

Therapeutic Apheresis in Transplantology

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Abstract

Background: The problem of organ transplantation is still far from being solved. Until now, to eliminate both acute and chronic rejection, medicines with many side complications are used.

Methods: At the same time, apheresis therapy methods aimed at removing antibodies from the body have not received enough proper and timely application.

Results: Among them plasmapheresis plays the leading role, which can be used to block both acute and chronic rejection of the transplanted organs as well as transplant-against-host reactions, giving less toxic doses of drugs.

Conclusion: The main aim of this work was to find evidence of the need for wider application of apheresis technologies in transplantation.

Keywords: Transplantation; Rejection; Graft-vs.-host response; Antibodies; Plasmapheresis

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Introduction

Despite all the achievements of modern transplantology, there is still a problem of acute or chronic rejection of the transplanted organs. These processes have many causes that not always can be eliminated. There is activation of the innate immunity elements against foreign microorganisms and of the antigenic structure disorders control of even their own cells (including the tumor ones) to perform their timely removal and cleansing of the body [1]. Humoral autoimmune mechanisms of antibody formation against the transplanted organ cell antigens are the main cause of graft rejection during the first year of the transplant [2,3]. T- and B-lymphocytes, natural killers, macrophages or polymorphonuclear leukocytes play a crucial role in rejection reactions [4]. At the same time, T-cells (T-killers) can affect the transplanted cells by release of cytokines (TNF- α , IL-2, etc.), contributing to the apoptosis of both their own damaged and foreign cells. On the other hand, they give a signal to the B-cells to form donor-specific autoantibodies [5].

Drug immunosuppression methods

Therefore, the main task after organ transplantation is to suppress these mechanisms of rejection by various immunosuppression methods. At the same time, it is necessary to immediately identify its potential dangers such as activation of microbial, viral, and fungal infection, up to the multiple organ

insufficiency, as well as carcinogenesis [6-9]. Post-transplant children may develop lymphoproliferative disorders with high mortality rates [10]. Patients predisposed to coronary heart disease after transplantation may experience activation of atherosclerosis with symptoms of heart failure and even sudden death [11,12]. Therefore, it is necessary to avoid toxic side effects of such drugs. Modern immunosuppressive therapy consists of a combination of several drugs – cyclosporine, azathioprine, prednisolone, and in recent years – rituximab, tacrolimus, and mycophenolate mofetil. However, such treatment is fraught with a number of complications. Glucocorticoids in particular contribute to development Cushingoid syndrome, hypertension, diabetes, and osteoporosis. In addition in 30% of patients they do not prevent the transplant rejection [13]. It should be noted that the incidence of diabetes after transplantation of various organs reaches 40% [14]. Cyclosporine A has a pronounced nephrotoxicity [15,16]. One of the cyclosporin-A side effects is hypertriglyceridemia that develops after transplantation of bone marrow cells, which can be controlled with plasma exchange or double filtration plasmapheresis [17,18]. Azathioprine often causes pancytopenia and interstitial pneumonitis [19,20]. Methotrexate also contributes to development of lymphopenia, infectious and pulmonary complications [21]. Its nephrotoxicity

may lead to acute renal failure [22]. Calcineurin inhibitors (tacrolimus, sirolimus) are also widely used in transplantology but their use is also fraught with serious complications such as thrombotic microangiopathy with development of nephropathy (acute tubular necrosis), cholestasis, encephalopathy (headaches, seizures), and high rate of lymphoproliferative tumors [23-28]. At the same time, the rejection rate remains quite high [29,30]. Tacrolimus may also cause acute demyelinating polyneuropathy of the Guillain-Barré syndrome type [31]. In addition, it also has nephrotoxic and diabetogenic effects, causing apoptosis of β -cells [32-35]. This often negates the results of operations and leads to the most adverse consequences. In these cases, plasmapheresis helps relieve such toxic complications and save the organ [31].

Rituximab (anti-lymphocytic immunoglobulin) is able to stop acute rejection reactions, but it is less effective in chronic forms [36]. In particular, it is far from being the most "magic drug" in recurrence of nephrotic syndrome after kidney transplantation [37]. Its long-term use is associated with increasing endotoxemia, contributing to neutropenia development followed by septic complications [38], and progressive leukoencephalopathy [39]. The use of mycophenolate mofetil is accompanied by persistent colitis and even colon ulcers [40-42]. The same complications occur with combined use with tacrolimus and mycophenolate mofetil [43]. According to the data of two transplantation centers chronic rejection processes of the transplanted lungs occurs in 60 to 80% of cases. The use of anti-lymphocytic antibodies during transplant rejection episodes is associated with activation of viral infections and lymphoproliferative diseases. In addition, in some cases, a graft-vs.-host reaction develops with the same consequences as after stem cell transplantation. In such cases, plasmapheresis enables to treat such complications [44]. Immune conflict after organ transplantation may result from anti-HLA antibodies appeared induced by isoimmunization due to previous transplants, blood transfusions or pregnancy. Despite cross-compatibility with the donor organ, presence of such HLA antibodies leads to early rejection. Growth of donor-specific alloantibodies occurs during the first few weeks [45]. In some cases, affected by natural autoantibodies IgG and IgM isotopes to endothelin-1 and angiotensin II and activation of the complement system and endothelial cells an ultra-acute rejection reaction may develop within the next few hours and even minutes [46]. However, it is to be taken into account that the cost of such drug therapy in acute crises is very high reaching up to \$49,000-155,000 [47,48]. Mycophenolate mofetil therapy cost can vary from \$638,018 to \$752,107 in the course of a year [49].

Research Methodology

Methods of apheresis therapy

In cases of upcoming transplantation, without removing the remaining autoantibodies in the body, the transplanted kidney is at risk of the same autoimmune lesion as the removed one. Recurrence of focal segmental glomerulosclerosis in the transplanted kidney is observed in 30-80% of cases, and the transplant death within three years reaches 90% and only systematic conventional or cascade plasmapheresis after

surgery prevents such complications [25,50-60]. A much higher incidence of rejection due to prior IgA nephropathy or focal segmental glomerulosclerosis is noted in cases of "live" kidney transplantation than in "cadaveric" one [61]. Sometimes this requires weekly plasmapheresis for almost four years [62-64]. The rise of hypoproteinemia with prolonged courses of plasma exchange forces to perform immunoadsorption instead [65]. Combination of plasmapheresis with rituximab or bortezomib enabled to achieve a longer remission of proteinuria [66-68]. However, it should be considered that rituximab itself can lead to severe complications, which also have to be treated with plasmapheresis [69,70]. Extracorporeal photopheresis, being quite efficient in cell-mediated rejection of the transplanted lungs and heart, is not applied in renal transplantation [71].

Immunoadsorption prior to transplantation improves outcomes and prognosis, even in cases of early rejection of the transplanted kidneys [72]. However, repeated procedures of plasmapheresis more reliably prevent rejection of the transplanted kidney than high-dose immunoglobulins [55]. High incidence of graft rejection in hemolytic-uremic syndrome also requires a preliminary course of plasmapheresis with eculizumab [73]. Plasmapheresis was necessary after transplantation, too [74]. After transplantation, a new situation arises when an antigenic signal comes from the transplanted kidney, in response to which new antibodies begin to form reaching the peak of acute "rejection crisis" in 1-2 weeks. In this case apheresis therapy can relieve these immune responses associated with lower level of immunosuppressive therapy. The course of plasmapheresis in such cases contributes to restoration of diuresis, reduction of creatinine level and gradual restoration of the graft function, which in 60% of patients enabled to avoid "graftectomy" [75]. Moreover, the transplanted organ, whether kidney, heart, lung, liver or bone marrow, is a constant driver of antibody reproduction during the whole life long, making this process one of the options of autoimmune diseases. Thus, anti-myosin antibodies are often found in case of the transplanted heart rejection [76]. Graft rejection can occur even after 10 years. Alloreactive antibodies are formed against the graft and they are also retained after its rejection, which reduces the chances of survival of the subsequent graft. Cells dysfunction is one of the causes of such resistant alloimmunization or hyperreactivity. In this case, uremia in renal failure plays the leading role even more than T-cell regulation [77]. Significant problems arise when kidney transplantation has to be performed in case of ABO-incompatibility. One of the reasons is the presence of A or B antigens, not only on the erythrocytes membranes, but also on the blood vessels walls, including the transplanted kidney [78]. In this case, the recipient antibodies begin to interact with antigens on the walls of the vessels, leading to microcirculation disorders and subsequent rejection of the transplanted organ ("high incompatibility"). "Low incompatibility" arises in the result of the donor lymphocytes production, being left in the transplanted organ as a kind of "passengers", against the recipient erythrocytes, causing their hemolysis [79]. In these cases, preliminary removal of anti-A or anti-B antibodies by plasma exchange significantly reduces the antibodies level, thereby minimizing the rejection reaction [80-82]. In organ

transplantation in the case of blood incompatibility according to the ABO system cascade plasmapheresis was successfully used [83,84]. At the same time, during 3-4 procedures of cascade plasmapheresis it was possible to reduce the anti-AB antibodies titer to the concentration ratio of 1:32, which is an acceptable criterion for subsequent kidney transplantation in ABO-incompatibility. The same problems with ABO-incompatibility arise in hematopoietic stem cells transplantation in hematology and the course of plasmapheresis before such transplantation to a large extent prevents crises of rejection [85]. In cases of incompatible ABO transplantation of relative's donor organs plasmapheresis is performed to reduce the relevant antibodies titer in ration of 1:2 to 1:4. At the same time in cases of the initial antibodies titer, up to 350-550% of the circulating plasma volume (CPV) has to be removed, and in case of the repeated increase of the antibodies titer up to 1:32 in the post-transplantation period it was necessary to resort to high-volume plasmapheresis with removal of up to 400% of the CPV [86]. Courses of plasmapheresis, both before and after the kidney transplantation in case of ABO-incompatibility, eliminated episodes of hyperactive antibody-dependent graft rejection [87]. Moreover, during the year, the transplanted kidneys were viable in 100% of cases. Positive results were also achieved when using immunoadsorption prior to ABO-incompatible kidney transplantation [88]. However, an increase in blood loss in such operations was observed [89]. More than 30% of potential kidney transplant recipients have a high level of anti-HLA antibodies, which can also be overcome by pre-transplant immunosuppression using plasmapheresis [90]. When it is needed to perform the kidney transplantation on the background of antiphospholipid syndrome, a preventive plasma exchange is also advisable due to the risk of early vascular thrombosis of the transplanted organ [91]. It should be considered that organ transplantation inevitably leads to a number of disorders, the treatment of which should require apheresis therapy. After all, almost all patients who require organ transplantation, by the time of the operation have very significant homeostasis disorders and endotoxemia due to the organ failure. It's either renal, cardiac, pulmonary, or even multiple organ failure. The donor organ, which has just suffered hypoxia during the period of removal and transportation and before that it has suffered stress and pre-death endotoxemia of the donor, does not get into the best conditions of the new host's internal environment after the transplantation, which hinders its adequate functioning on the new site. It was noted that content of average molecular weight toxins and malon dialdehyde even before the operation was higher than normal values and it continued to increase during the operation, reaching its maximum in 1 hour after the blood flow started in the transplanted kidney [92]. Therefore, apheresis therapy and detoxification of the recipient in the preoperative period seems relevant, and ideally, of the donor as well before his organ to be removed. It is believed that the graft dysfunction in the early postoperative period is due to hypoxia during preservation, immunological conflict, development of intravascular coagulation and microcirculation disorders in the transplanted organ, fraught with its functional failure [93]. Therefore, in 11 highly sensitized patients with high

antibody titer, 30-40% of CPV were eliminated immediately prior to the kidney transplantation and 40-50% of CPV – during re-transplantation (replacement of the removed plasma was carried out by albumin and fresh frozen plasma). In all cases, the graft began to function normally on the operating table with normalization of creatinine and urea levels on the 3rd – 4th days and without rejection crises. If necessary, repeated procedures of plasmapheresis were carried out in different postoperative periods [94]. One of such patients underwent a program plasmapheresis successfully for three years at 4 months intervals [95]. It was found that the level of the main proinflammatory cytokines (IL-6, IL-8, IL-10), toxins of the average molecular weight and lipid peroxidation products increases and reaches maximum by the end of the operation, which was the main indication for intraoperative plasmapheresis immediately after the inclusion of the transplanted kidney in the bloodstream [96]. Besides the reduction of such endotoxins in the blood, restoration of the initial nitrogen and water excretion function of the kidneys occurred much faster - by 5-6 days to compare with 12-18 days in cases without plasmapheresis [97]. Apheresis therapy is also advisable in the postoperative period for sanitation of the internal environment from intraoperative stress agents, which should simplify the "inclusion" of the transplanted organ [98,99]. Plasmapheresis in the volume of 1 CPV in the next 2-3 hours after inclusion of the transplanted kidney in the bloodstream reduced the level of average molecules by 25% below the preoperative level, prevented oligoanuria, the need for hemodialysis, contributed to a faster normalization of creatinine and increased actuarial survival of the transplants [100]. Plasmapheresis, performed immediately after the liver transplantation also prevented its dysfunction [101]. The same tactics of intra- and postoperative plasmapheresis was used in the heart transplantation on the background of high antibody and tissue incompatibility [102]. It was possible to stop acute rejection, which occurred 10 days after kidney transplantation, using anti-CD3 mouse monoclonal antibodies in combination with intensive course of plasmapheresis (7 procedures with removal of up to 2.5-3 liters of plasma per procedure) [103]. In type I diabetes, the pancreas transplantation is also often used [104], although this does not always stop the diabetic foot syndrome development with the need for amputations of the lower limbs [105]. Plasmapheresis on the background of immunosuppressive therapy was able to block the rejection reactions after transplantation of the small bowel loops [106]. In case of the liver transplantation on the background of ABO-incompatibility, plasmapheresis was also used before the operation [107,108], as well as both before and after surgery [109]. The same course of plasmapheresis before the liver transplantation is used in the presence of autoantibodies against tissue antigens of the liver [110]. Preoperative administration of rituximab alone (without plasmapheresis) was not able to block the antibodies production [80]. Splenectomy in combination with rituximab is not reported to have positive results, too [111]. Plasmapheresis should be applied as soon as the first signs of organ rejection appear [112,113]. After the liver transplantation, ischemic parenchymal disorders may develop with formation of infarction, which can

also be stopped with help of plasmapheresis courses [114]. In addition, development of cholestatic fibrosing hepatitis is also described, for the relief of which cascade plasmapheresis was used [115]. Since in such autoimmune processes practically the only and truly pathogenetic treatment is the apheresis therapy and among its methods the most effective is plasmapheresis, the principle of periodic courses of plasmapheresis aiming to remove antibodies against the transplanted organ should be introduced in transplantology [116-118]. At the same time, of course, immunosuppressive therapy remains relevant, but plasmapheresis will help to carry it out in subtoxic doses without the risk of side effects. The best effect was achieved by combining courses of plasmapheresis with intravenous administration of immunoglobulins [119]. Reduction of organ dysfunction after the liver transplantation by both conventional plasmapheresis and MARS (molecular adsorbent recirculating system) was equally effective [112].

After the heart transplantation, even in cases where the cause of transplantation was not ischemic heart disease, but dilated cardiomyopathy, immunosuppressive therapy leads to significant disturbance of the blood lipid composition and the occurrence of ischemic disorders in the transplanted heart. Since plasmapheresis is one of the most effective methods of treatment of hyperlipidemic conditions, it was included in the treatment program of 8 patients who underwent orthotopic heart transplantation 2-7 years after the operation. After courses of plasmapheresis, conducted twice a year, there was a pronounced positive dynamics in the state of hemorheology and lipid metabolism. Blood levels of total cholesterol and low-density lipoproteins were significantly reduced without significant changes in the concentration of immunosuppressors. Scintigrams showed significant improvement in myocardial perfusion in the ischemic areas [120]. It is especially important to conduct plasmapheresis to relieve acute crisis of rejection, even with unstable hemodynamics [121]. And it was emphasized that after 7 episodes of rejection in 7 patients treated without plasmapheresis, only 2 survived, and after 11 such episodes in 6 patients with plasmapheresis all of them survived [122]. At Birmingham University (Alabama, USA) plasmapheresis method is considered first-line method to overcome the crises of the heart rejection [123]. Plasmapheresis has been successfully used in crises of rejection of the transplanted heart even in children, starting from the age of 3.5 months [124]. Plasmapheresis was also effective after combined heart and liver transplantation in the presence of donor-specific antibodies [125]. Taking into account HLA-antigen allosensitization development in "candidates" for heart transplantation, it is advisable to carry out plasmapheresis before transplantation [126]. In addition, removal of antibodies with help of plasmapheresis is used during extracorporeal blood circulation during the heart transplantation [127]. Currently, the optimal tactics to prevent graft rejection is considered to be a combination of plasmapheresis with intravenous introduction of immunoglobulins (sometimes with addition of rituximab) [82,128,129]. Along with plasmapheresis, immunoabsorption is also used [118]. Cascade plasmapheresis

is also an effective and safe treatment method to relieve the crises of the transplanted organ acute rejection [130]. The lung transplantation has the lowest median survival rate compared to other organs [131]. Therefore, such preventive apheresis therapy is particularly relevant here, especially in cases of allosensitization to HLA-antigens [90]. First, the donor lung, in principle, cannot be "normal", because at the time of death, the donor should develop such endotoxemia, which can lead to respiratory distress development in their lungs, and ischemia with hypoxia at the time of the organ removal and its transportation can add additional damage, too. This was shown in the research of the experimental pathology laboratory, Institute of Pulmonology by E.N. Danilov, G.M. Kudryashov and E.D. Shekhunov back in 1980-1990-ies [132]. In addition, as the period of ischemic tissue increases, release of such toxic substances as "large histocompatibility complex" class II, and IL-2 and IFN- γ content increases in the bronchoalveolar lavage fluid, which significantly increases the rejection risk [133].

Secondly, immediately after the transplantation and inclusion in the bloodstream, pathological metabolites of the recipient, who has been in the state of severe respiratory failure for a long time, together with intraoperative BAS overwhelms them. Everything abovementioned also requires detoxification in the earliest postoperative, and maybe even in the intraoperative period [134]. This is especially true in lung transplantation for emergency indications on the background of severe respiratory distress syndrome, when only with help of plasmapheresis it is possible to ensure the normal function of the transplanted lungs [135,136].

The risk of acute rejection of the transplanted lung during the first year is 55% [137]. Even if the transplanted lung avoid the rejection reaction a significant number of patients (up to 60-80%) develop a progressive obliterative bronchiolitis, which is not less difficult to treat [138,139], including in children [140]. The risk of death often forces the patient to resort to repeated lung transplantation [141]. It is detected by reduction of the forced exhalation volume for 1 second to less than 80% compared to the early (basic) post-transplantation period. This syndrome can be considered a kind of chronic rejection reaction. It is characterized by a progressive fibroproliferative process of *lamina propria* of the small bronchi wall with lumen narrowing down to 2 mm with a relatively normal surrounding parenchyma [142]. Conducting plasmapheresis with subsequent administration of immunoglobulins or rituximab can prevent not only the rejection crisis, but also obliterative bronchiolitis [134,137,143,144], including in children [145]. When thrombotic microangiopathy develops, especially with tacrolimus administration or Guillain-barré syndrome, a more intensive course of plasmapheresis is required [28,31,146]. Extracorporeal photopheresis is also used [147]. In some cases, in highly sensitized patients particularly in those with ABO-incompatibility it is advisable to perform preoperative courses of rituximab and plasmapheresis [148,149], although it may pose a risk of infectious complications in the postoperative period [150].

Results

The bone marrow cell transplantation often results in acute or chronic graft-vs.-host disease (GVHD) with rate of up to 46% [151]. In acute GVHD, cells and tissues of the body are recognized to be "foreign" for the transplanted donor cells, which contributes to the launch of pathological reactions involving effector T- and B-lymphocytes, natural killer cells with release of a number of proinflammatory cytokines (IL-1 β , IL-6, IL-17, IL-18, TNF- α) and this "cytokine cascade" activates other effector cells – natural killers and macrophages, leading to direct tissues damage of the skin, liver, and gastrointestinal tract [152-154]. GVHD plays the leading role in complications and deaths after the bone marrow or stem cell transplantation [155]. The gastrointestinal form of GVHD is the most severe and is the leading cause of death [156,157]. It is especially severe in children when the mortality rate reaches 60% [158]. The liver damage appears to be rather common consequence caused by veno-occlusive liver disease, chronic viral or fungal infection, and cholestatic disorders [159,160]. In GVHD acute lung damage develops in the form of severe respiratory distress syndrome showing 28-day mortality of 46.6% of patients, and by the end of the year it reaches 66.9% [161]. It is aggravated by activation of viral-bacterial and fungal infection associated with leukopenia [162]. However, GVHD can develop after transplantation of solid organs and it is rather severe in such cases with fatal outcome. In particular, it occurs after the liver transplantation, when after interaction of the donor T-lymphocytes with cellular antigens of the patient (recipient), they proliferate and clonally expand leading to the liver, skin, bowels, bone marrow lesions and subsequent development of sepsis, multiple organ failure with fatal outcome [163]. All this are indications for plasmapheresis and we are convinced of its effectiveness in the treatment of GVHD [164,165]. It should be noted that chronic GVHD can develop in 50% of such patients with mortality up to 25% [166], which once again emphasizes the urgency of this problem and the need for more active treatment involving plasmapheresis.

Discussion

A frequent complication of stem cell transplantation is thrombotic microangiopathy associated with intima edema and fibrinoid necrosis of the vascular walls. This is due to high doses of conventional chemotherapy drugs, radiation therapy, calcineurin inhibitors (used for prevention and treatment of GVHD) and infection [167,168]. The kidneys are primarily affected and the renal insufficiency appears to be a poor prognostic factor with 44-90% mortality rate [169,170]. Plasma exchange, carried out daily until the positive effect appears, can control this complication [164,171,172]. In case of the lungs lesions ECMO may be required [173,174]. However, attempts are also made to directly suppress

T- and B- lymphocytes hyperactivity (producers of cytokines and antibodies), with help of extracorporeal photopheresis, when irradiation of the isolated cells with ultraviolet rays is carried out [175]. After ingestion of methoxalen - photosensitizer drug in a dose of 1 mg/kg to achieve the blood plasma concentration of more than 50 mg/DL, 90 minutes later leukopheresis is performed with extracorporeal irradiation of thin layer of white blood cells with long-wave ultraviolet rays followed by subsequent reinfusion of these cells in apoptosis state. The latter are photoactive and covalently bind to pyridine bases of leukocyte molecules of the membrane and cytoplasm, which leads to their lethal damage. These cells then reinfused to the patient and die within 1-2 weeks, but during this interval they stimulate autosuppression reaction, partly directed against T-cells, but also damaging the non-irradiated clones of T-cells. This method made it possible to reduce the frequency of both rejection crises and infectious complications [176]. Although such procedures have been successfully carried out for more than 20 years, the mechanisms of their therapeutic effect are still not fully understood [177,178]. Of course, it would be tempting to affect this way the specific T-cells that damage the graft, but such a reaction is not selective and possible lethal damage due to ultraviolet irradiation of other lymphocytes clones can lead to the most unpredictable consequences for the body as a whole. Besides, photopheresis does not remove the formed autoantibodies, which makes this procedure incomplete, since the remaining autoantibodies continue their damaging effect on the tissues and organs of the patient. That's why plasmapheresis was successfully performed weekly before each photopheresis procedure [179-182]. And we also think photopheresis and plasmapheresis to be the most appropriate combination.

Conclusion

The problem of organ transplantation is still far from being solved. Until now, to eliminate both acute and chronic rejection of the transplanted organs they use drugs with many side effects. At the same time, apheresis therapy methods, aimed at removing antibodies from the body, have not received enough proper and timely application. Indications to remove pathological agents that caused organ damage arise even in the preoperative period. It is advisable to use them even in the intraoperative period and immediately after the transplantation for faster and optimal restoration of the transplanted organs functions. The simplest and safest method of apheresis therapy is plasmapheresis, which removes gradually formed autoantibodies. Performing it regularly enables to prevent rejection crises, using drugs in less toxic doses. Russian equipment with "Rosa" or "PFM-500" plasma filters allows conducting membrane plasmapheresis even in outpatient settings.

References

- 1 Wood KJ, Goto R (2012) Mechanisms of rejection: Current perspectives. *Transplantation* 93: 1-10.
- 2 Petty M (2016) Antibody-mediated rejection in solid organ transplant. *AACN Adv Crit Care* 27: 316-323.
- 3 Cozzi E, Colpo A, De-Silvestro G (2017) The mechanisms of rejection in solid organ transplantation. *Transfus Apher Sci* 56: 498-505.

- 4 Moreau A, Varey E, Anegon I, Cuturi MC (2013) Effector mechanisms of rejection. *Cold Spring Harb Perspect Med* 3: a015461.
- 5 Baran T, Boratynska M (2017) Immunoregulatory role of B lymphocytes in alloresponse to kidney transplant. *Postepy Hig Med Dosw* 71: 254-266.
- 6 Jeulin H, Agrinier N, Guery M (2013) Human herpesvirus 6 infection after allogeneic stem cell transplantation: Incidence, outcome and factors associated with HHV-6 reactivation. *Transplantation* 95: 1292-1298.
- 7 Mencarelli F, Marks SD (2012) Non-viral infections in children after renal transplantation. *Pediatr Nephrol* 27: 1465-1476.
- 8 Matthes-Martin S, Boztug H, Lion T (2013) Diagnosis and treatment of adenovirus infection in immunocompromised patients. *Expert Rev Anti Infect Ther* 11: 1017-1028.
- 9 Miloh T, Barton A, Wheeler J (2017) Immunosuppression in pediatric liver transplants: Unique aspects. *Liver Transpl* 23: 244-256.
- 10 Stréhn A, Szőnyi K, Kriván G (2014) Post-transplantation lymphoproliferative disorders in childhood. *Orv Hetil (Hungarian)* 155: 313-318.
- 11 Gillis KA, Patel RK, Jardine AG (2014) Cardiovascular complications after transplantation: Treatment options in solid organ recipients. *Transplant Rev (Orlando)* 28: 47-55.
- 12 Ketelhuth DF, Hansson GK (2015) Modulation of autoimmunity and atherosclerosis-common targets and promising translation approaches against disease. *Circ J* 79: 924-933.
- 13 Rekers NV, De-Fijter JW, Claas FH, Eikmans M (2016) Mechanisms and risk assessment of steroid resistance in acute kidney transplant rejection. *Transpl Immunol* 38: 3-14.
- 14 Gosmanov AR, Dagogo-Jack S (2012) Predicting, managing and preventing new-onset diabetes after transplantation. *Minerva Endocrinol* 37: 233-246.
- 15 Damiano S, Ciarcia R, Montagrano S (2015) Prevention of nephrotoxicity induced by cyclosporine-A: Role of antioxidants. *J Cell Biochem* 115: 364-369.
- 16 Tolou-Ghamari Z, Mortazavi M, Palizban AA, Najafi MR (2015) The investigation of correlation between Iminoral concentration and neurotoxic levels after kidney transplantation. *Adv Biomed Res* 4: 59.
- 17 Giannini G, Valbonesi M, Morelli F (2005) Hypertriglyceridemia: Apheretic treatment. *Int J Artif Organs* 28: 1018-1024.
- 18 Moorman MT, Epstein RB, Smith JW (2011) Management of cyclosporine overdose in a hematopoietic stem cell transplant patient with sequential plasma exchange and red blood cell exchange. *J Clin Apher* 26: 156-158.
- 19 Willerding-Möllman S, Wilkens L, Schlegelberger B, Kaiser U (2004) Azathioprine-associated myelodysplastic syndrome with cytogenetic aberrations. *Dtsch Med Wochenschr (German)* 129: 1246-1248.
- 20 Nagy F, Molnár T, Makula E (2006) Azathioprine-associated interstitial pneumonitis. *Orv Hetil (Hungarian)* 147: 259-262.
- 21 Lateef O, Shakoob N, Balk RA (2005) Methotrexate pulmonary toxicity. *Expert Opin Drug Saf* pp: 723-730.
- 22 Howard SC, McCormick J, Pui CH (2016) Preventing and managing toxicities of high-dose methotrexate. *Oncologist* 21: 1471-1482.
- 23 Hesselink DA, Bouamar R, Van-Gelder T (2010) The pharmacogenetics of calcineurin inhibitor-related nephrotoxicity. *Ther Drug Monit* 32: 387-393.
- 24 Matsusaki T, Morimatsu H, Sato T (2010) Thrombotic microangiopathy after living-donor liver re-transplantation. *J Anesth* 24: 614-617.
- 25 Mitome J, Yamamoto H, Maruyama Y (2010) Successful treatment of recurrent focal segmental glomerulosclerosis combined with calcineurin inhibitor nephrotoxicity four year after kidney transplantation. *Clin Transplant* 24 (22): 48-53.
- 26 Pomerantz RG, Campbell LS, Jukic DM, Geskin LJ (2010) Posttransplant cutaneous T-cell lymphoma: Case reports and review of the association of calcineurin inhibitor use with posttransplant lymphoproliferative disease risk. *Arch Dermatol* 146: 513-516.
- 27 Oto T, Okazaki M, Takata K (2010) Calcineurin inhibitor-related cholestasis complicating lung transplantation. *Ann Thorac Surg* 89: 1664-1665.
- 28 Reig-Mezquida JP, Jover AS, Ansótegui-Barreta E (2015) Thrombotic microangiopathy associated with tacrolimus in lung transplantation. *Arch Bronconeumol* 51: 23-24.
- 29 Lemaitre F, Blanchet B, Latournerie M (2015) Pharmacokinetics and pharmacodynamics of tacrolimus in liver transplant recipients: Inside the white blood cells. *Clin Biochem* 48: 406-411.
- 30 Sikma MA, Van-Maarseveen EM, Van de Graaf EA (2015) Pharmacokinetics and toxicity of tacrolimus early after heart and lung transplantation. *Am J Transplant* 15: 2301-2313.
- 31 Sharma NS, Wille KM, Hoopes CW, Diaz-Guzman E (2014) Acute demyelinating polyneuropathy after lung transplantation: Guillain-Barré syndrome or tacrolimus toxicity?. *Case Rep Transplant p*: 685010.
- 32 Rangel EB (2014) Tacrolimus in pancreas transplant: A focus on toxicity, diabetogenic effect and drug-drug interactions. *Expert Opin Drug Metab Toxicol* 10: 1585-1605.
- 33 Öztürk Z, Gönc EN, Akcan L (2015) A rare but important adverse effect of tacrolimus in a heart transplant recipient: Diabetic ketoacidosis. *Turk J Pediatr* 57: 533-535.
- 34 Scalea JR, Levi ST, Ally W, Brayman KL (2016) Tacrolimus for the prevention and treatment of rejection of solid organ transplants. *Expert Rev Clin Immunol* 12: 333-342.
- 35 Akhtar T, Sheikh N, Shan T (2017) Tacrolimus induced nephrotoxicity and pulmonary toxicity in Wistar rats. *J Biol Regul Homeost Agents* 31: 1061-1066.
- 36 Macklin PS, Morris PJ, Knight SR (2017) A systematic review of the use of rituximab for the treatment of antibody-mediated renal transplant rejection. *Transplant Rev (Orlando)* 31: 87-95.
- 37 Grenda R, Jarmužek W, Rubik J (2016) Rituximab is not a magic drug in post-transplant recurrence of nephrotic syndrome. *Eur J Pediatr* 175: 1133-1137.
- 38 Ahmadi F, Dashti-Khavidaki S, Khotami MR (2016) Rituximab-related late-onset neutropenia in kidney transplant recipients treated for antibody-mediated acute rejection. *Exp Clin Transplant* 15: 414-419.
- 39 Loyaga-Rendon RY, Taylor DO, Koval CE (2013) Progressive multifocal leukoencephalopathy in a heart transplant recipient following rituximab therapy for antibody-mediated rejection. *Am J Transplant* 13: 1075-1079.
- 40 Liapis G, Boletis J, Skalioti C (2013) Histological spectrum of mycophenolate mofetil-related colitis: Association with apoptosis. *Histopathology* 63: 649-658.
- 41 Curtin BR, Rachakonda VP, Von-Rosenvinge EC (2014) Unusually late-

- onset mycophenolate mofetil-related colitis. *Am J Health Syst Pharm* 71: 1858-1861.
- 42 Sonoda A, Wada K, Mizukami K (2017) Deep ulcers in the ileum associated with mycophenolate mofetil. *Intern Med* 56: 2883-2886.
 - 43 Van-Boekel GA, Aarnoutse RE, Van der Heijden JJ (2012) Effect of mild diarrhea on tacrolimus exposure. *Transplantation* 94: 763-767.
 - 44 Rossi AP, Bone BA, Edwards AR (2014) Graft-vs.-host disease after simultaneous pancreas-kidney transplantation: A case report and review of the literature. *Am J Transplant* 14: 2651-2656.
 - 45 Stegall MD, Gloor J, Winters JL (2009) A comparison of plasmapheresis versus high-dose IVIG desensitization in renal allograft recipients with high levels of donors specific alloantibody. *Am J Transplant* 6: 346-351.
 - 46 Cozzi E, Calabrese F, Schiavon M (2017) Immediate and catastrophic antibody-mediated rejection in a lung transplant recipient with anti-angiotensin II receptor type 1 and anti-endothelin-1 receptor type A antibodies. *Am J Transplant* 17: 557-564.
 - 47 Kim M, Martin ST, Townsend KR, Gabardi S (2014) Antibody-mediated rejection in kidney transplantation: A review of pathophysiology, diagnosis, and treatment options. *Pharmacotherapy* 34: 733-744.
 - 48 Muduma G, Odeyemi I, Smith-Palmer J, Pollock RF (2016) Review of the clinical and economic burden of antibody-mediated rejection in renal transplant recipients. *Adv Ther* 33: 345-356.
 - 49 Martinez-Mier G, Salazar-Ramirez A (2016) The cost of gastrointestinal adverse events and the impact of dose-reductions/discontinuations on acute rejection in kidney transplant patients of mycophenolate mofetil-related compared to enteric-coated mycophenolate sodium: A pharmaco-economic study. *Transplant Proc* 48: 588-595.
 - 50 Sener A, Bella AJ, Nguan C (2009) Focal segmental glomerular sclerosis in renal transplant recipients: Predicting early disease recurrence may prolong allograft function. *Clin Transplant* 23: 96-100.
 - 51 Varma PP, Hooda AK, Kumar A, Singh L (2009) Highly successful and low-cost desensitization regime for sensitized living donor renal transplant recipients. *Ren Fail* 31: 533-537.
 - 52 Fuentes GM, Meseguer CG, Carrion AP (2010) Long-term outcome of focal segmental glomerulosclerosis after pediatric renal transplantation. *Pediatr Nephrol* 25: 529-534.
 - 53 Satoscar AA, Pelletier R, Adams P (2010) *De novo* thrombotic microangiopathy in renal allograft biopsies - role of antibody-mediated rejection. *Am J Transplant* 10: 1804-1811.
 - 54 Schachter ME, Monahan M, Radhakrishnan J (2010) Recurrent focal segmental glomerulosclerosis in the renal allograft: Single center experience in the era of modern immunosuppression. *Clin Nephrol* 74: 173-181.
 - 55 Stegall MD, Gloor JM (2010) Deciphering antibody-mediated rejection: New insights into mechanisms and treatment. *Curr Opin Organ Transplant* 15: 8-10.
 - 56 Bartel G, Schwaiger E, Bohmrig GA (2011) Prevention and treatment of alloantibody-mediated transplant rejection. *Transplant Int* 24: 1142-1155.
 - 57 George SM, Balogun RA, Sanoff SL (2011) Therapeutic apheresis before and after kidney transplantation. *J Clin Apher* 26: 252-260.
 - 58 Montgomery RA, Lonze BE, King KE (2011) Desensitization in HLA-incompatible kidney recipients and survival. *N Engl J Med* 365: 318-326.
 - 59 Rummel S, Maier K, Barz D (2013) Therapeutic apheresis in transplantation medicine, experience with cardiac and lung transplantation in Jena. *Atheroscler Suppl* 14: 33-38.
 - 60 Straatmann C, Kallash M, Killakey M (2014) Success with plasmapheresis treatment for recurrent focal segmental glomerulosclerosis in pediatric renal transplant recipients. *Pediatric Transplantation* 18: 29-34.
 - 61 Deng R, Dai Y, Zhang H (2018) Higher incidence of renal allograft glomerulonephritis in living-related donor kidney transplantation. *Transplant Proc* 50: 2421-2425.
 - 62 Deegens JK, Andresdottir MB, Croockewit S, Wetzels JF (2004) Plasma exchange improves graft survival in patients with recurrent focal glomerulosclerosis after renal transplantation. *Transpl Int* 17: 151-157.
 - 63 Canaud G, Zuber J, Sberro R (2009) Intensive and prolonged treatment of focal and segmental glomerulosclerosis recurrence in adult kidney transplant recipients: A pilot study. *Am J Transplant* 9: 1081-1086.
 - 64 Noorlander I, Hesselink DA, Wabbijn M, Betjes MGH (2011) High cut-off haemodialysis induces remission of recurrent idiopathic focal segmental glomerulosclerosis after renal transplantation but is no alternative to plasmapheresis. *NDT Plus* 4: 321-323.
 - 65 Fencel F, Simková E, Vondrák K (2007) Recurrence of nephrotic proteinuria in children with focal segmental glomerulosclerosis after renal transplantation treated with plasmapheresis and immunoadsorption: Case reports. *Transplant Proc* 39: 3488-3490.
 - 66 Hickson LJ, Gera M, Amer H (2009) Kidney transplantation for primary focal segmental glomerulosclerosis: Outcomes and response to therapy for recurrence. *Transplantation* 87: 1232-1239.
 - 67 Jordan SC, Reinsmoen N, Peng A (2010) Advances in diagnosing and managing antibody-mediated rejection. *Pediatr Nephrol* 25: 2035-2048.
 - 68 Sakai K, Takasu J, Nihei H (2010) Protocol biopsies for focal segmental glomerulosclerosis treated with plasma exchange and rituximab in renal transplant patient. *Clin Transplant* 24(22): 60-65.
 - 69 Levine MH, Abt PL (2012) Treatment options and strategies for antibody mediated rejection after renal transplantation. *Semin Immunol* 24: 136-142.
 - 70 Hastings D, Patel B, Torloni AS (2009) Plasmapheresis therapy for rare but potentially fatal reaction to rituximab. *J Clin Apher* 24: 28-31.
 - 71 Deborska-Materkowska D, Kozińska-Przybył O, Mikaszewska-Sokolewicz M, Durlik M (2014) Fatal late-onset pneumocystis pneumonia after rituximab: Administration for post-transplantation recurrence of focal segmental glomerulosclerosis-case report. *Transplant Proc* 46: 2908-2911.
 - 72 Sanchez AP, Ward DM (2012) Therapeutic apheresis for renal disorders. *Semin Dial* 25: 119-131.
 - 73 Higgins RM, Bevan DJ, Vaughan RW (1996) 5-year follow-up of patients successfully transplanted after immunoadsorption to remove anti-HLA antibodies. *Nephron* 74: 53-57.
 - 74 Nester C, Stewart Z, Myers D (2011) Pre-emptive eculizumab and plasmapheresis for renal transplant in atypical hemolytic uremic syndrome. *Clin J Am Soc Nephrol* 6: 1488-1494.
 - 75 Mack R, Mastrovovich J, Grimes D, Davis O (2014) Significance in establishing a differential diagnosis in management of thrombotic microangiopathies. *J Clin Apher* 29: 38-39.

- 76 Biryukova LS, Moseshvili EG, Fetisova EV (2001) Plasmapheresis in the treatment of acute transplant rejection]. Proc. IX Conference, Moscow Society of Hemapheresis, Moscow (Rus) pp: 9-10.
- 77 Shütz A, Beuer M, Kemkes BM (1997) Antimyosin antibodies in cardiac rejection. *Ann Thorac Surg* 63: 578-581.
- 78 Okashe K, Saxena A, El-Bedowey MM, Shoker AS (1997) Immunoglobulin G subclasses and susceptibility to allosensitization in humans. *Clin Nephrol* 48: 165-172.
- 79 Fidler ME, Gloor JM, Lager DJ (2003) Histologic findings of antibody mediated rejection in ABO blood group incompatible living donor kidney transplantation. *Am J Transplant* 3: 1-7.
- 80 Crespo M, Pascual M, Tolkoff-Rubin N (2001) Acute humoral rejection in renal allograft recipients: I. Incidence, serology and clinical characteristics. *Transplantation* 71: 652-658.
- 81 Shimoda M, Marubashi S, Dnj K (2009) ABO-incompatible adult liver transplantation when the anti-ABO antibody titer is high. *Hepatogastroenterology* 56: 1174-1177.
- 82 Ponticelli C, Glassock RJ (2010) Posttransplant recurrence of primary glomerulonephritis. *Clin J Am Soc Nephrol* 5: 2363-2372.
- 83 Silvestre C, Furian L, Marson P (2014) Desensitization with plasmapheresis and anti-Cd20 for ABO incompatible kidney transplantation from living donor: Experience of a single center in Italy. *Transplant Proc* 46: 2209-2213.
- 84 Tanabe K, Inui M (2013) Desensitization for prevention of chronic antibody-mediated rejection after kidney transplantation. *Clin Transplant* 27(26): 2-8.
- 85 Higgins R, Lowe D, Hathaway M (2010) Double filtration plasmapheresis in antibody-incompatible kidney transplantation. *Ther Apher Dial* 14: 392-399.
- 86 Stussi G, Halter J, Buchelli E (2009) Prevention of pure red cell aplasia after major or bidirectional ABO blood group incompatible hematopoietic stem cell transplantation by pretransplant reduction of host anti-donor isoagglutinins. *Haematologica* 94: 239-248.
- 87 Ragimov AA, Solovyeva IN, Dashkova NG (2012) Efferent hemocorrection in preparing recipients for transplantation-related kidney, incompatible ABO. Proc XX Confer Moscow Society of Hemapheresis, Dubna pp: 109-110.
- 88 Tobian AA, Shirey RS, Montgomery RA (2008) The critical role of plasmapheresis in ABO-incompatible renal transplantation. *Transfusion* 48: 2453-2460.
- 89 Hickstein H, Loball S, Lehmann R (2014) ABO incompatible kidney transplantation using unspecific immunoadsorption. *Transfus Apher* 50: 263-266.
- 90 De Weerd AE, Van Agteren M, Leebeek FW (2015) ABO-incompatible kidney transplant recipients have higher bleeding risk after antigen-specific immunoadsorption. *Transpl Int* 28: 25-33.
- 91 Riella LV, Safa K, Yagan J (2014) Long-term outcomes of kidney transplantation across a positive complement-dependent cytotoxicity crossmatch. *Transplantation* 97: 1247-1252.
- 92 Snyder LD, Gray AL, Reynolds JM (2014) Antibody desensitization therapy in highly sensitized lung transplant candidates. *Am J Transplant* 14: 849-856.
- 93 Sofue T, Hayashida Y, Hara T (2014) Plasmapheresis in a patient with antiphospholipid syndrome before living-donor kidney transplantation: A case report. *BMC Nephrol* 15: 167.
- 94 Solovyova IN, Ragimov AA (2007) Plasmapheresis in reconstructive surgery and transplantation. Proc confer Actual issues of extracorporeal treatment, Moscow (Rus) pp: 19-20.
- 95 Shumakov VI, Morozov BN (1997) The first experience of application of plasmapheresis in the intraoperative period when re-transplant renal transplant. Proc V Confer Moscow society of hemapheresis, Moscow p: 63.
- 96 Akalin E, Dinavahi R, Friedlander R (2008) Addition of plasmapheresis decreases the incidence of acute antibody-mediated rejection in sensitized patients with strong donor-specific antibodies. *Clin J Am Soc Nephrol* 3: 1160-1167.
- 97 Morozov BN, Kazakov EN, Ostroumov EN (2001) Experience of application software plasmapheresis in a patient after the orthotopic heart transplantation. Proc IX Confer Moscow Society of Hemapheresis Moscow pp: 33-34.
- 98 Kaabak MM, Goryainov VA, Ragimov AA (2012) Ten years experience in the application of early plasmapheresis after kidney transplantation. Proc XX Confer Moscow Society of Hemapheresis, Dubna pp: 68-69.
- 99 Vatazin AV, Sinyutin AA, Zulkarnaev AB (2014) The impact of plasmapheresis on the intraorgan blood flow of the transplanted kidney in the early postoperative period. *Urology (Rus)* 1: 16-19.
- 100 Cochat P, Kassir A, Colon S (1993) Recurrent nephrotic syndrome after transplantation: Early treatment with plasmapheresis and cyclophosphamide. *Pediat Nephrol* 7: 50-54.
- 101 Solovyova IN, Ragimov AA (2001) Plasmapheresis in reperfusion syndrome. Proc IX Confer Moscow Society of Hemapheresis, Moscow p: 43.
- 102 Sinyutin AA, Vatazin AV, Kantaria RO (2011) The use of plasmapheresis in ischemic reperfusion injury of the renal graft. Proc XIX confer Moscow Society of Hemapheresis, Moscow pp: 89-90.
- 103 Mandal AK, King KE, Humphreys SL (2000) Plasmapheresis: An effective therapy for primary allograft nonfunction after liver transplantation. *Transplantation* 70: 216-220.
- 104 Lick SD, Vaidya S, Kollar AC (2008) Peri-operative alemtuzumab (Campath-1H) and plasmapheresis for high-PRA positive lymphocyte crossmatch heart transplant: A strategy to shorten left ventricular assist device support. *J Heart Lung Transplant* 27: 1036-1039.
- 105 Moseshvili EG, Biryukova LS, Petrov VI, Kalinin NN (2007) Features of allogeneic renal graft engraftment with full restoration of its function in combination therapy with polyclonal and monoclonal antibodies in combination with plasmapheresis. Proc confer "Topical issues of extracorporeal therapy", Moscow pp: 48-49.
- 106 Tondolo V, Manunza R, Pellegrino RA, Zamboni F (2015) Pancreas transplantation: Small-center experience in type 1 diabetes mellitus in a high-incidence region. *Transplant Proc* 47: 2169-2172.
- 107 Woeste G, Wullstein C, Pridöhl P (2003) Incidence of minor and major amputations after pancreas/kidney transplantation. *Transpl Int* 16: 128-132.
- 108 Ruiz P, Carreno M, Weppler D (2010) Immediate antibody-mediated (hyperacute) rejection in small-bowel transplantation and relationship to cross-match status and donor-specific C4d-binding antibodies: Case report. *Transplant Proc* 42: 95-99.
- 109 Kozaki K, Egawa H, Ueda M (2006) The role of apheresis therapy for ABO incompatible living donor liver transplantation: The Kyoto University experience. *Ther Apher Dial* 10: 441-448.
- 110 Matsuno N, Iwamoto H, Nakamura Y (2008) ABO-incompatible

- adult living donor liver transplantation for hepatocellular carcinoma. *Transplant Proc* 40: 2497-2500.
- 111 Ashizawa T, Matsuno N, Yokoyama T (2006) The role of plasmapheresis therapy for perioperative management in ABO-incompatible adult living donor liver transplantation. *Transplant Proc* 38: 3629-3632.
 - 112 Hong G, Yi NJ, Suh SW (2014) Preoperative selective desensitization of liver transplant recipients considering the degree of T-lymphocyte cross-match titer, model for end-stage liver disease score, and graft liver volume. *J Korean Med Sci* 29: 640-647.
 - 113 Raut V, Mori A, Kaido T (2012) Splenectomy does not offer immunological benefits in ABO-incompatible liver transplantation with a preoperative rituximab. *Transplantation* 93: 99-105.
 - 114 Kheradmand F, Shan M, Xu C, Corry DB (2012) Autoimmunity in chronic obstructive pulmonary disease: Clinical and experimental evidence. *Expert Rev Clin Immunol* 8: 285-292.
 - 115 Lee JY, Kim SB, Chang JW (2010) Comparison of the molecular adsorbent recirculating system and plasmapheresis for patients with graft dysfunction after liver transplantation. *Transplant Proc* 42: 2625-2630.
 - 116 Kamar N, Lavayssi er L, Muscari F (2009) Early plasmapheresis and rituximab for acute humoral rejection after ABO-compatible liver transplantation. *World J Gastroenterol* 15: 3426-3430.
 - 117 Yoon SY, Hwang S, Ahn CS (2013) Clinical outcome of idiopathic hepatic parenchymal infarct following living donor liver transplantation. *Transplant Proc* 45: 3072-3075.
 - 118 Murakami K, Kawagishi N, Ishida K (2014) Fibrosing cholestatic hepatitis developing within one month after living donor liver transplantation for chronic hepatitis C-related cirrhosis: A case report. *Transplant Proc* 46: 995-998.
 - 119 Korach JM, Guillevin L, Petitpas D (2000) Apheresis registry in France: indications, techniques, and complications. *Ther Apher* 4: 207-210.
 - 120 Keren A, Hayes HM, O'Driscoll G (2006) Late humoral rejection in a cardiac transplant recipient treated with the anti-CD20 monoclonal antibody rituximab. *Transplant Proc* 38: 1520-1522.
 - 121 Rummel S, Barz D (2012) Plasma exchange and immunoadsorption of patients with thoracic organ transplantation. *Transfus Med Hemother* 39: 234-240.
 - 122 Statinska J, Honsova E, Burgelova M (2009) Plasmapheresis and intravenous immunoglobulin in early antibody-mediated rejection of the renal allograft: A single-center experience. *Ther Apher Dial* 13: 108-112.
 - 123 Dzemeshevich S, Ragimov A, Mikhaylov Y (1998) Plasmapheresis in the treatment of posttransplant cardiomyopathy. *Artif Organs* 22: 197-202.
 - 124 Verheyen J, Vermeulin T, Van-Doorn J (2011) Treating humoral rejection after cardiac transplantation. *Acta Cardiol* 66: 263-266.
 - 125 Grauhan O, Knosalla C, Ewert R (2001) Plasmapheresis and cyclophosphamide in the treatment of humoral rejection after heart transplantation. *J Heart Lung Transplant* 20: 316-320
 - 126 Singh VK, Lin SX, Yang VC (1998) Serological association of measles virus and human herpesvirus-6 with brain autoantibodies in autism. *Clin Immunol Immunopathol* 89: 105-108.
 - 127 Pollock-Barsim SM, Den-Hollander N, Ngan BY (2007) Pediatric heart transplantation in human leukocyte antigen sensitized patients: Evolving management and assessment of intermediate-term outcomes in a high-risk population. *Circulation* 116(11): 172-178.
 - 128 Saheb S, Euraud D, Ouldammam S (2014) Strongly donor-specific antibodies and absence of acute rejection after combined heart and liver transplantation: Case report. *J Clin Apher* 29: 62-63.
 - 129 Velez M, Johnson MR (2009) Management of allosensitized cardiac transplant candidates. *Transplant Rev (Orlando)* 23: 235-247.
 - 130 Holt DB, Lublin DM, Phelan DL (2007) Mortality and morbidity in pre-sensitized pediatric heart transplant recipients with positive donor crossmatch utilizing peri-operative plasmapheresis and cytolytic therapy. *J Heart Lung Transplant* 26: 876-882.
 - 131 Chih S, Tinckam KJ, Ross HJ (2013) A survey of current practice for antibody-mediated rejection in heart transplantation. *Am J Transplant* 13 : 1069-1074.
 - 132 Roman PE, Devore AD, Welsby IJ (2013) Techniques and applications of perioperative therapeutic plasma exchange. *Curr Opin Anaesthesiol* 27: 57-64.
 - 133 Otsuka Y, Takeda A, Horike K (2014) Early recurrence of active IgA nephropathy after kidney transplantation. *Nephrology (Carlton)* 19 (3): 45-48.
 - 134 Potestio C, Jordan D, Kachulis B (2017) Acute postoperative management after transplantation. *Best Pract Res Ckin Anaesthesiol* 31: 273-284.
 - 135 Voinov VA (2016) Therapeutic apheresis. *Constan a: Celebris* 2: 1.
 - 136 Serrick C, Giaid A, Reis A, Shennib H (1997) Prolonged ischemia is associated with more pronounced rejection in lung allograft. *Ann Thor Surg* 63: 202-208.
 - 137 Bittner HB, Dunitz J, Hertz M (2001) Hyperacute rejection in single lung transplantation: Case report of successful management by means of plasmapheresis and antithymocyte globulin treatment. *Transplantation* 71: 649-651.
 - 138 Bok JS, Jun JH, Lee HJ (2014) A successful bilateral lung transplantation in a patient with high panel reactive antibody and positive cross matching. *Korean J Thorac Cardiovasc Surg* 47: 420-422.
 - 139 Madurka I, Elek J, Sch onauer N (2017) Urgent lung transplantation in severe acute respiratory failure based on rapidly progressive interstitial lung disease: A case report. *Transplant Proc* 49: 1544-1448.
 - 140 Martinu T, Chen DF, Palmer SM (2009) Acute rejection and humoral sensitization in lung transplant recipients. *Proc Am Thorac Soc* 6: 54-65.
 - 141 Aguilar PR, Michelson AP, Isakov W (2016) Obliterative bronchiolitis. *Transplantation* 100: 272-283.
 - 142 Gr onnings eter IS, Tsykunova G, Lilleeng K (2017) Bronchiolitis obliterans syndrome in adults after allogeneic stem cell transplantation—pathophysiology, diagnostics and treatment. *Expert Rev Clin Immunol* 13: 553-569.
 - 143 Towe C, Chester-Ogborn A, Ferkol T, Sweet S, Huddleston C, et al. (2015) Bronchiolitis obliterans syndrome is not specific for bronchiolitis obliterans in pediatric lung transplant. *J Heart Lung Transplant* 34: 516-521.
 - 144 Fakhro M, Broberg E, Algotsson L (2017) Double lung, unlike single lung transplantation might provide a protective effect on mortality and bronchiolitis obliterans syndrome. *J Cardiothorac Surg* 12: 100.

- 145 Verleden SE, Sacreas A, Vos R (2016) Advances in understanding bronchiolitis obliterans after lung transplantation. *Chest* 150: 219-225.
- 146 Vacha M, Chery G, Hulbert A (2017) Antibody depletion strategy for the treatment of suspected antibody-mediated rejection in lung transplant recipients: does it work?. *Clin Transplant* p: 31.
- 147 Lus F, Verboom M, Sommer W, Poyanmehr R, Knoefel AK, et al. (2018) Preemptive treatment of early donor-specific antibodies with IgA- and IgM-enriched intravenous human immunoglobulins in lung transplantation. *Am J Transplant* 18: 2295-2304.
- 148 Jackups R Jr, Canter C, Sweet SC (2013) Measurement of donor-specific HLA antibodies following plasma exchange therapy predicts clinical outcome in pediatric heart and lung transplant recipients with antibody-mediated rejection. *J Clin Apher* 28: 301-308.
- 149 Boctor FN (2006) Tacrolimus (FK506) associated thrombotic thrombocytopenic purpura/hemolytic uremic syndrome in lung transplant salvage with a plasmapheresis and cyclosporine. *Egypt J Immunol* 13: 95-99.
- 150 Yung GL, Craig V (2015) Lung transplantation and extracorporeal photopheresis: The answer to bronchiolitis obliterans?. *Transfus Apher Sci* 52: 162-166.
- 151 Hadjiliadis D, Chaparro C, Reinsmoen NL (2005) Pre-transplant panel reactive antibody in lung transplant recipients is associated with significantly worse post-transplant survival in a multicenter study. *J Heart Lung Transplant* 24(7): 249-254.
- 152 Strüber M, Warnecke G, Hafer C (2008) International ABO-incompatible lung transplantation. *Am J Transplant* 8: 2476-2478.
- 153 Turza KC, Shafique M, Lobo PI (2014) Infectious complications in living-donor kidney transplant recipients undergoing multi-modal desensitization. *Surg Infect (Larchmt)* 15: 182-186.
- 154 Liu JM, Hockenberry M (2011) Review of chronic graft-vs.-host disease in children after allogeneic stem cell transplantation: Nursing perspective. *J Pediatr Oncol Nurs* 28: 6-15.
- 155 Hymes SR, Alousi AM, Cowen EW (2012) Graft-versus-host disease: Part I, Pathogenesis and clinical manifestations of graft-vs.-host disease. *J Am Acad Dermatol* 66: 1-18.
- 156 Beres AJ, Drobowski WR (2013) The role of regulatory T cells in the biology of graft vs. host disease. *Front Immunol* 4: 163.
- 157 Zeiser R (2015) Activation of innate immunity in graft-versus-host disease: Implications for novel targets?. *Oncol Res Treat* 38: 239-243.
- 158 Kavand S, Lehman JS, Hashmi S (2017) Cutaneous manifestations of graft-versus-host disease: Role of the dermatologist. *Int J Dermatol* 56: 131-140.
- 159 Teshima T, Reddy P, Zeiser R (2016) Acute graft-vs.-host disease: Novel biological insights. *Biol Blood Marrow Transplant* 22: 11-16.
- 160 Strong-Rodrigues K, Oliveira-Ribeiro C, Fiuza-Gomes SDA, Knobler R (2018) Cutaneous graft-vs.-host disease: Diagnosis and treatment. *Am J Clin Dermatol* 19: 33-50.
- 161 Feito-Rodríguez M, De Lucas-Laguna R, Gómez-Fernández C (2013) Cutaneous graft versus host disease in pediatric multivisceral transplantation. *Pediatr Dermatol* 30: 335-341.
- 162 Ferrara JLM, Levine JE, Reddy P, Holler E (2009) Graft-vs.-host disease. *Lancet* 373: 1550-1561.
- 163 Carreras E, Diaz-Beyá M, Rosiñol L (2011) The incidence of veno-occlusive disease following allogeneic hematopoietic stem cell transplantation has diminished and the outcome improved over the last decade. *Biol Blood Marrow Transplant* 17: 1713-1720.
- 164 Yadav H, Nolan ME, Bohman JK (2016) Epidemiology of acute respiratory distress syndrome following hematopoietic stem cell transplantation. *Crit Care Med* 44: 1082-1090.
- 165 Rieger CT, Rieger H, Kolb HJ (2009) Infectious complications after allogeneic stem cell transplantation: Incidence in matched-related and matched-unrelated transplant settings. *Transpl Infect Dis* 11: 220-226.
- 166 Rai V, Dietz NE, Agrawal DK (2016) Immunological basis for treatment of graft vs. host disease after liver transplant. *Expert Rev Clin Immunol* 12: 583-593.
- 167 Voinov VA, Pugachev AA, Karchevskii KS (2009) The use of the device "Hemophenix" at children. Proc confer "Topical issues of hemapheresis and hemocorrection", Moscow p: 19.
- 168 Rossi AP, Bone BA, Edwards AR (2014) Graft-vs.-host disease after simultaneous pancreas-kidney transplantation: A case report and review of the literature. *Am J Transplant* 14: 2651-2656.
- 169 Wolff D, Bertz H, Greinix H (2011) Problems of graft-versus-host-disease. *Dtsch Arztebl Int* 108: 732-740.
- 170 Choi CM, Schmaier AH, Snell MR, Lasarus HM (2009) Thrombotic microangiopathy in hematopoietic stem-cell transplantation. *Drugs* 69: 183-198.
- 171 Willems E, Baron F, Seidel L (2010) Comparison of thrombotic microangiopathy after allogeneic hematopoietic cell transplantation with high-dose or nonmyeloablative conditioning. *Bone Marrow Transplant* 45: 689-693.
- 172 George JN, Li X, McMinn JR (2004) Thrombotic thrombocytopenic purpura-hemolytic uremic syndrome following allogeneic HPC transplantation: A diagnostic dilemma. *Transfusion* 44: 294-304.
- 173 Laskin BL, Goebel J, Davies SM, Jodele S (2011) Small vessels, big trouble in the kidney and beyond: Hematopoietic stem cell transplantation-associated thrombotic microangiopathy. *Blood* 118: 1452-1462.
- 174 Christidou F, Athanasiadou A, Kalogiannidis P (2003) Therapeutic plasma exchange in patients with grade 2-3 hematopoietic stem cell transplantation-associated thrombotic thrombocytopenic purpura: A ten years later experience. *Ther Apher Dial* 7: 259-262.
- 175 Kennedy GA, Kearey N, Bleakley S (2012) Transplantation-associated thrombotic microangiopathy: Effect of concomitant GVHD on efficacy of therapeutic plasma exchange. *Bone Marrow Transplant* 45: 699-704.
- 176 Voinov VA (2013) Respiratory distress syndrome. St. Petersburg: RIZ PSPbGMU.
- 177 Morris SH, Haight AE, Kamat P, Fortenberry JD (2010) Successful use of extra-corporeal life support in hematopoietic stem cell transplant patient with diffuse alveolar hemorrhage. *Pediatr Crit Care Med* 11: 4-7.
- 178 Pierelli L, Bosi A, Olivieri A (2018) Best practice for extracorporeal photopheresis in acute and chronic graft-vs.-host disease by Societa' Italiana di Emaferesi and Manipolazione Cellulare and Gruppo Italiano Trapianto Midollo Osseo: a national survey to ascertain its degree of application in Italian transplant centers. *Transfusion* 58: 217-222.
- 179 Schneiderman J (2017) Extracorporeal photopheresis: Cellular

- therapy for the treatment of acute and chronic graft-vs.-host disease. *Hematology Am Soc Hematol Educ Program* 1: 639-644.
- 180 Franklin C, Cesko E, Hillen U (2015) Modulation and apoptosis of neutrophil granulocytes by extracorporeal photopheresis in the treatment of chronic graft-versus-host disease. *PLoS One* 10: 0134518.
- 181 Patel J, Klapper E, Shafi H, Kobashigawa JA (2015) Extracorporeal photopheresis in heart transplant rejection. *Transfus Apher Sci* 52: 167-170.
- 182 Mohammadi AM, Norooznexhad AH, Seghatchian J (2018) Photopheresis of a less than 10-kg child with acute graft versus host disease accompanied with hyperbilirubinemia: A case report. *Transfus Apher Sci* 57: 428-430.