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European Journal of Experimental Biology, 2012, 2 (4):1144-1150



The impact of elevated CO₂ on growth and competitiveness of C₃ and C₄ crops and weeds

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ABSTRACT

This experiment was conducted to evaluate the effects of normal CO₂ level (350ppm) and elevated concentration (700ppm) on growth and competitive ability of millet and soybean against pigweed and lambsquarters. The plants were planted as mono and multicultural to study inter- and intra-specific competition in the greenhouse. Root and shoot dry weights and chlorophyll value was measured at the end of vegetative growth. The results showed that plant chlorophyll content rose up by increasing CO₂ concentration, especially in C₃ plants when they were intercropped with C₄ plants. CO₂ elevation caused considerably higher root, stem and leaf weight in C₃ plants than in C₄ plants. In intercropping condition, C₄ plants shoot dry weights decreased under elevated CO₂ concentration. It indicated that competitive ability of those plants reduced in these situations. In all investigated plants, root shoot ratio reduced by increasing CO₂ concentration. Generally, PRY comparisons showed that competitive ability of soybean and lambsquarter increased and millet and pigweed decreased under elevated CO₂ concentration. Therefore weed – crop interactions would be highly affected by CO₂ concentration.

Key words: Climate change, competition, weed management, photosynthetic pathways

INTRODUCTION

Weeds are usually known as plants that interfere in growth, yield and production of cropping systems. Weeds, due to their competition with crops for soil and water resources, cause reduction of yield quantity and quality and land value and farmers could not encountered the weed damages [1]. Nowadays, great portion of production expenditures belongs to weed control. For example it is reported that weeds caused 12 percent reduction in crop production and their control costs is 35 billion dollars [2]. Developing countries spend much more amounts [3]. Furthermore, weeds could be a pest and diseases host and these increase their control complexities. Recognition the characteristics which play role in weed competition ability are important in weed management. Environmental factors are important in alteration of weed competition ability. From the most recent attractive environmental topics is climatic change. CO₂ concentration have been risen up from 285 in 1950 to 370 (30 percent increase) [4]. CO₂ changes also caused the temperature to change. Therefore it is important to understand the effects of elevated CO₂ on plant growth and metabolism. It's reported that elevated CO₂ induced growth and development of more than 100 plant species [5,6,7].

One of the important agricultural aspects which influenced from CO₂ elevation is weed-crop competition [8]. The quality of crop and weed competition is shown to be affected by environmental condition and varied by increasing CO₂ concentration [9]. Different responses in C₃ and C₄ plants to crescent CO₂ and temperature might change their competition ability. This could be important because most of the world crops are C₃ and often the noxious weeds are C₄ [10].

It was showed that C₃ plant growth would be induced by CO₂ concentration [11]. However it was showed that there is a great interspecific variation in plants to respond to CO₂. Growth of C₄ plant also could be induced by CO₂ at lower rates [12,13,14]. Porter, 1993 reported that by duplicating CO₂ concentration, the average growth of 156 species would be increased 37%. C₃ plants growth (41%) was higher than C₄ (22%). CAM plants showed the lower responses. Ziska [13] examined the competitive ability of sorghum against *Xanthium strumarium* under normal and higher CO₂ concentrations and concluded that by increasing CO₂ concentration, the competitive ability of sorghum decreased. It was observed that by increasing CO₂ concentration the photosynthesis, growth and competitive ability of C₃ plants would increase. Therefore time and dose of herbicides application like glyphosate would be changed for C₃ plants and had to be applied earlier or in higher concentrations. Such changes had not been observed in C₄s [15]. Some researchers demonstrated that C₄ plants responded better to elevated CO₂. For example, Owensby *et al.* [16] observed the higher response to CO₂ in C₄ wheatgrass than C₃ plants. This variation in plant response can be related to different temperate, soil, water and nutrient ability [17].

The primary and transient response of plants to increasing ambient CO₂ is to increase photosynthesis rate and decrease in transpiration rates. Increasing CO₂ fixation is due to decrease in photorespiration; however decreased transpiration is related to stomata closure [18]. Nevertheless the permanent effects of CO₂ on growth and physiology of plants has been little detected. For example the advantageous of photosynthesis increase might be ruined by adverse effects of feedback [19].

With respect to increasing atmospheric CO₂ concentration and the necessity of understanding the interactions of crops and weeds in these circumstances, in order to improve managing methods, this experiment was conducted to examine the twin and separated responses of C₃ and C₄ crops and weeds to elevated CO₂ concentrations.

MATERIALS AND METHODS

This experiment was conducted in Lamerd region (52°54'19"N , 28°27'31" E , 500m above sea level, 250mm mean annual participation and 4000 mm annual evaporation), Fars province, Iran in 2010. The experiment was carried out in the greenhouse. Soybean and millet as C₃ and C₄ crops respectively and lambsquarter and pigweed as respective C₃ and C₄ weeds were selected. The experimental design was factorial based on completely randomized design with 20 treatments and four replications. The first factor included of two levels of CO₂ concentrations, 350ppm as normal and 700ppm as elevated concentration. The second factor consisted of monoculture and intercropping (50:50 ratio) of illustrated crops (millet, soybean, lambsquarter, pigweed, millet-soybean, millet-pigweed, millet-lambsquarter, soybean-pigweed and soybean-lambsquarter, pigweed-lambsquarter).

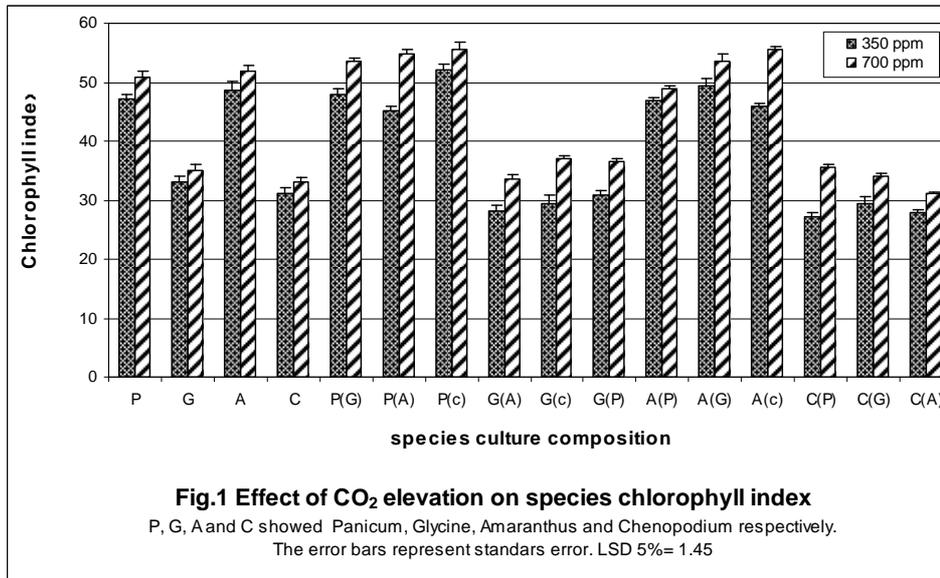
In order to elevating CO₂ concentration, the CO₂ container capsule was used from the 2-3 leaf stage. For measuring and controlling CO₂ concentration, the portable CO₂ meter (Model AZ77535, Thailand) was used. The greenhouse temperatures remained constant at 35°C day and 25°C night. The plants were cultivated in plastic pots (45cm diameter and 55cm height), which were filled by loamy soil and manure to avoid soil crusting. 50 % density of monoculture of each plant was considered as a mixed culture density. Appropriate phosphate and ammonium were applied according to soil chemical analysis.

The measured parameters were chlorophyll index during the growth season, root, stem and leaf dry weight of each plant at the beginning of flowering. Chlorophyll was measured from the three random points of ultimate fully expanded leaf by SPAD method using chlorophyll measuring probe (Model CI 200, Optiscience, USA). At the beginning of flowering the plants were cut from the soil surface and the stems and leaves were separated and oven dried at 70°C for three days. The root also were washed up and after cleaning, were put in oven at 60°C (50°C for millet) for 3 days and then weighted. Root/shoot ratio were measured by dividing root dry weight to shoot dry weight and plant relative yield (PRY) were measured as, shoot dry weight in monoculture to shoot dry weight in mixed culture. The data were analyzed using GENSAT 11 software. Means were compared by LSD examination. The graphs were drawn by Excel.

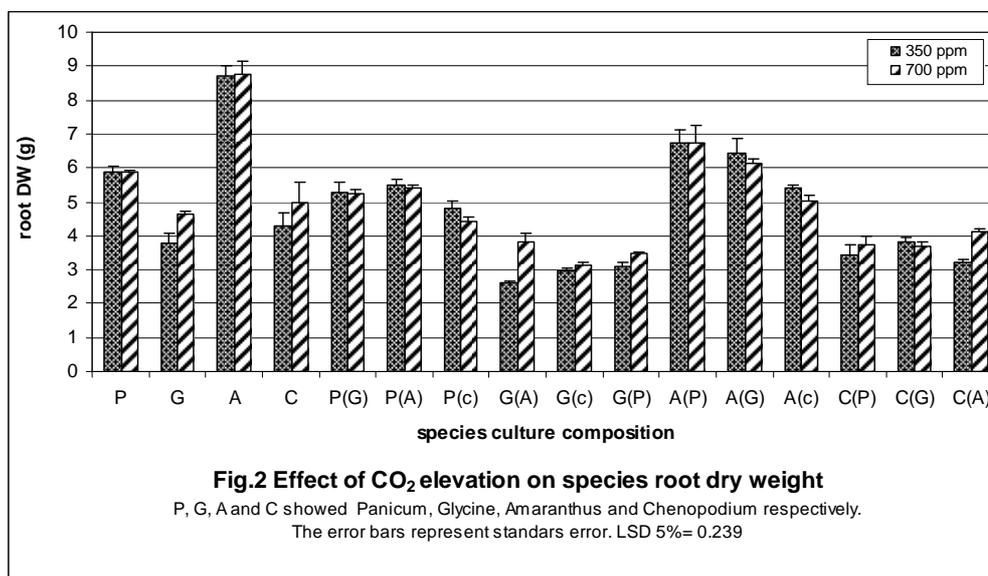
RESULTS AND DISCUSSION

Chlorophyll index- Chlorophyll indices of all examined species were increased by increasing CO₂ concentration but with different rates (Fig 1). The rate of chlorophyll increase in monoculture for millet, soybean, pigweed and lambsquarter were 7.8, 6.6, 6.5 and 7 percentages, respectively. It seems that C₃ plants responded better to CO₂ especially in mixed culture and in competition with C₄ plants. The highest chlorophyll enhancement (30.7%) was seen in lambsquarter when sown with millet (C(P)) followed by 25.4% increase which was seen in mixed cropping of lambsquarter and soybean (G(C)). The lowest chlorophyll increase (6.7%) was observed in competition of millet and pigweed (C₄). Higher chlorophyll increase in mix cropping may be related to higher competitive ability of C₃

plants under CO₂ elevation. There were contradictory cited results about the effect of CO₂ on chlorophyll. CO₂ enrichment, caused chlorophyll to increase in cotton [20] and clover [21]. Heagle *et al.* [22] also reported the 3% increase in wheat chlorophyll content due to CO₂ enrichment. However, decrease in chlorophyll content in *Brassica oleraceae* and lambsquarter was observed by Sage *et al.* [23] in elevated CO₂ condition. Different chlorophyll responses were might be related to different experimental conditions as well as soil nitrogen content variation. According to Hinmann *et al.* [24] the whole plant response to CO₂ would be altered due to biochemical limitations like lower rubisco activity, Ultrastructural limitations like chloroplast degradation and changes in canopy status like leaf area fluctuations.

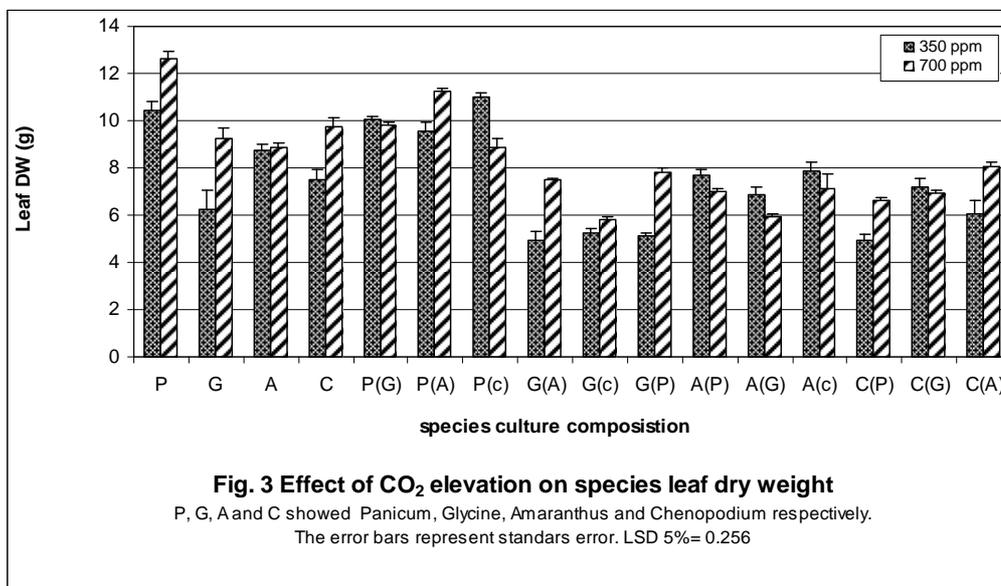


Root dry weight- Effects of CO₂ increasing on root dry weight are shown in Fig. 2. The species showed different responses to elevated CO₂. In pure culture, root dry weight of soybean and lambsquarter increased but any obvious root dry weight increase was seen in millet and pigweed. This led us to conclude that C₃ plants respond better to CO₂ elevation then C₄. In competitive circumstances (mixed culture) the millet root weight remained constant when planted in mixture with soybean and pigweed, but decreased when planted with lambsquarter (Fig. 2). It shows that competitive ability would differ among C₃ plants and also under elevated CO₂ condition competitive ability of millet (C₄ plant) would decrease. The similar results were obtained from pigweed as in the vicinity of soybean and lambsquarter, its root dry weight decreased but when adjacent with pigweed its root dry weight remained constant. It could be concluded that millet and pigweed could compete better under normal CO₂ concentration. Golvi [25] reported that in normal situation pigweed competitive ability is higher than soybean but in higher CO₂ concentration soybean was stronger competitor.



It was seen that soybean root weight increased in the mix culture with millet and pigweed, but remain constant in the adjacent with lambsquarter. Also lambsquarter's root weigh increased in the vicinity of millet and pigweed. Pritchard *et al.* [26] demonstrated that root growth of C₄ plants like sorghum would decrease under elevated CO₂ condition. Derner *et al.* [14] perceived that root biomass response in cotton and sorghum depended to density and type of species competition. As in low density, root biomass of sorghum respond better to elevated CO₂, but in dense planting the cotton root weight increased (126% increasing) more than sorghum (13% increasing). Dipperry *et al.* [27] observed that velvetleaf (*Abutilon theophrasti*) root weight respond better to CO₂ enhancement than pigweed. By increasing CO₂ concentration from 150 ppm to 700 ppm, root dry weight of velvetleaf increased from 0.38 to 11.7g. Such biomass and growth changes could cause plants to alter competition ability. Bazaz *et al.* [28] stated that pigweed competitive ability is high because of higher root growth and ability of absorbing nitrogen from subsoil. But such competitive ability decreased under increasing CO₂ concentration.

Leaf dry weight- Under elevated CO₂ condition, leaf weight of millet, lambsquarter and soybean increased when they were mono cultured (Fig. 3). The highest leaf weight increase was seen in soybean (47.9%) followed by lambsquarter (29.5%) and millet (21.2%). The lowest leaf weight increase was observed in pigweed (1.8%). Therefore the C₃ plants respond better to CO₂ increase. Ziska [13] observed that in elevated CO₂ concentrations, leaf weight and area increased 50 and 35 %, respectively in cocklebur (*Xanthium pensylvanicum*) but the increase of leaf weight and area was 0.5 and 2.4% in sorghum. The reaction rate was decreasing after a while. The same trend was seen in cotton and sorghum in response to elevated CO₂ [14]. Ghanoum *et al.* [29] believed that young leaves of C₄ plants react as similar as C₃ plants to elevated CO₂.

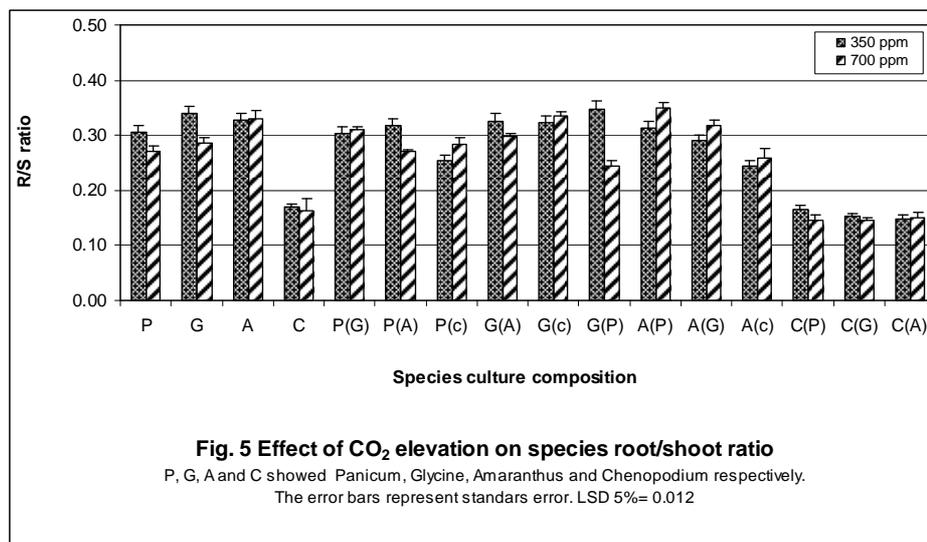
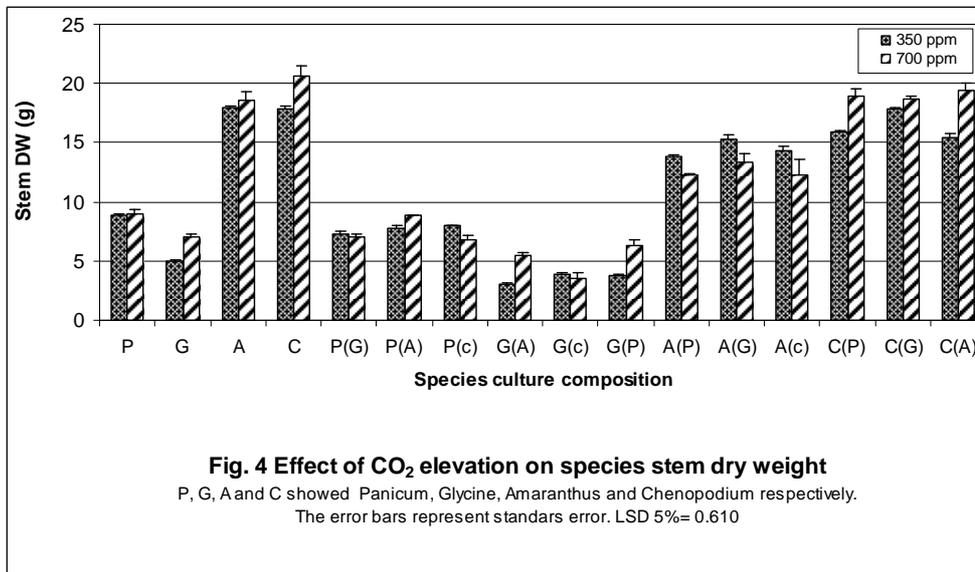


The different responses were seen in multiculture condition among the plants. In elevated CO₂ concentration, leaf weight of millet increased in the vicinity of pigweed and decreased in the vicinity of lambsquarter and soybean (Fig. 3). In pigweed, also the leaf weight decreased in the vicinity of all tested species especially lambsquarter and soybean. Contrary to C₄ species, the C₃ leaf weights increased in the vicinity of C₄ species and remain constant in the vicinity of each other. So that by increasing CO₂ concentration, competitive ability of C₃ plants increased and this led to leaf higher growth.

Wand *et al.* [30] stated that by increasing CO₂ concentration, vegetative growth of both C₃ and C₄ plants increased. But in C₃ plants the response was much more visible. Also the species reaction was different under inter- and intra-specific competition situations. Ziska [13] observed that under interspecific competition (monoculture) the velvetleaf's leaf weight increased only 16% in response to elevated CO₂, but under intraspecific competition situation (multicultural) the leaf weight increased 42%. Ishizaki *et al.* [31] also reported that CO₂ concentration increase caused increasing in root shoot ratio and leaf weight ratio.

Stem dry weight- Fig. 4 shows the effect of CO₂ elevation on stem dry weight. The stem weights of millet, soybean, pigweed and lambsquarter increased 1.9, 42.2, 3.4 and 15.6%, respectively by increasing CO₂ in monoculture condition. In multicultural condition, stem weight of millet decreased (13%) in the adjacency of lambsquarter, increased (15%) in the adjacency of pigweed and remain constant in the adjacency of soybean. This indicates that competitive ability of millet increased against pigweed and decreased against lambsquarter. Soybean stem weight increased 76 and 69% in the vicinity of millet and pigweed, respectively but, in the adjacency of lambsquarter the

soybean stem weight decreased slightly. Pigweed stem weight decreased 11, 12 and 15% in the vicinity of millet, soybean and lambsquarter, respectively. This indicated that although both millet and pigweed are C_4 plants but millet could enhance its growth and photosynthesis under elevated CO_2 condition. By increasing CO_2 concentration lambsquarter's stem weight increased 19, 5 and 26% in the vicinity of millet, soybean and pigweed respectively. Mishra *et al.* [32] demonstrated that stem weights of *Brasica* species increased by increasing CO_2 concentration. The highest increase was seen in *B. juncea* (50%) followed by *B. campestris* (45%) and *B. nigra* (only 10%). Similar results were reported by Wand *et al.* [30], Porter [7], Collatz *et al.* [33] and Ghanom *et al.* [29].



Root shoot ratio- The root shoot ratio (R/S) often decreased by increasing CO_2 concentration (Fig. 5). In monoculture in pigweed remained constant and in soybean, millet and lambsquarter decreased 16, 11 and 3 percent respectively. R/S ratio also decreased in mixcropped plants except for millet and pigweed. R/S ratio increased 11% in millet mixcropped with lambsquarter. In mixcropping of pigweed with soybean, millet and lambsquarter, the R/S ratio increased 9, 12 and 6% respectively. It indicated that elevated CO_2 concentration altered translocation of assimilates and decreased the R/S ratio. Besides, R/S ratio increased in C_4 plants the under intraspecific competition (mixcropping). It indicated that by elevating CO_2 concentration, the competitive ability (for above ground resources like light and CO_2) and shoot growth of these species decreased. But in the case of underground resources, C_4 plants could be competitive under elevated CO_2 concentration.

Dippery *et al.* [27] also showed that by increasing CO_2 concentration, R/S ratio increased in pigweed. Polley *et al.* [34] reported that under duplicated CO_2 concentration, root and shoot biomass of *Prosopis glandulosa* increased 37 and 46%, respectively and consequently R/S decreased. R/S ratio and root growth depend exactly to soil fertility and

mineral availability could alter the plant response. For example by increasing CO₂ concentration, root growth of C₄ plants like *Bouteloua gracilis* would decreased in unfertile and dry soils [35]. De Luis *et al.* [36] also demonstrated that drought stress altered plant response to elevated CO₂ as in normal concentration R/S ratio increased 108% under drought stress but in elevated CO₂ concentration, drought stress increased R/S ratio up to 269%.

Plant relative yield (PRY)- By increasing CO₂ concentration, PRY increased in soybean when intercropped with millet and pigweed but decreased when planted with lambsquarter (Table 1). The results also showed that PRY and consequently competitive ability of millet decreased facing with soybean and lambsquarter and increased when intercropped with pigweed under elevated CO₂ concentration. Competitive ability of pigweed decreased when intercropped with others. Competitive ability of lambsquarter increased against pigweed and millet and decreased in counter with soybean. Generally the results showed that CO₂ elevation increase competitive ability of C₃ plants versus C₄s. Among the C₄ species, millet responded better to CO₂ elevation therefore its competitive ability increased against pigweed. Similarly, soybean responded better to CO₂ than lambsquarter. The same results were cited about the competitive ability alteration by CO₂ enhancement. Ziska [13] showed that PRY of sorghum and cocklebur were similar in normal CO₂ concentration but in elevated CO₂ concentration the competitive ability of cocklebur increased against sorghum. Bazaz *et al.* [28] also showed that PRY of C₃ plants increased versus C₄s under elevated CO₂ concentration. Ziska [12] also demonstrated that competitive ability of pigweed and soybean yield loss decreased under elevated CO₂ concentration.

Table 1. Plant relative yield (PRY) for soybean, millet, pigweed and lambsquarter (base on shoot DW) in CO₂ concentration of 350 ppm and 700 ppm

Species	In competition with	PRY at 350 ppm	PRY at 700 ppm
Millet	Soybean	0.902	0.781
Millet	Pigweed	0.901	0.927
Millet	Lambsquarter	0.984	0.723
Soybean	Millet	0.794	0.872
Soybean	Pigweed	0.717	0.793
Soybean	Lambsquarter	0.819	0.573
Pigweed	Millet	0.807	0.723
Pigweed	Soybean	0.831	0.728
Pigweed	Lambsquarter	0.835	0.726
Lambsquarter	Millet	0.819	0.841
Lambsquarter	Soybean	0.987	0.844
Lambsquarter	Pigweed	0.849	0.904

CONCLUSION

Generally the results indicated that by CO₂ elevation vegetative growth of C₃ plants (soybean and lambsquarter) can increase. These inductions were much more visible in shoots than roots. Therefore root shoot ratio would be decreased in these plants. Although C₄ species responded to elevated CO₂ in lower degree, but millet responded higher than pigweed to elevated CO₂. Consequently the competitive ability (referred to PRY) increased in soybean and lambsquarter and decreased in C₄ species. It indicated that in future, the C₃ plants will be more competitive against weeds while competitive ability of C₄ plants will be decrease. With respect to the interactions between weeds and crops, it's necessary to evaluate the effect of CO₂ on seed bank and herbicides efficiency.

REFERENCES

- [1] Oliver, LR, *Weed Technol*, **1988**, 2, 403.
- [2] Pimentel D, McNair S, Janecka J, *Agric Eco Environ*, **2001**, 84, 1.
- [3] Macias FA, Lopez A, Varela RM, Torres A, Molinillo JMG, *Phytochem*, **2004**, 65, 3057.
- [4] Keeling CD, Whorf TP, *A Compendium of Data on Global Changes*, Oak Ridge Natl. Lab., U.S. Dep. of Energy, Oak Ridge, Tenn, **2001**.
- [5] Kimball BA, *Agron J*, **1983**, 75, 779.
- [6] Kimball BA, *J Agric Meteorol*, **1993**, 48, 559.
- [7] Poorter H, *CO₂ and Biosphere*, Kluwer Academic Publishers, Dordrecht, Netherlands, **1993**.
- [8] Prestidge RA, Pottinger RP, *The Impact of Climate Change on Pests, Diseases, Weeds and Beneficial Organisms Present in New Zealand Agricultural and Horticultural Systems*, Hamilton, MAF Technol, **1990**.
- [9] Patterson DT, Flint EP, *Weed Sci*, **1980**, 28, 71.
- [10] Duke SO, *Weed Physiology*, CRC Press, Boca Raton, Florida, **1985**.
- [11] Jongen M, Jones J, *Ann Bot*, **1998**, 82, 1111.
- [12] Ziska LH, *Glob Chan Biol*, **2000**, 6, 899.
- [13] Ziska LH, *Weed Sci*, **2001**, 49, 62.

- [14] Derner J, Johnson H, Kimball BA, Pinter PJ, Polley HW, Tishler CR, Otman RL, Matthias AD, Brooks TJ, *Glob Chan Biol*, **2003**, 9, 452.
- [15] Ziska LH, Bunce JA, Caulfield F, *Aust J Plant Physiol*, **1998**, 25, 801.
- [16] Owensby CE, Ham JM, Knapp AK, Auen LM, *Glob Chan Biol*, **1999**, 5, 497.
- [17] Chen DX, Hunt HW, Morgan JA, *Ecol Model*, **1996**, 87, 11.
- [18] Lambers H, Chapin SF, Pons TL, *Plant Physiological Ecology*, Springer Science, Business Media, LLC, **2008**.
- [19] Woodward FI, *Cur Opin Plant Biol*, **2002**, 5, 207.
- [20] Pinter JP, Idso SB, Hendrix DL, Rokey RR, Rauschkolb RS, Mauney JR, Kimball BA, Hendrey GR, Lewin KF, Nagy J, *Agric Forest Meteo*, **1993**, 70, 163.
- [21] Cave G, Tolley LC, Strain BR, *Physiol Plant*, **1981**, 51, 171.
- [22] Heagle AS, Miller JE, Booker FL, *Crop Sci*, **1998**, 38, 113.
- [23] Sage RF, Sharkey TD, Seeman JR, *Plant Physiol*, **1989**, 89, 590.
- [24] Heinemann AB, Maia HN, Dourado-Neto AD, Ingram KT, Hoogenboom G, *Europ J Agron*, **2005**, 24, 52.
- [25] Golvi M, PhD Thesis, Ferdosi University, Iran, **2003**.
- [26] Pritchard SA, Prior H, Rogers H, Davis MA, Runion GB, Popham TW, *Agric Ecosys Environ*, **2006**, 113, 175.
- [27] Dipperry JK, Tissue DT, Thomas RB, Strain BR, *Oecologia*, **1995**, 101, 13.
- [28] Bazzaz FA, Garbutt K, Reekie EG, Williams WE, *Oecologia*, **1989**, 79, 223.
- [29] Ghannoum O, Von Caemerer S, Ziska LH, Conroy JP, *Plant Cell Environ*, **2000**, 23, 931.
- [30] Wand SJE, Midgley GF, Jones MH, Curtis PS, *Glob Chang Biol*, **1999**, 5, 723.
- [31] Ishizaki SH, Hikosaka K, Hirose T, *Annal Bot*, **2003**, 91, 905.
- [32] Mishra RS, Abdin MZ, Uprety DC, *J Agron Crop Sci*, **1999**, 182, 223.
- [33] Collatz GJ, Berry JA, Clark JS, *Oecologia*, **1998**, 114, 441.
- [34] Polley HW, Tischler CR, Johnson HB, Pennington RE, *Tree Physiol*, **1999**, 19, 359.
- [35] Read JJ, Morgan JA, *Annal Bot*, **1996**, 77, 487.
- [36] De Luis I, Irigoyen JJ, Sanchez-Diaz M, *Physiol Plant*, **1999**, 107, 84.