁴⁻¹⁷

Literature survey reveals that several studies were published on anti-fungal, antibacterial and anti-oxidant activities of roses¹⁸. The physiological functions of *Rosacea* may be partially due to the abundance of flavonoids and phenolic compounds, also known as bioactive agents ¹⁹. Phenolic compounds retaina wide range of bio-chemical activities, such as freeradical scavengers, anti-bacterial, antioxidants²⁰⁻²², anti-inflammatory²³, antimutagenic²⁴ and anticancer²⁵. Like phenolic compounds, flavonoids also show antioxidant activities, they were observed to protect DNA against oxidative damages^{26,27}. Beside that they possess strong resistance against UV radiation having wavelength of 254 nm and potentially it is believed that they could be used in anti-solar creams²⁸.

In a nutshell medicinal plants are the most important source for varieties of drugs because phytochemicals present in these plants are more precise, environment friendly. easily decomposable. Photochemical bid exclusive platform for biological functions and structural diversity which are essential for drugs discovery, their biological properties help in reducing the risk of various chronic disease²⁹⁻³¹. An extensive range of phytochemicals exists within Rosacea^{32,33}.

One of the main problem that concerns open wellbeing, is usuallv microbial opposition in opposition to antibiotics^{34,35}. Consequently, researchers are hectic in screening natural resources for antimicrobial agents vet undiscovered³⁶⁻³⁸. Extract of Rosa. damascena were found to antimicrobial activities³⁹. This have acknowledgement that a lot of contagious pathogenic creatures are building resistance (s) from the relevant medications rapidly offers necessitated to search for new sources of anti-microbial compounds⁴⁰. That's why; the intention of this study was to evaluate the antimicrobial activities and phytochemical screening of rose.

MATERIALS AND METHODS

Collection of plant

Rose flowers were collected early morning from Bannu, Khyber Pakhtunkhwa, Pakistan. The sample rose flowers were collected from the plants being kept in the gardens. The rose flowers were collected during February to March in 2014.

Preparation of plant extract with ethanol

After collection rose petals dried under shade for a couple of hours and were then extracted with ethanol. The extract obtained was dried using a Rota-evaporator at 60°C. The extract was further dried at room temperature. After complete drying, it was fractionated consequently with hexane, chloroform, ethyl-acetate and finally with butanol. All solvent fractions were allowed to dry under room temperature.

Bacterial strains and growth conditions

Bacterial strains used in the research were obtained from the microbial collection, from the Department of Microbiology, KUST, Kohat (Table 1). Bacteria were subcultured from stock maintained in nutrient agar at 37 °C.

Determination of antifungal activity

Using Agar tube dilution method, the stock solutions of crude and other three fractions were prepared in DMSO i.e. in 4mg/1ml ratio. (1 ml from each dilution was added to sterilize each test tube, and then 9 ml of nutrient agar (Sterilized media) was added to the test tubes.). Small piece of previously grown fungus was added to each test tube with the help of sterilizing loop. The tubes containing positive and negative control were incubated at 250 °C for a week. After a week the fungal growth in each tube was observed with reference to the positive control⁴¹. The anti-fungal activities of *R. indica* was checked by using four different

fungal strains i.e. A. Flavus, A. Fumi, A. Niger and F. Solani.

*positive control contains fungus, *negative control is without fungus.

Phytochemical analysis

Phytochemical screening of rose extracts were carried out according to the methods described by Arun Joshi, Maya Bhobe and AshmaSattarkar⁴², for the detection of bioactive components like alkaloids, carbohydrates flavonoids and saponins etc.

RESULTS ANS DISCUSSION

Most of the journals provide overview about whether plant extract/fractions possess activity against Gram-negative and Gram-positive bacteria and fungi or not. Nevertheless, not all provide details information about the extent of their activity. The consequence of the study reveals that the polar and non-polar solvent fractions of Rosa Indica were active against the bacterial strains that are often responsible for infections. Anti-bacterial and anti-fungal activities of Rosa Indica are shown intable-1 and table-2. Anti-bacterial activities of Rosa Indica were performed against.

Streptococcus pneumoniae, Shigella flexneri, Proteus mirabilis, Pneumonia, Salmonella typhimurium, Vibia cholera, *E. coli*, Pneumonia aeruginosa strains whereas anti-fungal activity was performed against Aspergillus flavus, Fusarium solani, Aspergillus niger and Aspergillus fumigatus strains.

All solvent soluble fractions of *Rosa Indica* showed excellent activity against Streptococcus pneumoniae, Shigella flexneri, Proteus mirabilis, Pneumonia, Salmonella typhimurium, Vibia cholera, *E. coli*, Pneumonia aeruginosa but no result was recorded against Proteus mirabilis by hexane and chloroform fraction. The crude

and butanol fraction showed highest activity against all strains of bacteria. Similarly comprehensive anti-fungal activity was shown by crude against all fungal strains. Chloroform fraction showed zero percent activity against Aspergillus fumigatus. The n-hexane fraction was active against Aspergillus fumigatus and Aspergillus niger while n-butanol fractions exhibited best Antifungal activity against all fungal strains except Aspergillus niger. There are so many reasons to put a question, that why Rosa Indica show anti-fungal and anti-bacterial activities. The most effective reason is the presence of various phytochemicals like alkaloids, phenolic acids, flavonoids, tannins, and volatile oils. Rosa Indica exhibited best anti-fungal and anti-bacterial activity against such bacterial and fungal strains that are often present in our environment particularly in our food and cause serious infections. It can be used as antimicrobial, for the treatment of different bacterial and fungal diseases and infections.

CONCLUSION

The above mentioned result recommends that Rosa Indica have excellent anti-fungal as well as anti-bacterial properties and can be used for disease management and therapy. It may also be used or suggested as a new source in antimicrobial medications and for disease therapy. Beside this Rosa Indica have also gain important in scientific ethno-botanical studies because of their phytochemicals. Further research is necessary for isolation and characterization of effective elements of the extracts and also to elucidate the mechanism of their action.

ACKNOWLEDGEMENTS

Authors are grateful to the faculty members especially to the Chairman Dr. Murad Ali Khan for their appreciativeness and making the facilities available for the research, department of chemistry, Kohat University of science and technology, Kohat, Pakistan.

REFERENCES

- 1. S.M.M. Shah. Evaluation of Phytochemicals and Antimicrobial Activity of White and Blue Capitulum and Whole Plant of Silybum Marianum, *World Applied Sciences Journal* 2011, 12 (8): 1139-1144.
- 2. Chopra, R.N. and K. Nayar Chopra, 1986. Glossary of Indian medicinal plants.Coancil of Scintific and Industrial research, New Dheli India.
- 3. Ferrence, S.C. and G. Bendersky, 2004. Therapy with saffron and the goddess at thera. *Perspectives in Biol. Med.*, 47: 199-226.
- 4. Owolabi, J., E.K.I. Omogbai and O. Obasuyi, 2007. Antifungal and antibacterial activities of the ethanolicandaque *C. erectus paperous* extract of *Kigeliaafricana* (Bignoniaceae) stem bark. *Afr. J. Biotechnol.*, 6: 882-85.
- 5. Chitravadivu, С., M. Bhoopathi, T. S. Elavazhagan, Javakumar and V. Balakrishnan, 2009. Screening of antimicrobial activity of medicinal plant oils prepared by herbal venders, South India, Middle-East J. Med. Pl. Res., 4: 115-117.
- Kviecinski, M.R., K.B. Felipe, T. Schoenfelder, L.P. de Lemos Wiese, M.H. Rossi, E. Gonçalez, J.D. Felicio, D.W. Filho and R.C. Pedrosa, 2008. Study of the antitumor potential of Bidenspilosa (Asteraceae) used in Brazilian folk medicine, *J. Ethnopharmacol.*, 117: 69-75.
- N. B. Hirulkar and Mona Agrawal, Antimicrobial Activity of Rose petals Extract Against Some Pathogenic Bacteria", *International Journal of Pharmaceutical & Biological Archives* 2010; 1(5):478-484
- Akhmadieva AK, Zaichkina SI, Ruzieva RK, Ganassi EE. 1993. [The protective action of a natural preparation of anthocyanin (pelargonidin-3, 5-diglucoside)]. *Radio biologiia*. 1993 May-Jun; 33(3): 433-5. Russian.
- 9. M. Shohayeb, S. Abdel-Hameed, S.A. Bazaid and I. Maghrabi, Antibacterial and Antifungal

Activity of Rosa damascena MILL. Essential Oil, Different Extracts of Rose Petals. *Global Journal of Pharmacology* 8 (1): 01-07, 2014.

- Hirulkar NB and Mona A., *International J. of Pharmaceutical & Biological Archives*, 2010, 1(5):478-484.
- 11. Joo SS, Kim Yun B and Lee DI., *Plant Pathol. J*, 2010, 26(1): 57-62.
- Vinokur Y, Victor R, Natalie R, Genady G, Batia H, Nakdimon U And Haya F., J. of Food Science, 2006, 71(1):42-47.
- 13. Özkan G, Sağdıç O, Göktürk-Baydar N, Baydar H (2004) Antioxidant and antibacterial activities of *Rosa damascena* flower extracts. *Food Sci Technol Int* 10:277– 281.
- A.D. Kashani, I. Rasooli, M.B. Rezaee, P. Owlia, Antioxidative Properties and Toxicity of White Rose Extract, *Iranian journal of toxicity*, 415-425, Volume 5, No.1&2, Spring & Summer 2011.
- 15. Rusanov K, Kovacheva N, Vosman B, Zhang L, Rajapakse S, Atanassov A, Atanassov I. Microsatellite analysis of Rosa damascena Mill. Accessions reveals genetic similarity between genotypes used for rose oil production and old Damask rose varieties. *Theor Appl Genet* 2005; 111(4):804-9.
- 16. Rangaha MK. Rose Rosa damascena 'Trigintipetala. *Crop Food Res Broad Sheet* 2001; 29.
- 17. Das, A., S. Bhui and D. Chakraborty, 2012. Growth Behavior of Rose Plants in Low Cost Hydroponics Culture, *J. Hort. Sci. and Ornamen. Plants*, 4: 01-06.
- Basim E, Basim H (2003) Antibacterial activity of *Rosa damascena* essential oil. *Fitoterapia* 74:394–396.
- 19. Wen D, Li C, Di H, Liao Y, Liu H. A universal HPLC method for the determination of phenolic acids in compound herbal medicines. *J Agric Food Chem* 2005; 53(17):6624-9.
- Leenen R, Roodenburg AJC, Tijburg LBM, Wiseman SA. A single dose of tea with or without milk increases plasma antioxidant activity in humans. *European Journal of Clinical Nutrition* 2000; 54(1):87-92.
- 21. Ng TB, Liu F, Wang ZT. Antioxidative activity of natural products from plants. *Life Sciences* 2000; 66(8):709-23.

- 22. Jafari M, Zarban A, Pham S, Wang T. Rosa damascena decreased mortality in adult Drosophila. *J Med Food* 2008; 11(1):9-13.
- Crespo ME, Ivez J, Cruz T, Ocete MA, Zarzuelo A. Anti-inflammatory activity of diosmin and hesperidin in rat colitis induced by TNBS. *Planta Medica* 1999; 65(7):651-3.
- Miyazawa M, Okuno Y, Nakamura SI, Kosaka H. Antimutagenic activity of flavonoids from Pogostemon cablin. J Agric Food Chem 2000; 48(3):642-7.
- 25. Ren W, Qiao Z, Wang H, Zhu L, Zhang L. Flavonoids: Promising anticancer agents. *Medicinal Research Reviews* 2003; 23(4):519-34.
- 26. Pawlak, K., W. Bylka, B. Jazurek, I. Matlawska, M. Sikorska, H. Manikowski and G. Bialek-Bylka, 2010. Antioxidant activity of flavonoids of different polarity, assayed by modified abtscation radical decolorization and epr technique, *Acta Biol. Cracoviensia Series Botanica*, 52: 97-104.
- 27. Kalim, M.D., D. Bhattacharyya and A. Banerjee, 2010. Chattopadhyay S. Oxidative DNA damage preventive activity and antioxidant potential of plants used in Unani system of medicine, *BMC Complement Altern. Med.*, 16: 10:77.
- Tabrizi, H., S.A. Mortazavi and M. Kamalinejad, 2003. An *in vitro* evaluation of various *Rosa damascene* flower extracts as a natural antisolar agent, *Int. J. Cosmet. Sci.*, 25: 259-265.
- 29. J.A. Khan and S. Tewari, A Study on Antibacterial Properties of Rosa indica against Various Pathogens, *Asian Journal of Plant Science and Research*, 2011, 1 (1): 22-30
- 30. Verpoorte R, Contin A and Memelink, J., *Phytochemistry Reviews*, 2002, 1 (1): 13–25.
- 31. Curin Y, Andriantsitohaina R. Polyphenols as potential therapeutical agents against cardiovascular diseases. *Pharmacological Reports* 2005; 57(Suppl.):97-107.
- 32. Olsson ME, Andersson S, Werlemark G, Uggla M, Gustavsson KE. Carotenoids and phenolics in rose hips. *Acta Horticulturae* 2005; 490:249-53.
- Uggla M, Gustavsson KE, Olsson ME, Nybom H. Changes in colour and sugar content in rose hips (Rosa dumalis L. and R.

rubiginosa L.) during ripening. *J Hortic Sci Biotechnol* 2005; 80(2):204-8.

- 34. Karlowsky, J.A., A.J. Walkty, H.J. Adam, M.R. Baxter, D.J. Hoban and G.G. Zhanel, 2012. Prevalence of Antimicrobial Resistance among Clinical Isolates of Bacteroidesfragilis Group in Canada in 2010-2011: CANWARD Surveillance Study. *Antimicrob. Agents Chemother.*, 56: 1247-1252.
- 35. Gayathri, V.R., P. Perumal, A. Pazhanimuthu and B. Prakash, 2013. Epidemiology and Molecular Variations in Methicillin Resistant Staphylococcus aureusIsolated from Different Clinical Samples of Private Hospitals of Salem District, India, *Global J. Pharmacol.*, 7: 81-86.
- Srivastava, J., J. Lambert and N. Vietmeyer, 1996 Medicinal plants: an expanding role in development, World Bank Technical Paper no, pp: 320.
- 37. Ahmed, A., A.G. Murtaza, Z.I.A.T. Akhter and F.A. Minhas, 2013. Ethnobotanical study of some highly medicinal important wild plants of Leepa Valley District Muzaffarabad, *World Appl. Scie. J.*, 22: 1760-1765.

- 38. Mahesh, B. and S. Satish, 2008. Antimicrobial activity of some important medicinal plant against plant and human pathogens. *World J. Agricul. Sci.*, 4: 839-843.
- Ulusoy, S., G. Bo gelmez-Tinaz .H. Seçilmi -Canbay, 2009. Tocopherol, carotene phenolic contents and antibacterial properties of essential oil, hydrosol and absolute, *Curr. Microbiol.* 59: 554-558.
- Williams, D. H., Stone M. J., Houck P. R. and Rahman S. K., *J. Nat. Prod.*, 1989, 52, 1189– 1208.
- 41. BushraImtiaz, Fozia, Abdul Waheed, Ali Rehman, HussainUllah, Hamid Iqbal, Abdul Wahab, Mamoona Almas, Ijaz Ahmad, "Antimicrobial Activity of Malva Neglecta and Nasturtium Microphyllum" 3(6), Nov – Dec 2012.
- 42. A. Joshi, M. Bhobe and A. Sattarkar, Physicochemical and Phytochemical Investigation of the roots of Grewia Microcoslinn., *American Journal of Pharmacy & Health Research* 2013. Volume 1, Issue 4, 54-65.

S. No	Bacterial Strains used	MTCC code	Crude	Hexane	Chloroform	Butanol	1MP
1	Streptococcus pneumonia	6305	14	14	11	10	49
2	Shigella flexneri	12022	11	13	12	10	52
3	Proteus mirabilis	12453	13	0	0	15	42
4	Pneumonia	-	22	11	9	26	29
5	Salmonella typhimurium	14028	16	12	9	14	26
6	Vibia cholera	9459	24	12	7	28	27
7	E. coli	25922	27	11	9	20	19
8	Pneumonia aeruginosa	27853	23	10	10	28	26

Table 1. Showing anti-bacterial activity of rose fractions against different strains

S. No	Phytochemical scr	Inference	
1	Alkaloids		
a.	Mayer's test	+	Presence
b.	Wagner's test	+++	Presence
с.	Hager's test	++	Presence
2	Carbohydrate		
a.	Molish's test	+++	Presence
b.	Benedict's test	++	Presence
с.	Fehling's test	+++	Presence
3	Flavanoids		
a.	Lead acetate test	++	Presence
4	Triterpenoids &St		
a.	Salkowshi test	++	Presence
5	Phenolic compounds	+++	Presence
6	Resins +		Presence
7	Proteins		
a.	Biuret's test	-	Absence
b.	Millen's test	-	Absence
8	Glycosides		
a.	Legal's test	++	Presence
b.	Liebermann's test	+	Presence
9	Saponins		
a.	Foam test	-	Absence

Table 2. Phytochemical screening	
----------------------------------	--

 Table 3. Anti-fungal activity of rose fractions

Fractions	A.Fumi	A.niger	A.Flavos	F.Solani
Curde	+	+	+	+
n-Butanol	+	-	+	+
n-Hexane	+	+	-	-
Chloroform	-	+	+	+

