Prognostic Significance of TIA-1 Cytotoxic T cells versus SIRM in Cutaneous T-Cell Lymphoma

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Abstract

We have previously reported that Synchrotron Infrared Microspectroscopy (SIRM) is a promising tool for studying of epidermotropism in CTCL.

Objective: In this study, we tried to establish new prognostic markers, which could be beneficial in estimating the prognostic index.

Methods: We performed a study on twenty patients with Mycosis Fungoides (MF): Group I (I A) and group II (IB and IIA) with age range from 16-61 years; they were diagnosed clinically in the department of dermatology, Cairo University-Egypt. A 6 mm biopsy was taken from each patient in order to confirm the diagnosis, routine investigations and serum LDH were also done. Immunophenotyping using antibodies CD3, CD4, CD8, CD20, CD30 and TIA-1 were evaluated. Further examination with SIRM was done in NSLS, BNL - USA. Statistical analysis was done by Student t-test and Cluster analysis.

Results: Results revealed that both TIA-1 and LDH were correlated with the progression of the disease significantly. Also, both TIA-1 and LDH confirmed that PUVA therapy was significantly effective to all our patients. Furthermore, SIRM proved that group I and II clustered separately and amide was significantly higher within the lymphocytes of group II than group I. These findings confirm that both TIA-1 and SIRM detected the cellular changes and they showed a unique biochemical signature within the lymphocytes of MF with the disease progress which could be used as prognostic markers for CTCL.

Keywords: Cutaneous T-cell lymphoma; Mycosis fungoides; Prognosis; SIRM; TIA-1

Introduction

Mycosis Fungoides (MF) is one of the most common forms of primary Cutaneous T-Cell Lymphomas (CTCL) [1]. It is very difficult to distinguish between malignant T cells in early-stage MF and benign reactive cells in inflammatory dermatoses [2]. Also, Cytotoxic T-Lymphocytes (CTLs) can recognize and induce target cell cytolysis [3]. Moreover, activated lymphocytes can mediate cell lysis by releasing perforin, granzymes, and TIA-1 (T-cell intracytoplasmic antigen) [4,5]. It should be pointed out that TIA-1 is a cytotoxic protein that is specifically expressed by cytotoxic CD8 positive T cells and natural killer cells. TIA-1 is an intracellular protein of cytotoxic lymphocytes that can induce apoptosis [6]. In 1999, Vermeer and his colleagues revealed that expression of cytotoxic proteins by neoplastic T cells in MF; increases with progression from plaque stage to tumor stage disease [7].

The prognosis of CTCL is usually based on the classification of the International Society for Cutaneous Lymphomas/European Organization for Research and Treatment of Cancer (ISCL/EORTC), which developed the following markers: stage, age, sex, cutaneous histologic features of folliculotropism, CD30 positivity, proliferation index, large-cell transformation, WBC/lymphocyte count, serum lactate dehydrogenase, and identical T-cell clone in blood and skin [8]. However, efficient non-invasive tools for the early diagnosis of lymphoma remain a necessity to be applied clinically. Several studies have reported vibrational spectroscopies and infrared absorption to characterize biological tissues. Infrared spectroscopy can provide insight into protein structure. This technique is sensitive to the backbone amide arrangement of peptide and protein molecules. Synchrotron Infrared Microspectroscopy (SIRM) is a favorable method for a number of reasons: it is adequately sensitive to minimal sample amounts and is not limited by the molecular weight of the sample; yields spectra that are simple to evaluate; does not require protein modifications and is applicable to soluble and discrete samples.
membrane proteins [9]. Psoralen Plus UVA (PUVA) is an established first line of treatment of early MF [10].

**Aim of Work**

The aim of the current study is to evaluate MF progression using TIA-1 and SIRM as prognostic markers. Also, appraise the efficacies of PUVA therapy in MF patients through studying lymphocytic changes before and after treatment.

**Patients and Methods**

This study is a prospective comparative study that was approved by the Research Ethical Committee (REC) of Cairo University. Patient-informed detailed consent forms were completed by all patients before starting this work. It included twenty patients of early stage mycosis fungoides (IA-IB-IIA) who were recruited from the outpatient dermatology clinic, Cairo University. Diagnosis was based on clinical examination and confirmed by histopathological evaluation. Full detailed history and complete clinical examination were done for all patients by two fixed observers. Routine investigations in the form of CBC, ESR, liver and kidney function tests, fasting blood sugar, urine analysis, Serum Lactic Dehydrogenase (LDH), chest x-ray, abdominal ultrasound, lymph node examination, biopsy, and funds examination were done for all patients.

Early stage MF patients with normal liver and kidney functions were included in the study whereas children less than 12 years, pregnant and lactating females, or any patient with any contraindication to phototherapy were excluded from this study. Patients included in this study, their age ranged from 16-61 years were divided into two groups. Group 1 included 10 patients, 5 males (50%) and 5 females (50%). Three patients were skin type III and seven patients were skin type IV. All patients were stage IA. The duration of disease ranged from 4 month to 15 years. The duration of treatment ranged from 5 month to 12 month. The total number of sessions ranged from 67-144 sessions, and the total number of joules ranged from 230-1114 J/cm². Group II included 10 patients, 5 males (50%) and 5 females (50%). Two patients were skin type III and eight patients were skin type IV. The stages were IB and IIA. The duration of disease ranged from 6 month to 13 years. The duration of treatment ranged from 5 months to 14 months. The total number of sessions ranged from 60-170 sessions, and the total number of joules ranged from 216-912 J/cm². All patients in the current study received PUVA in the form of oral administration of 8-methoxypsoralen at a dose of 0.7 mg/kg and after 2 h UVA sessions were received 3 times/week.

**Immunohistochemical Study**

A 6 mm punch biopsies were retrieved from the lesional areas in MF patients. Skin biopsies were prepared and fixed in paraffin blocks. These were cut to obtain 5-6 μ thick sections placed on infrared-reflective MirrIR glass low e-slides (Keveler Technologies, Chesterland, OH) for SIRM studies and then stained by HandE. Immunohistochemical staining for CD3, CD4, CD8, CD20, CD30 and TIA-1. Staining of formalin fixed tissues required pretreatment for retrieving the masked epitopes of the antigens. Immunostaining for TIA-1 required pretreatment by boiling tissue in 10 mM citrate buffer cat number AP-9003 pH 6 for antigen retrieval, while this was done for 10 min and left to cool in room temperature for 20 min. The primary mouse monoclonal antibody (Biocare Medical LLC, USA) for TIA-1 was applied for 60 min. Immunohistochemical staining was completed by the use of ultravision detection system ABC kit (TP-015-HD) (Labvision Vermont, USA), DAB (3,3 di-amino-bezidine tetrahydrochloride) as chromogen was applied for each slide and the slides were incubated for 5 min at room temperature. Counterstaining was done using Mayer’s hematoxylin (TA-060-MH) (Labvision Vermont, USA). Positive immunoreactivity appeared as brown cytoplasmic granular deposits.

**Morphometric Study**

The area of positive immune staining was measured in TIA-1 immunohistochemically stained sections from all subjects involved in the study. Measurements were done in 5 non-overlapping fields at 400x magnification in a field area of 7286.783 μm² for every subject using Leica Qwin 500C image analyzer computer system (England). Images were captured live on the screen from sections under a light microscope (Olympus BX-40, Olympus Optical Co. Ltd., Japan) with affixed video camera (Panasonic Color CCTV camera, Matsushita Communication Industrial Co. Ltd., Japan). The video images were digitized using Leica Qwin which is a Leica’s windows based image analysis tool kit fitted to an IBM compatible personal computer with a color monitor. Brown deposits demonstrate positive immune-reactivity. Binary images were generated by color thresholding for the brown color then the area of these binaries is measured by the Leica Qwin 500 software. This is done for every field of the five fields for each subject to obtain a mean area for each. Mean area percent is the relation between the areas of positivity marked by the binary images to the field area which is 7286.783 μm².

**Synchrotron Infrared Microspectroscopy (SIRM) Study**

Synchrotron Infrared (IR) microspectroscopy was performed at Beamline U2B at the National Synchrotron Light Source, Brookhaven National Laboratory, NY, USA. The synchrotron IR source was used to improve the spatial resolution to 10 μm such that only the lymphocyte nuclei were probed. Fourier Transform Infrared Microspectroscopy (FTIRM) spectra were collected in reflection mode with a 10 μm beam. For each spectrum, 128 scans were accumulated with 8 cm⁻¹ resolution. For each biopsy, spectra from 50-60 nuclei were collected from epidermal and dermal cells, all spectra were averaged, the first derivative was generated, and a 9-point Savitsky-Golay smoothing was applied. Principal Components Analysis (PCA) was performed on the average first derivative spectra using the spectral region from 1900-900 cm⁻¹ (Unscrambler 9.8). In addition, the absorbance intensity of the nucleic acids (1210-1265 cm⁻¹, baseline: 1185-1285 cm⁻¹), RNA (1119-1123 cm⁻¹, baseline: 995-1140
cm$^{-1}$), DNA (1018-1022 cm$^{-1}$, baseline: 995-1140 cm$^{-1}$), Amide I (1600-1700 cm$^{-1}$, baseline: 1480-1715 cm$^{-1}$) and Amide II (1544 cm$^{-1}$). Specifically, were calculated for each spectrum using Omnic software v7.2 (Thermo Scientific, MA-USA).

All SIRM was analyzed by Principal Components Analysis (PCA) is a nonparametric procedure that reduces the number of variables into principle components, or the variables that most describe the variability within a set of data. With spectroscopy data, each wavenumber can be seen as a single variable, and therefore there are many variables that must be considered when comparing multiple spectra.

Statistics

All data were subjected to statistical analysis. Cluster analysis and statistical significance between groups for each parameter was determined using a Student t-test and ANOVA (SPSS).

Results

In the current study, 20 MF patients were included 50% male and 50% females with mean of male age was 52.2 ± 7.254, while in case of females was 47.2 ± 11.212.

Evaluation of TIA-1 level revealed a significant difference between group I and group II (P<0.05). Also, TIA-1 level showed a highly significant difference before and after PUVA treatment (P<0.001). TIA-1 level after PUVA treatment showed a highly significant difference between group I and group II (P<0.05). Furthermore, Serum LDH showed a highly significant correlation with MF progression (P<0.001). Also, LDH revealed a direct proportional correlation with TIA-1 before and after PUVA treatment as shown in Figure 5 and Table 1.

SIRM spectra collected from the nuclei of lymphocytes from each sample revealed that the differences between Amide I/RNA and Amide II/RNA between group I and II were highly significant (P=0.00 and 0.01) respectively as shown in Table 2.
Figure 4: Positive TIA immunoreactivity involving the abundant lymphocytes around the blood vessels. 144 x 108 mm (150 x 150 DPT).

Figure 5: Comparison between TIA and S-LDH in MF patients before and after PUVA treatment.

Table 1: Shows comparison between different MF stages before and after treatment using TIA and S-LDH.

<table>
<thead>
<tr>
<th>Group</th>
<th>Before treatment</th>
<th>After treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>I (IA)</td>
<td>155.245 ± 59.33</td>
<td>90.98±</td>
</tr>
<tr>
<td></td>
<td>± 83.168**</td>
<td>± 68**</td>
</tr>
<tr>
<td>II (IB and IIA)</td>
<td>44.815 ± 12.608</td>
<td>66.15±</td>
</tr>
<tr>
<td></td>
<td>± 12.852**</td>
<td>± 54**</td>
</tr>
</tbody>
</table>

Student t-test: **<0.001=highly significant, *<0.05=significant

Moreover, both groups before and after PUVA treatment showed significant correlation (P=0.02). When PCA was performed on the dermal SIRM spectra of both groups of MF patients, the scores plot of (PC1) vs. (PC2) revealed that all group I spectra clustered together, while group II clustered separately, these findings are consistent with the usual progression of MF. The PC loadings plot shows that the protein region variability indicates a structural difference with a unique biochemical signature which is important in differentiating MF spectra as shown in Figure 6.

Table 2: Shows comparison between different MF stages before and after treatment using SIRM.

<table>
<thead>
<tr>
<th></th>
<th>MF (IA)</th>
<th>MF (IB and IIA)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Before PUVA</td>
<td>After PUVA</td>
<td>Before PUVA</td>
</tr>
<tr>
<td>Amide I/RNA</td>
<td>4.427 2.2607</td>
<td>4.269 1.83*</td>
</tr>
<tr>
<td>Amide II/RNA</td>
<td>3.479 1.8967</td>
<td>3.257 0.95*</td>
</tr>
</tbody>
</table>

Student t-test: **<0.001=highly significant, *<0.05=significant.

Figure 6: Shows comparison between MF groups with cluster analysis.

Discussion

The affinity of T lymphocytes on early stages of MF for the epidermis and papillary dermis; is apparent as there is rarely significant involvement in the underlying reticular dermis. In 1999, Vermeer and his Colleagues showed that Granzyme B (Gr B) and TIA-1 are cytotoxic proteins that are specifically expressed by cytotoxic CD4 or CD8 positive T cells and natural killer cells. They noticed that the expression of cytotoxic proteins by neoplastic T cells in 54 MF patients increases with progression from plaque stage to tumor stage disease. Moreover, the expression of TIA-1 and GrB was found in 35% and 10% patches or plaques, 55% and 20% tumors without blastic transformation, and 75% tumors with blastic transformation (defined as >50% blast cells). So, they concluded that TIA-1 defines a subpopulation of CD8+ T-cells and NK cells that have cytotoxic activity [7]. Furthermore, tumor-stage MF, showed a significant relation between CD8+ CTLs and survival rate; the high proportions of these cells correlated with a better prognosis [11].

The risk of disease progression from early to advanced disease was anticipated by the Cutaneous Lymphoma International Prognostic Index (CLIPi) developed by Benton and his colleagues in 2013; significant adverse prognostic factors at
early diagnosis consisted of five factors were associated with a worse Overall Survival (OS) consisting of age >60 years, male gender, presence of plaques (T1b/T2b), folliculotropic disease and lymph node stage N1/Nx. Moreover, other prognostic variables such as Lactatic Dehydrogenase (LDH), B0a/b, and CD30 status and Total Body-Surface Area Affected (TBSA) and the type of skin lesions were suggested by several authors [12,13].

In the current study, the prognostic value of TIA-1 was evaluated; it was found the expression of TIA-1 was significantly higher in early MF, which was declined significantly by about 71% with progression of the disease. Moreover, PUVA therapy increased the level of TIA-1 in group I and II by 24.1% and 66% respectively. According to ISCL/EORTC classification, serum LDH could be used as a prognostic marker in CTCL. In this study, serum LDH was directly correlated with TIA-1 in both group I and II as shown in (Figure 5 and Table 1), these results confirms that TIA-1 could be used as a prognostic marker for MF.

On the other hand, the current study revealed that lymphocytes in MF lesions were clustered in two clusters, one for group I and the other for group II. Furthermore, there was a significant difference within Amide/RNA ratio within the lymphocytes in both groups of patients according to the disease progression. Both Amide I/RNA and Amide II/RNA in group II were higher than group I significantly. These findings are consistent with the results of Andrus, who found that lymphoma cells in vitro have higher RNA and protein/RNA ratio than normal lymphocytes based on IR spectral differences between proliferating and non-proliferating lymphocytes. In summary, TIA-1 together with SIRM seems to be promising methods for prognosis of MF patients [14-16].

References

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