

## Histochemical Localization of Crystals and their Taxonomic Significance in Five Species of the Subfamily Papilionoideae

Ojimba Chioma F\*,  
Mbagwu Ferdinand N and  
Inyama Chiemeka N

Department of Plant Science and  
Biotechnology, Imo State University,  
Owerri, Nigeria

### Abstract

An investigation into the histochemical localization of crystals and their taxonomic significance in five species of the subfamily Papilionoideae was carried out. The result showed the presence of rhomboidal type of crystals in the leaf of *M. aboensis* and *L. cyanescens*; stellate type in *C. pubescens* but absent of crystals in *C. retusa* and *I. spicata*. The petiole of *M. aboensis* has stellate, rhomboidal and styloid types of crystals; *I. spicata* has crystal sand type; *C. pubescens* has both stellate and rhomboidal types; *L. cyanescens* has stellate but absence of all these types in *C. retusa*. At stem of these species, *M. aboensis* has stellate, rhomboidal and crystal sand crystals; *I. spicata* has rhomboidal; *C. pubescens* has stellate, rhomboidal and crystal sand crystals; *L. cyanescens* has only crystal sand type while *C. retusa* has none. There is absence of crystals at the root of these species. These results showed that *M. aboensis* and *C. pubescens* are closely related in terms of crystal types while *C. retusa* can be distinguished from the rest of the taxa investigated.

**Keywords:** Taxonomy; Papilionoideae; Identification; Histochemistry

\*Corresponding author: Ojimba Chioma F

✉ chingor01@yahoo.com

Department of Plant Science and  
Biotechnology, Imo State University, Owerri,  
Nigeria.

**Citation:** Chioma FO, Ferdinand NM, Chiemeka NI (2017) Histochemical Localization of Crystals and their Taxonomic Significance in Five Species of the Subfamily Papilionoideae. Am J Ethnomed. Vol. 4 No. 1:4

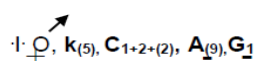
**Received:** March 07, 2017, **Accepted:** March 28, 2017, **Published:** March 31, 2017

### Introduction

Leguminosae is usually divided into three subfamilies: Papilionoideae, Caesalpinioideae and Mimosoideae because of its large size [1]. *Milletta aboensis*, *Crotalaria Retusa*, *Indigofera spicata*, *Centrosema pubescens* and *Lonchocarpus cyanescens* belong to the subfamily Papilionoideae. The subfamily is easily recognized by their characteristic Papilionaceous (butter fly-like) flower [2]. The Papilionoideae contains most of the important leguminous crop species such as soya bean (*Glycine max*), common pea (*Pisum sativum*), chickpea (*Cicer arietinum*), French bean (*Phaseolus vulgaris*), Lentil (*Lens culinaris*) and peanut (*Arachis hypogaea*). Papilionoideae is the largest of the three subfamilies with about 476 genera and 13,855 species [3].

They are herbs, shrubs or trees. Leaves are pinnately or simple with inflorescence racemose and axillary. It is characterized by markedly bilaterally symmetrical, papilionaceous corolla. This means that 5 sepals and five petals are differentiated into one standard petal, two wing petals and a keel formed from two petals that are united along their ventral margins and enclosing the stamens and the pistil. Ovary is superior with marginal placentation. Fruit is a legume and seeds are non endospermic.

### Floral formula of Papilionoideae



Some members of Leguminosae (e.g. the beans) provide man and his livestock with a highly nutritional food resource. Species of many genera (eg *Albizia*, *Azalia*, *Daniella*, *Brechystegia*, *Afromosia*, *Berlinia*) are very good for timber. Species of *Copaifera* and *Lonchocarpus* are sources of dyes. Species of some genera (eg *Pterocarpus*) are sources of leaf vegetables. Many members produce edible oil in large quantities eg. soyabean, groundnut. Many members of legumes yield important fibers, green manures and forages. Species of many genera (eg *Brachystegia*, *Acacia* etc.) are used to produce commercial gum. Some seed species (eg *Brachystegia earycoma*, *Mucuna aurens* are used for thickening soup. Many members of the family are grown as *ornamentals* eg. *Crotalaria* and *Dilonixca esalpinia*. All the tree species are important sources of firewood.

The botanical literature shows that the use of histochemistry in taxonomic conclusions is now a common practice. For example, the occurrence of calcium oxalates crystals in various plant families have been reported by various authors. Edeoga and

Okoli [4], Ayensu [5], Okoli and Green [6], observed the presence of calcium oxalate in the chloroplasts and trichomes of some *Dioscorea* species and suggested that they could be employed for taxonomic purposes. Tomlinson [7], Edeoga and Ogbebor [8] equally reported that calcium oxalate crystals characterize different members of *Commelina* species and Zingiberales and reported their diverse distribution in most parts of the plants. The deployment of histochemistry as a tool for identification and classification of plants has been indicated in various families such as Verbenaceae Mathew and Shah [9], Dioscoreaceae Okoli and Green [10], Edeoga and Okoli, [4] Cucurbitaceae Okoli [6] and Commelinaceae [11]. Other authors that have used ergastic substances for clarity in taxonomy at all hierarchies are Itoh and Karg [12], Burkill [13], Wu and Huang [14] in Moraceae.. Crystals of calcium oxalate which occur in organs and tissues of many plant species are among the most widespread ergastic substance known in Angiosperm [15].

Calcium oxalate crystals exist in different forms such as raphides, bundle of needles, styloids, elongated columnar crystals, druses, spheroidal, aggregates of prismatic crystals and crystals sand [16]. Itoh and Kang [12] found calcium oxalate crystal in the intercellular or middle lamella of the radial walls of the secondary phloem in the Taxodiaceae family. Calcium oxalate crystal occurring in vertical parenchymatous cells include, the raphids of *Morinda citrifolia* and *Hamella patterns*, the solitary crystals of *Psidium guajava* and *Anogeissus acuminata*, the crystal sand of *Cordia subcordata* and *Santivia tomentosa*.

The taxonomic importance of calcium oxalate crystals found in leaves, stems and roots of most taxa lies in the variation in their shape and localization. For example, the rhomboidal crystal of *Aneilema acquinociale* and *A. beniniense* were different from the irregularly shaped oxalate crystals of *A. paludosum* and *A. umbrosum*. From these findings, these taxa could be separated on the basis of crystal shape. The localization of crystals within the root cortex is also important as this region in addition to its storage functions offers protection against desiccation. The persistent association of the crystals with the ground tissues (supportive), cortex (storage) and mesophyll (nutritional) clearly attests to the possible roles of calcium oxalate in the taxonomy and classification of *Aneilema* species.

The use of histochemical attribute of plant in taxonomic conclusions are recently becoming frequently. Edeoga and Ugbo [11], Edeoga and Okoli [4] had suggested that these ergastic substances could have nutritional, mechanical and transport roles in plants.

It is very unfortunate that despite the contributions of the above authors on crystals, little or no information exist on the shape of crystals that are found on the subfamily Papilionioideae hence the need for this work with the objective of finding the type and shape of crystals present in the subfamily. Thus using the crystal type to characterize and identify the species.

## Materials and Methods

### Specimen collection

The five species used in the investigation namely *Milletia aboensis*, *Crotalaria retusa*, *Indigofera spicata*, *Centrosema pubescens*, and *Lonchocarpus cyanescens* were all collected from Imo State. *Milletia aboensis* and *Crotalaria retusa* were collected from Okigwe Local Government Area while *Indigofera spicata*, *Centrosema pubescens*, and *Lonchocarpus cyanescens* were collected from Agriculture farm of Imo State University.

### Specimen identification

The specimens were identified by a taxonomist, Professor S. E. Okeke and were confirmed at Forest Herbarium institute (FHI) Ibadan, Oyo State. Voucher specimens were deposited at Imo State University Herbarium (IMSUH) Owerri. With Voucher numbers IMSUH 89,90,91,92 and 93 respectively.

### Anatomical studies

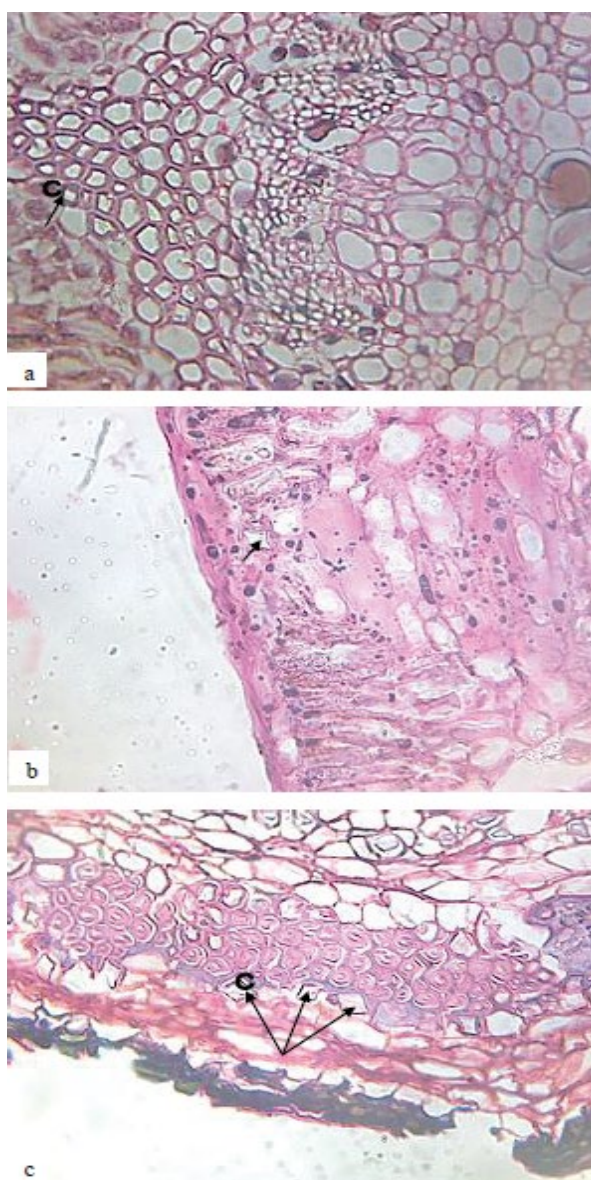
Mature and fresh parts of the leaves, petioles and the stems were collected and sectioned. The cut sections of the fresh, leaves, petioles and stems were done at various portions. For the leaves, cutting was done from the middle portion, the petioles, cutting was at 0.5 cm from the node; while that of stem was 4 cm from the terminal portion. Before sectioning commences, the specimens undergo a pretreatment process referred to as killing and fixing. The aim of this process was to terminate suddenly and permanently, all life processes within the specimens and preserve the cells composing the materials as close to their original conditions as possible [17]. For the pretreatment, the specimens were treated using Formalin Acetic Alcohol (FAA) for 48 hours and then washed thoroughly in distilled water. The specimens were washed in two changes of 30% ethanol and are dehydrated in a graded series of ethanol (30%-50%-70%-95%) for at least 15 minutes in each grade.

They were covered for 3 hours in each of the following solutions containing a ratio of absolute alcohol to pure chloroform (v/v: 3:1, 1:1, 1:3), then pure chloroform. At the stage of pure chloroform, wax pellets (60°C melting point) were added and the wax is changed periodically. The containers of the specimen were transferred to an oven for 2-7 days to remove the chloroform. The contents of the vial were carefully transferred into moulds. The specimens were arranged using a flamed (hot) mounting needle and transferred to a cold water bath where it remains until the wax is sufficiently solid and was later stored in a refrigerator for two days.

A very thin section of 10-20  $\mu$  was made using a Reichert rotary microtome. Ribbon cut sections obtained from the microtome sectioning were placed on clean slides covered with a thin film of Haupt's albumen and allowed to dry. Three or four drops of distilled water were added before mounting. The slides were placed on a hot plate at 40°C for few minutes kept overnight.

The slides were immersed in pure xylene for 2-5 minutes in a solution of xylene and absolute alcohol with a ration of 1:1 (v/v). They were later transferred to another solution of xylene and alcohol graded series (95%, 90%, 70% and 50%) in the ratio of 3:3 (v/v) for few minutes. Drops of safranin were used to stain the section for 5 minutes, washed off with distilled water and then counter stained with alcian blue for 2 minutes and then dehydrate in 50% alcohol, 70%, 80% and 90% alcohol solution respectively and pure xylene at intervals.

Drop of D.P.X mountant were introduced on to the slides enough to cover the length of the sectioned material and then covered with cover slip. The slides containing the mounted sections were placed on a hot plate at 30°C to dry. Photographs of materials sectioned were taken using a Leitz wetzler ortholux digital microscope filled with a camera (Figures 1-3).



**Figure 1** Transverse section of leaves of a; *M. aboensis*; b, *C. pubescens* and c, *L. cyanescens* with arrows indicating calcium oxalate crystals. X400.

## Results

The presence of calcium oxalate crystals in some parts of the five taxa investigated was observed and recorded in **Table 1**. In the mesophyll of the leaves of *M. aboensis* and *L. cyanescens*, rhomboidal shaped crystals was observed and stellate were observed in *C. pubescens* while no crystal was observed in the other two taxa (**Figure 1**). At the petiole stellate, rhomboid and styloid shaped crystals was observed for *M. aboensis*, crystal sand crystal was found in *I. spicata*, in *C. pubescens*, stellate and rhomboid shaped crystals were observed, in *L. cyanescens*, it was stellate shaped crystal while no crystal was observed in *C. retusa* (**Figure 2**). In the stems, *M. aboensis* and *C. pubescens* have three different shapes of crystals: stellate shaped, rhomboid and crystal sand shapes, in *I. spicata*, shaped of crystal was rhomboid, *L. cyanescens* has only crystal sand shapes while no crystal was observed in *C. retusa* (**Figure 3**).

## Discussion

The variation in shape of calcium oxalate crystals is of taxonomic importance and could be used to distinguish and classify different plant species. The leaf anatomy showed that *M. aboensis* and *L. cyanescens* have rhomboidal crystal; for *C. pubescens* has stellate crystal, while no crystal was observed on the other two taxa. This is of systematic importance as it delineates the species. At the petiole, *M. aboensis*, *C. pubescens* and *L. cyanescens* have star-like crystals. In addition, *M. aboensis* and *C. pubescens* have rhomboidal crystals; *M. aboensis* could be separated from the other taxa due to the presence of *styloid* observed. Also, crystal sand can be used to delimitate *I. spicata* from the other taxa while the absence of crystals at *C. retusa* further separates it from the other taxa investigated.

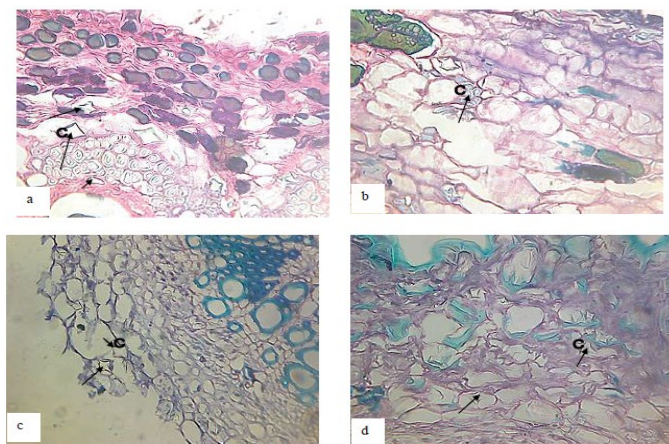
Only rhomboidal shaped crystals were observed in *I. spicata* in the stem anatomy is considered. Also, the presence of crystal sand separates *L. cyanescens* from the other taxa. *M. aboensis* and *C. pubescens* showed close affinity by having star-like rhomboidal and crystal sand crystals in common while *C. retusa* was delineated due to absence of calcium oxalate crystal.. The absence of crystals at the leaf, petiole and stem of *C. retusa* is a distinctive character of this taxon and thus separates it from the other four taxa [18]. The presence of calcium oxalate in four of the taxa investigated showed close phylogenetic relationship between them. While different shapes of the crystal observed could explain why they are placed under different genera or

**Table 1:** Histochemical studies of the five species of the subfamily Papilionoideae investigated.

Part of Plant	<i>M. aboensis</i>	<i>C. retusa</i>	<i>I. spicata</i>	<i>C. pubescens</i>	<i>L. cyanescens</i>
Leaf	Rhomboidal	-	-	Stellate	Rhomboidal
	Stellate,	-	Crystal sand	Stellate,	Stellate,
Petiole	Rhomboidal,				Rhomboidal,
	Styloid				
Stem	Stellate	-	Rhomboidal	Stellate	Crystal sand
	Rhomboidal,			Rhomboidal,	
	Crystal sand			Crystal sand	



species. It was observed that calcium oxalate crystals are present in one form or the other among the taxa considered. Lack of crystals in the vegetative organ distinguished *C. retusa* from the rest of the taxa investigated. *I. spicata* can be separated from others because of the absence of crystal in its leaves. Therefore, this study has strengthened intra specie relationships among the species investigated and the usefulness of histochemistry in delimitation and identification of taxa.

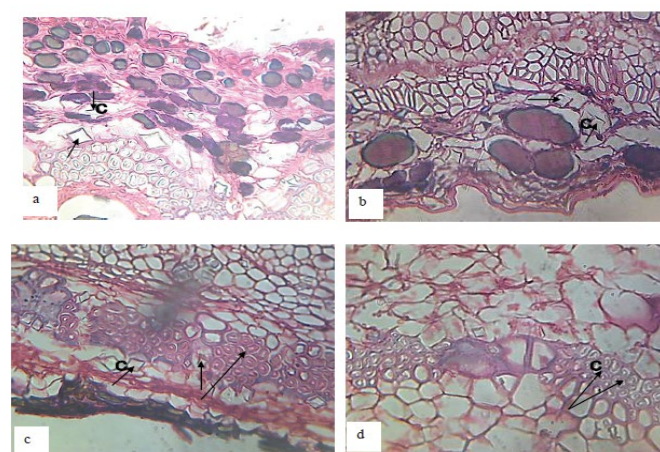


**Figure 2** Transverse section of petiole of a; *M. aboensis*; b, *I. spicata*; c, *C. pubescens* and d, *L. cyanescens*; with arrows indicating calcium oxalate crystals. X400.

species. Authors such as Edeoga and Okoli [4] used shapes of crystals to distinguish species of Dioscorea; Okoli and McEuen also distinguished species of Cucurbitaceae using crystal shapes.

## Conclusion

The data accrued from histochemical study of these species investigated however presented some important characters that could be exploited in improving the characterization of the five



**Figure 3** Transverse section of stem of a; *M. aboensis*; b, *I. spicata*; c, *C. pubescens* and d, *L. cyanescens*; with arrows indicating calcium oxalate crystals. X400.

## References

- 1 Heywood VH, Davis PH (1973) Principles of Angiosperm Taxonomy. Oliver and Boyd, London.
- 2 Dutta AC (2005) Botany for Degree Students. Leth Edition, Oxford University Press.
- 3 Hutchinson J, Dalziel JM (1954) Flora of West Tropical Africa. Vol. I and II. Crown Agenda for Overseas Government and Administration, London.
- 4 Edeoga HO, Okoli BE (1995) Histochemical Studies in the Leaves of some Dioscorea, L (Dioscoreaceae) and the taxonomic importance. Feddes Report 106: 113-120.
- 5 Ayensu ES (1972) Anatomy of the Monocotyledons. VI. Dioscoreales. Oxford Univ. Press, New York.
- 6 Okoli BE (1987) Anatomical Studies of the Leaf and Probract of Telfaira. Hooker (Cucurbitaceae). Feddes Report. 98: 231-239.
- 7 Tomlinson P (1969) Anatomy of Monocotyledons, Commelinales Zingiberales. Clarendon Press. Oxford.
- 8 Edeoga HO, Ogbekor NO (1999) Vegetative Anatomy in some Nigeria Species of Aneilema (Commelinaceae). Acta phytotax Geobot 50: 51-58.
- 9 Matthew L, Shah GL (1984) Crystals and their Taxonomic Significance in some Verbebaeeae. 83: 279-289.
- 10 Okoli BE, Green BO (1987) Histochemical Localization of Calcium Oxalate Crystals in Starch Grains of Yams. (Dioscorea L) 60: 391-394.
- 11 Edeoga HO, Ugbo HM (1997) Histochemical Localization of Calcium Oxalate Crystal in the Leaf Epidermis of some Commelina, L (Commelinaceae) and its bearing on Taxonomy. Acta Phytotax. Geobot, 48: 23-30.
- 12 Itoh T, Kang KD (1993) The occurrence of Calcium Oxalate Crystals in the Cell Walls of the Secondary Phloem of Taxdiaceae. Holzforchung 47: 465-472.
- 13 Burkill HM (1995) The useful Plants on West Tropical Africa. In: Families JL (ed.) Royal Botanic Gardens (2ndedn) Vol. 3, Kew, Richmond, UK.
- 14 Wu-Chichih C, Kuo Hang LL (1997) Calcium Crystals in the Leaves of some Species of Moraceae. Bot Bull. Aca. Sin. 38: 97-104.
- 15 Metcalf CR, Chalk L (1950) Anatomy of the Dicotyledons. Vol 1 and II. Crown agent for Overseas Government and Administration. London.
- 16 Al-Rais AH, Meyers A, Watson L (1971) The Isolation and Properties of Oxalate Crystals from Plants. Ann. Bot 35: 1213-1218.
- 17 Peacork EA (1973) Elementary Microtecheque. London: Edward Arnold.
- 18 Gill LS (1988) Taxonomy of Flowering Plants. Onitsha. Africana Feb. Publishers.